

Available Online at www.ijpba.info

# International Journal of Pharmaceutical & Biological Archives 2013; 4(2): 265-267

# **REVIEW ARTICLE**

# Disease Management for the Post Larvae of Fresh Water Cultivable Prawn, Macrobrachium malcolmsonii (H. Milne Edwards, 1844)

# P. Soundarapandian\* and D. Varadharajan

Centre of Advanced Study in Marine Biology, Annamalai University, Parangipettai – 608 502, Tamil Nadu, India

#### Received 24 Oct 2012; Revised 08 Mar 2013; Accepted 19 Mar 2013

### ABSTRACT

*Macrobrachium malcolmsonii* is the second largest among fresh water prawns and has a great potential for culture in Tamil Nadu. More yields can be obtained by stocking of good quality, healthy and disease free hatchery produced post larvae. The common diseases occurring in the post larvae include infestation of ciliates, bacterial, fungal and viral diseases as well as MCD (Midcycle Disease), EED (Exuvia Entrapment Disease) and IMN (Idiopathic Muscle Necrosis). All these diseases can be effectively treated and controlled in the hatchery itself for producing healthy post larvae. The disease management largely depends on the use of disinfected water free of pathogens, good quality feeds and general hygiene.

#### Key words: Monsoon river prawn, disease, symptoms, culture management.

### INTRODUCTION

The freshwater prawn, Macrobrachium malcolmsonii is second largest and commercially important fresh water prawn of India, Bangladesh and Pakistan. It is reported to be a possible candidate species for culture and is highly suitable species for both mono and polyculture with carps in Tamil Nadu<sup>[1,2]</sup>. This is competed promisingly with other freshwater prawns in several respects like growth feeding habits. rate, lower cannibalistic tendencies and cultivability in impounded freshwater, euryhaline adaptability in breeding, fairly good fecundity, larval hardiness in withstanding starvation and heavy turbulence <sup>[3]</sup>. It is generally claimed that *Macrobrachium* spp are less susceptible to disease. However, shrimp larval stages are more susceptible to diseases as a result of their high stocking densities in hatcheries with heavy organic loading associated with daily [4] feeding The known diseases of M. malcolmsonii are caused by ciliates, bacteria, fungi, viral and nutritional deficiencies <sup>[5]</sup>, as well as Mid Cycle Disease (MCD), Exuvia Entrapment Disease (EED) and Idiopathic Muscle Necrosis (IMN). All the disease causing organisms are probably present in the rearing water and only affect post larvae when they are stressed due to inadequate feeding, overcrowding and poor water quality. In running a successful hatchery, good management and monitoring measures are to be taken care so to get a healthy and good quality seeds.

### MID CYCLE DISEASE (MCD)

Mid cycle disease is so called because it occurs roughly at the mid cycle of the larval phase. Mortality sets in around the end of last third of the larval stage of larval rearing cycle, say day 10<sup>[6]</sup>. Mortality increases quickly for 3-5 days, then ceases or dramatically decreases. Post larval production may be reduced to 1 or 2 PL/L.

### **Symptoms**

- The larvae swim weakly, often in a corkscrew or spiral manner,
- Loss of appetite and poor feed consumption,
- Moribund individuals are eaten by the healthy larvae,
- The larvae turn bluish gray.

### **Control Measures**

MCD does not respond to any tested antibiotics. Under this situation it is advisable to discard the infected larvae and disinfect the affected tank. The disease condition can be reversed by reducing stocking density and improving pond husbandry and sanitation <sup>[7]</sup>. During this period, larvae should be provided with good quality *Artemia* nauplii <sup>[8]</sup>. If the infection spreads through the hatchery, general disinfection through washing the tanks, filters and equipment with formalin and / or chlorinated water, followed by drying for at least one week is necessary.

# EXUVIA ENTRAPMENT DISEASE (EED)

This disease affects stage XI larvae and early post larvae. It is also known as the metamorphosis moult mortality syndrome. Mortality usually ranges from 20-30% <sup>[7,8]</sup>. Nutritional deficiencies are the principle cause of this disease.

### **Symptoms**

- Infected larvae are unable to free their appendages, eyes or rostrum from their exuviae during moult and become entrapped.
- The larvae generally have malformed appendages and die shortly after moult.

# **Control Measures:**

Adding lecithin to the prepared fed may help to prevent or reduce EED<sup>[9]</sup>.

# IDIOPATHIC MUSCLE NECROSIS (IMN)

This disease is also known as white muscle disease, muscle necrosis, spontaneous muscle necrosis, muscle opacity or milky prawn disease and causes massive larval mortalities in hatcheries. IMN causes mortalities up to 40 to 60% in the post larvae [10]

#### Symptoms:

- This disease appears as focal and multifocal diffused opacity of striated muscle [9-11]
- This disease is considered to be associated with environmental stressors including salinity and temperature fluctuations, hypoxia, hyper activity and overcrowding [10, 11].
- IMN may occur within 1 or 2 days following stocking in production ponds due to stressful conditions.
- <sup>[12]</sup> suggested that the prevalence of IMN in a population of PL could serve as a useful indicator of their general health.

### **Control Measures**

In hatcheries, in general, if the necrosis has not progressed extensively, the disease can be reversed by changing the water <sup>[9]</sup>. There is no effective treatment for this disease besides minimizing the environmental stressors. IMN may occur within one or two days following stocking in production ponds due to stressful conditions. Stocking PL in nursery ponds before releasing into grow-out ponds may reduce this problem. INFESTATION BY MICROORGANISMS

Infestations by microorganisms including microscopic epibiont disease (MED) by a variety of ciliates, which are found attached to the exoskeleton of larvae, are *Zoothamnium* sp., *Epistylis* sp., *Vorticella* sp. and *Acineta* sp. Bacterial Necrosis (BN) caused by bacteria, and the diseases caused by fungi and virus won't cause much damage to the older larvae and post larvae.

### Microscopic Epibiont Disease:

**Symptoms:** A variety of Protozoa may be found attached to the exoskeleton of the larvae (*Zoothamnium* sp., *Epistylis* sp., *Vorticella* sp. and *Acineta* sp). It brings slight opaqueness in body colour. Some species may attack the eggs of brood stock. Others interfere with the feeding and moulting of larvae.

**Control Measures:** These can be generally controlled by formalin treatment. Treatment with 20-30 ppm formalin for 24h in a static bath is effective and safe in controlling larval Zoothamnium infection<sup>[13]</sup>. Repeated treatment of 1-minute 2-ppm acetic acid for dip is recommended for *Epistylis* sp.

### **Bacterial Necrosis:**

### Symptoms:

Larvae affected by this disease turn bluish and stop feeding and the weak larvae sink to the bottom. Brown spots may appear on the antennae and newly formed appendages. The intestinal tracts become empty. Small black spots and lesions are seen on the exoskeleton.

### TREATMENT

All these diseases can be effectively treated and controlled in the hatchery itself for producing healthy post larvae. The disease management largely depends on the use of disinfected water free of pathogens, good quality feeds and general hygiene. MCD does not respond to any tested antibiotics. The best procedure is to discard the larvae and disinfect the infected tanks. If the infection spreads through the hatchery, general disinfectation may be required. Disinfectation requires thorough washing of all tanks, filters and equipment with formalin and/or chlorinated water, followed by drying for at least one week. [7] reported that, the disease can be controlled by low stocking density and improving pond husbandry and sanitation. It is believed that nutritional deficiencies are the principle cause of EED. Adding lecithin to the prepared feed may help to prevent or reduce EED <sup>[9]</sup>. There is no effective

P. Soundarapandian *et al* / Disease Management for the Post Larvae of Fresh Water Cultivable Prawn, *Macrobrachium Malcolmsonii* 

treatment for Idiopathic Muscle Necrosis (IMN) besides minimizing the environmental stressors. Stocking Pl in nursery ponds before releasing into grow-out ponds may reduce this disease. The ciliates can be controlled by either formalin or acetic acid treatments. Bacteria, fungi and virus can be effectively treated with antibiotics like, Bipenicillinstrepromycin, Furnace, Erythromycin Phosphate, Chloramphenical and Novobiocin<sup>[9]</sup>.

#### MANAGEMENT MEASURES

The good quality seeds can be obtained by good management measures. Maintaining stable water conditions in the hatchery is what ensures successful larval rearing. If any, infection spreads through the hatchery, general disinfection through washing the tanks, filters and equipment with formalin and/or chlorinated water, followed by drying for at least one week is necessary.

#### ACKNOWLEDGEMENT

The authors are thanks to former director Dr. T. Kannupandi, Faculty of Marine Sciences and authorities of Annamalai University for providing facilities to carry out the work.

#### REFERENCES

- 1. Kanaujia, D. R. and A. N. Mohanty, 1996. Prospects of Both Mono and Mixed Culture of *Macrobrachium malcolmsonii* . *Fishing Chimes*, 15(12): 33-35.
- 2. Kanaujia, D.R., 2006. Freshwater Prawn Breeding and Culture. In: Handbook of Fisheries and Aquaculture, ICAR, New Delhi, Publication. 293-306.
- Thangadurai, A. J., 1992. Culture of Freshwater Prawn Macrobrachium malcolmsonii in Tamil Nadu - A case study. Proc. of the National Symp. Freshwater Prawns (Macrobrachium spp.), Kerala Agricultural University, Kochi. pp. 187-188.
- 4. Soundarapandian, P., K.S. Prakash and G.K. Dinakaran, 2009. Simple Technology for the Hatchery Seed Production of Giant Palaemonid Prawn *Macrobrachium rosenbergii* (De Man). *Inter. J. Ani. Veter. Advan.*, 1(2): 49-53.
- Sahoo, P.K., 2008. Diseases in Freshwater Prawns and their Management. In: National Training Course Manual on Seed Production and Grow out Culture of Freshwater Prawns, CIFA, Kausalyagang, Bhubaneswar. 61-67.

- Anderson, I.G., G. Nash and M. Shariff, 1990. Mass Larval Mortalities in Giant Freshwater Prawn, *Macrobrachium rosenbergii* de Man, Cultured in Malaysian Modified static "Green water" Systems. J. Fish. Dis., 13: 127-134.
- Brock, A.J., 1983. Diseases (infectious and non-infectious), Metazoan Parasites, Predators, and Public Health Consideration in *Macrobrachium* Culture and Fisheries. In: Handbook of Mariculture, Volume I, Crustacean Aquaculture, J.P. Mcvey (ed.): CRC Press Inc. Florida., 329-370.
- Johnson, D., 1980. Evaluation of various diets for optimal growth and survival of selected life stages of *Artemia*. In G. Persoone, P. Sorgeloos, O. Roels, & E. Jaspers (eds.). The Brine Shrimp Artemia, Universa, Wetteren, Belgium. Vol. 3. p. 185-192.
- Brock, A.J., 1988. Diseases and Husbandry Problems of Cultured *Macrobrachium rosenbergii*. In: Disease Diagnosis and Control in North American Marine Aquaculture. C.J. Sindermann and D.V. Lightner (eds.). *Else. Amst.*, 134-180.
- Nash, G., S. Chinabut and C. Limsuwan, 1987. Idiopathic Muscle Necrosis in the Freshwater Prawn *Macrobrachium rosenbergii* de Man cultured in Thailand. *J. Fish. Dis.*, 10: 109-120.
- 11. Akiyama, D.M., J.A. Brock and S.R. Haley, 1982. Idiopathic muscle necrosis in the cultured freshwater prawn, *Macrobrachium rosenbergii. Vet. Med. Small Clinician*, 1119-1121.
- Sarver, D., S.R. Malecha and D.R. Onizuka, 1982. Possible sources of variability in stocking mortality in post larval *Macrobrachium rosenbergii*, In: Giant Prawn Farming, M.B. New (ed.): Elsevier, Amsterdam. 99-113.
- 13. Roegge, M. A., W. P. Rutledge and W. C. Guest. 1979. Chemical Control of Zoothamnium on larval sp. Macrobrachium acanthurus. pp. 295–299. Proceedings of the Second Biennial Crustacean Health Workshop, Galveston, Texas. April 20-22, 1977. Texas A&M University Sea Grant Publication, 79-114:400.