

REVIEW ARTICLE

Hallmarks of cancer drug resistance and overcoming ways: a paradigm***Muhammad Torequl Islam****Department of Pharmacy, Southern University Bangladesh, Mehedibag-4000, Chittagong, Bangladesh***Received 02 Sep 2017; Revised 10 Oct 2017; Accepted 01 Nov 2017****ABSTRACT**

Over 100 types of cancers, affecting humans still remain most fearful disease in the world. To date, a number of anticancer therapies have been introduced. However, resistance in cancer is a running matter. This review focuses to seek out possible pathways of cancer resistance, ways of overcoming resistance and still remaining challenges to be resolved. The ways concerning cancer resistance are: cross resistance, resistance-promoting adaptive responses, tumor microenvironment, drug detoxification and inactivation, decreased drug accumulation, alterations in drug targets, drug efflux and compartmentalization, altered cell cycle events, alteration in apoptosis, autophagy, angiogenesis and proliferation, DNA damage repair, alterations in membrane lipids, resistance mutations, cancer stem cells and epigenetics. Along with host factors, some other factors such as p53, Pgp, MRP family, BCRP, LRP, sphingosine 1-phosphate, glucosylceramide, Bcl-2 family genes and PTEN also have contributions in anticancer drug resistance. MDR, MDR modulators, multifunctional nanoparticles, iRNA interference, targeted therapy, adaptive immunotherapy, or combination therapies are useful to overcome a number of cancer resistance. However, inevitable recurrences, cancer progression and metastasis along with the efficacy, safety, inherited/ acquired drug resistance, incomplete elucidation of anticancer drug resistance and the host factors are still-remained challenges in cancer therapy.

Keywords: cancer; challenges; resistance; overcome.**INTRODUCTION**

Cancer is known as a severe health threat. Cancer causes 21% death in developed, while 9.5% in developing countries. Cancer cells have the ability to develop resistance to a number of therapies, including traditional therapies. The word 'resistance' comes when pharmaceutical treatments are not enough to combat diseases. Primary and acquired are the two main type resistances. Primary drug resistance exists prior to any given treatment, while acquiring after initial therapy. Targeted therapies have revolutionized cancer treatment; however, the development of drug resistance within the tumor limits their success in cancer patients.

Resistance becomes more dangerous when the resistant cells dominate the population of the tumor^[1]. Cancers targeted to treat with combined drugs, better to be termed as 'multidrug' also undergoing to develop multi-drug resistance (MDR) due to cancer cells are structured and/or functionally different and may have different molecular targets. In intrinsic MDR, cancer cells exhibit resistance to chemotherapy at the initial

anticancer drug exposure. On the other hand, acquired MDR develops resistance during the course of the treatment or upon recurrence of the disease after successful chemotherapy^[2]. Several host factors are also involved in the development of both types of MDR, including those that impair the delivery of anticancer drugs and nullify their cytotoxic effects, and those that alter the genetic or epigenetic factors, leading to drug insensitivity. In MDR, resistance to one drug is accompanied by resistance to other drugs having the structures and mechanisms of action similar or even completely different. The term 'MDR' was first applied to antibiotic-resistant infection^[3] and now applied to cancer chemotherapy. Historically, the first significant advance of MDR came with the identification of the membrane transporter P-glycoprotein (Pgp), then followed by other transporters of the ATP-binding cassette (ABC) family, which is capable to catalyze the efflux of a range of structurally dissimilar anticancer drugs and helps to protect cells by ejecting a wide variety of toxins. Generally, a rapid resistance originates through multiple non-mutational and

non-genetic mechanisms [4]. This paper discusses cancer drug resistance, challenges and overcome pathways.

Source of resistant cancer cells

The occurrence of resistant cells is a broad phenomenon. Sometimes, some factors also play an important role to develop resistant cancer cells, such as hypoxia-inducible factor (HIF), that can cause a progression of renal cell carcinoma and acquired resistance to anti-angiogenic multi-kinase and mechanistic target of rapamycin (mTOR) inhibitors [5]. In general, factors related to genomic and epigenetic alterations, are the major source of resistant cancer cells, as the development of human cancers, a complex multistage process involving an accumulation of both of them [6]. During the progress of the tumor, some cells undergo genetic alterations with a superior growth advantage in a given context. Cancer cell selection obeys the Darwinian law of evolution, hence, under therapeutic pressure; those populations that are most adaptive or resistant to treatment will be selected for. If we can treat them properly, others will be treated so [7]. The resistant cell also possesses some favorable characteristics such as a mutated drug binding site [8] and some stochastic alterations within the cancer cells [9]. Cell breathing time is another important factor in the growth and multiplication of resistant cancer cells. Vaccine therapy may broaden the antigenic breadth and induce the immune responses against autologous cancer cells in cancer stem cells (CSCs) [10]. Thus, cells will find more time to augment their population with a greater extent of the resistance.

Drug resistance in cancer

Resistance often follows initial responses to chemotherapy, although combinations of chemotherapeutic agents led to improved survival [11]. Both primary and acquired resistance can be caused by alterations in drug metabolism (sequestrations or enhanced detoxification) or modifications to the drug targets [12]. Resistance involves drug metabolism, including its uptake, efflux, and detoxification. It may result from the mutations that modify the activity or reduce the expression of surface receptors and transporters. A number of cytotoxic anti-cancer drugs need to undergo metabolic activation. Thus, resistance may develop by decreasing drug activation [13] via downregulation or mutation to the enzymes involving a metabolic pathway [14]. Furthermore, drug inactivation plays a major role in the

development of anticancer drug resistance, through conjugation with reduced glutathione (GSH), which is a powerful anti-oxidant that protects the cells against the damaging effects of reactive oxygen species (ROS) [15].

Many cancer cells have an over reliance or dependency on an oncogene, better to be termed as 'oncogene addiction' [16]. The development of targeted therapies belongs to targeting such oncogenes. However, mutation of the targeted protein(s) may result in drug resistance [17]. On the other hand, amplification of alternative oncogenes or inactivation of alternative survival pathways are two causes of drug resistance related to this phenomenon [18]. Thus, targeting of one protein alone is not sufficient due to other parallel pathways may support tumor growth and survival. This may link to a synthetic lethal relationship [19]. Targeting both pathways may be an effective treatment in this type of resistance [20]. It is evident that, mutation rate in cancer cells is higher than in normal (non-tumor) cells. In chronic cancers, as the tumor grows for years before it is treated, have ample time to develop resistant mutations to emerge and get fixed within the population of tumor cells before beginning of treatment. Although, an aggressive treatment may reduce the overall size of the tumor and population size of the tumor cells, but it can cause mutation and develop *de novo* resistance [1]. Primary mechanisms of anticancer drug resistance have been depicted in **Figure 1**.

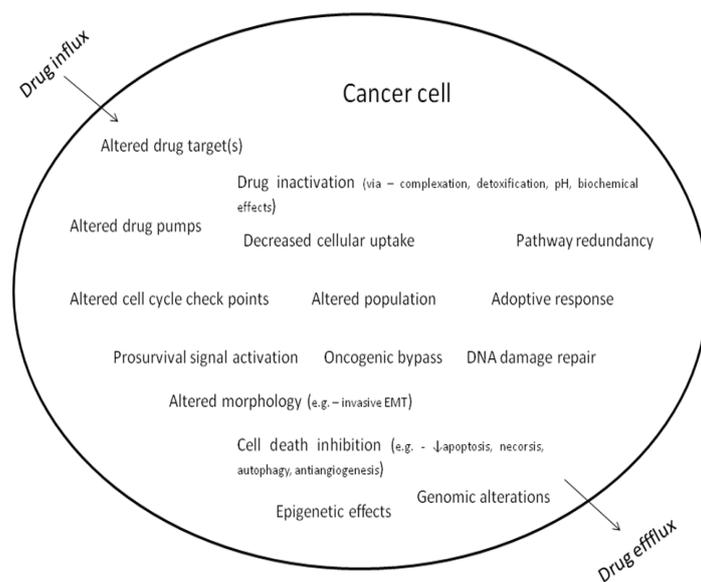


Figure 1. Primary mechanisms include anticancer drug resistance.

Cross resistance

By developing resistance to one drug, a cancer is capable to grow resistant to one or more anticancer drugs [12]. For an example, treatment of

advanced prostate cancer with gonadal testosterone-deprivation leads to the development of castration-resistant prostate cancer (CRPC) [21]. A high degree of cross-resistance was also observed between abiraterone and enzalutamide [22]. Loss of a drug transporter can lead to resistance to structurally diverse compounds [23]. MDR phenotype correlates with poor chemotherapy response to the drugs that are not recognized by transporters, thus evade efflux, efflux inhibitors, as well as drugs that are selectively lethal to Pgp expressing cells, and so on [24].

Resistance-promoting adaptive responses

Activation of prosurvival signalling: Numerous studies have reported for the activation of epidermal growth factor receptor (EGFR) as a resistance mechanism to various chemotherapies [25].

Oncogenic bypass and pathway redundancy: The molecular targeting agents, used to block prosurvival signals and tumor addiction helpful to specific gain-of-function mutations can develop various adaptive resistance mechanisms. For an example, ERBB3 (also known as HER3) and downstream signalling through the PI3K–AKT pathway, an important mechanism of adaptive resistance to EGFR-targeted therapies [26] termed as ‘oncogenic bypass or ‘kinome reprogramming’. In this case, the primary drug target remains unaltered and continues to be inhibited, but an alternative kinase becomes activated gives an adaptive feedback loop as well as a genetic mutation during treatment; emerging as a major mechanism of resistance.

Epithelial-mesenchymal transition (EMT): The epithelial to mesenchymal transition (EMT) causes solid tumors to metastatic. Epithelial cells from losing polarized organization and tight cell-cell junctions undergo changes in cell shape and develop a fibroblast-like morphology associated with increased motility and invasive capacity. There is a link between chemotherapy and targeted therapy resistance and the EMT phenotype [27]. Furthermore, angiogenesis, responsible to cause formation of new blood vessels around metastatic tumors [28] may result survival and resistance of cancer cells. The signaling processes of differentiation, essential for EMT also cause drug resistance in cancer cells [29].

Tumor microenvironment

Each tumor has a unique microenvironment. For example, ECM, cancer-associated fibroblasts, immune and inflammatory cells and blood vessels are found in solid tumors [30], while bone marrow stromal cells, bone marrow endothelial cells, osteoclasts, osteoblasts, macrophages and T cells in haematological malignancies [31]. The protection provided by the microenvironment provides refuge for cancer cells from cytotoxic agents, thus allowing them to evade apoptosis and to develop acquired resistance.

Integrins: Integrins, the cell surface adhesion molecules that connect the cells with ECM [32]. Higher expression of integrins increases survival and drug resistance in cancer cells [33]. Inhibition of apoptosis and altered drug targets is evident in integrin-mediated adhesion to the ECM [34]. Moreover, they can modulate many signalling pathways, such as PI3K–AKT, ERK and NF-κB pathways that promote cell survival and drug resistance [35], especially to the kinase-targeted agents.

Cytokines and growth factors: The cytokines and growth factors through activation of autocrine, paracrine and endocrine oncogenic signalling play key roles anticancer drug resistance, as they can activate various survival signalling pathways.

Drug detoxification and inactivation

Drugs may show underlying pharmacokinetics due to the malmetabolism in cancer cells. Molecules such as glutathione (GSH), a crucial antioxidant, prevents oxidative stress, and keeps redox homeostasis stable in cells are known to cause conjugation with many drugs [36], which essentially detoxify and/or facilitate excretion of drugs, leading to not only escaping cancer cells but also grow resistance.

During drug activation in a host, it interacts with a number of proteins or enzymes. Driver mutations by making a protein constitutively active, in the presence of drug can modify, partially degrade, or complex the drug with other molecules or proteins, causes activation for other kind of activity. Cancer cells can develop resistance through drug inactivation [37]. Drug inactivation also occurs *via* cytochrome P450 (CYP) system and uridine diphospho-glucuronosyltransferase (UGT) super family [38].

Decreased drug accumulation in cancer cells

ATP-dependent efflux pumps, member of ATP-binding cassette (ABC) transporter family,

expression of them is one of the common consequences in the anticancer drug resistance development^[39] *via* decreasing intracellular drug concentrations. A decreased drug uptake causes decrease in drug accumulation in cancer cells. Drugs are transported into the cells *via* several routes such as diffusion across the plasma membrane, loading of the drugs on specific receptors and either receptor mediated or non-specific endocytosis. The problems of these pathways lead to the development of drug resistance in cancer cells^[40].

Alterations in drug targets

A drug's efficacy is influenced by its molecular target and alterations of this target. Alteration in the signal transduction process and modified enzyme expression levels can also cause drug resistance in cancer cells^[41]. However, resistant cancer cells can produce mutated drug target(s) that retains its activity in the cell without being a target of that drug. Therefore, the target is not inhibited by it. Moreover, gene amplification *via* genomic instability in cancer cells may cause alterations in drug targets, which is one of the major causes of the development of acquired drug resistance.

Drug efflux

By reducing the drug accumulation through enhancing efflux, cancer cells can develop drug resistance. ATP-binding cassette (ABC) transporter family proteins, multidrug resistance protein 1 (MDR1), multidrug resistance-associated protein 1 (MRP1), and breast cancer resistance protein (BCRP) are known to cause many anticancer drug resistance^[42]. The constitutive activation of signaling molecules such as kinases causes the cell cycle out of control and results in cancer. These proteins also regulate Pgp expression and are able to develop drug resistance^[43].

Compartmentalization

Sequestration of the drugs in cellular compartments is an important reason to develop anticancer drug resistance. The physicochemical characteristics such as pH partition, generation of an electrical charge, membrane permeability, and so on also results in drug accumulation and anticancer drug resistance^[44].

Altered cell cycle events

In cancer cells an uncontrolled cell proliferation arises from the defects throughout the cell cycle

progression at G1, G2, S and mitotic phases. Cell cycle checkpoints, including a network of protein kinase signaling pathways, protect the cells from DNA damage induced by chemotherapeutic agents and provide the cells appropriate time to repair the damages^[45]. Thus, defects in cell cycle checkpoints can cause not only carcinogenesis, but also drug resistance.

Apoptosis, autophagy, angiogenesis and host factors in drug resistance

Apoptosis is recognized as a unique form of cell death. In necroptosis, a caspase-independent cell death occurs through receptor-interacting protein kinases (RIP1 and RIP3) or mixed lineage kinase domain-like protein (MLKL). Although, its importance in cancer treatment is controversial, but it is evident to develop resistance to an altered apoptosis in cancer cells^[46]. The Bcl-2, anti-apoptotic family of proteins elicits a broad cell survival program through promoting cell migration, invasion, and metastasis. In cancer cells, there is a higher expression level of these family proteins^[47], which is responsible for the development of anticancer drug resistance^[48]. Autophagy is a complex issue because it can play both a pro-death and a pro-survival role^[49]. Autophagy can affect the sensitivity of tumor cells to the treatments^[50]. In a study, cancer stem-like cells (CSCs) were found to develop resistance against photodynamic therapy (PDT)^[51]. In another study, TXNDC17 (thioredoxin domain containing 17)-mediated taxol resistance was evident *via* enhancing autophagy in colorectal cancer cells^[52]. Antiangiogenic therapy can also develop drug resistance, through multiple mechanisms^[53], including VEGF and VEGF receptor (VEGF-R), FGF and FGF receptor (FGF-R), and PDGF and PDGF receptor (PDGF-R) pathways. It may select metastatic clones to avoid hypoxia, thus clonal evolution gradually leads to aggressiveness of the cancer situation and develop resistance to treatment. In a recent study, regorafenib was found to progress the malignancy *via* prevention of autocrine and paracrine VEGF signaling in colorectal cancer cells^[54].

The 'host' can also play a major role in the development of cancer resistance. The tumor microenvironment is heterogeneous and can help to support a variety of resistance mechanisms. For example, cellular pathways affected by tumor hypoxia, can develop resistance^[55]. CSCs by using some mechanisms such as expression of ABC transporters, aldehyde dehydrogenase system, prosurvival proteins altered DNA damage

response and altered signaling pathways can develop resistance [56]. Abdel-Hafiz suggested that CSCs play an important role in driving metastasis and tamoxifen resistance in breast cancer cells [57].

MDR involving resistance to apoptosis

Suppression of pathways leading to apoptosis is thought to be an intrinsic feature of cancer cells [58]. Conversely, cytotoxic anticancer drugs commonly induce stress pathways such as p38 kinase [59] or suppress signalling pathways such as those coordinated by phosphatidyl-3-phosphate kinase (PI3K) and extracellular-regulated kinase-1 (ERK1) to reactivate pathways to apoptosis leading to pre-existing or acquired MDR, which increases suppression of apoptosis pathways. Inactivating mutations of the gene for p53 protein, activating mutations of the gene for PI3K, loss of expression of PTEN (a phosphatase controlling PI3K activity) and activating mutations of the genes for the RAS/RAF affect the balance of activity of the Bcl-2 family proteins and their relatives [60], which in turn control transition to apoptosis by modulating the stability of the outer mitochondrial membrane. Cytochrome *c* can induce a cascade of caspase enzymes that lead to convert the cell too small fragments, which can be recognized and ingested by nearby phagocytes. Loss of regulation of PI3K leads to increased activity of AKT (PKB), phosphorylation of bad, a member of the Bcl-2 family, results protection of mitochondria from the permeability transition and increased resistance to cell death. Similarly, activating mutations in the RAS or RAF genes lead to activation of the ERK1 enzyme, inactivated by phosphorylation of bid, another member of the Bcl-2 family and protection of mitochondria from the permeability transition. Phosphorylation of Bcl-2, following activation of the NF- κ B transcription factor and its downstream target twist-1 by cytokines and other cellular stress, can lead to apoptosis resistance [61]. Induced overexpression of Bcl-2 itself can also provide a mechanism of apoptosis resistance [62].

Altered proliferation

Apoptosis is the normal response to DNA damage that cannot be repaired. But the threshold of death is much higher in cells that are not growing [63]. A part of a transient reduction in cell growth is mediated by p53, as the levels of this protein rise and reduces cell cycle by stimulating apoptosis at a certain threshold [64]. MicroRNAs regulate gene expression at the post-transcriptional level, and are involved in many different biological

processes, including cell proliferation, differentiation, metabolism, stress response, and apoptosis. The aberrant expression of microRNAs plays a major pathogenic role in the carcinogenic process. MicroRNAs also play an important role in anticancer drug inducing resistance [65]. Thus, the mechanisms involving altered proliferation of cancer cells are linked to resistance directly or indirectly.

DNA damage repair

Cancers must acquire permanent genomic mutations. Once a mutation is acquired cancers become addicted to a different DNA repair pathway. Therefore, the repair of damaged DNA permits anticancer drug resistance via directly or indirectly reversing damaged DNA, and even DNA damage response (DDR) mechanisms [66].

Alterations in membrane lipids

Cancer cells are different from the differences in lipid profiles from their healthy forms (Leach, 1996). Alterations in membrane lipids are important factors for acquiring resistance and even MDR. Cieřlik-Boczula et al stated that the formation of domains with different content of pyrimidine analog (FPh-prm)/dipalmitoyl-sn-glycero-3-phosphocholine (DPPC) can be responsible for the membrane-related mechanism of chemoprevention of pyrimidine analogs [67]. It was also demonstrated that an inhibition of the outward transport of anticancer drugs by the Pgp in cancer cells may be the core mechanism in that case.

p53 in anticancer drug resistance

The protein 53 (p53) is an important transcription factor with tumor suppressor functions [68]. Except, of the stress conditions, cells have inactive p53 protein with a short half-life at low levels. In cellular stressful conditions such as DNA damage, hypoxia, nitric oxide exposure, decrease in ribonucleotides and oncogenic signaling, p53 becomes activated and intracellular levels of p53 proteins are increased [69]. Activation of p53 prevents cellular proliferation *via* arresting cell cycle at G1 or G2 phase or *via* triggering apoptotic signals. In case of any damage of DNA, p53 induces cell cycle arrest and provides the cell an appropriate time needed for repairing the DNA [70]. When the damage is serious that cannot be repaired, p53, in turn, makes the cells undergo apoptosis [68]. Activation and inactivation of p53 is regulated by the phosphorylation, ubiquitylation, acetylation and interactions with

other proteins ^[71]. Murine double minute 2 (Mdm2) is the protein that majorly regulates p53 activity *via* protecting it from interacting with the transcription proteins through binding to the transcriptional activation domain of p53, and also *via* providing the ubiquitin ligase-mediated degradation of p53 ^[72]. To date, abnormalities in p53 function have been reported in more than 60% of cancers ^[73]. These abnormalities lead to uncontrolled cell proliferation as DNA cannot be repaired and cell cycle cannot be arrested in the cells having mutated p53, leading to development of anticancer drug resistance ^[74].

Pgp in anticancer drug resistance

Among the ABC transporters involved in MDR, Pgp is the most common efflux pump in the plasma membrane ^[75]. Structurally, Pgp is a single polypeptide including two homologous parts, each having a hydrophobic transmembrane domain (TMD) and a nucleotide-binding domain (NBD); those are separated by an intracellular linker region. Each TMD consists of six membrane spanning helices. Transmembrane domains are responsible for the specificity of substrate drugs by forming channels, whereas nucleotide-binding domains participate in ATP binding and hydrolysis ^[76]. Pgp is able to bind a large variety of hydrophobic drugs, especially the conjugated anticancer drugs ^[77].

MRP family in anticancer drug resistance

Not only Pgp over expression but also MDR associated protein 1 (MRP1/ABCC1) was found to be amplified in anticancer drug resistance ^[78], as they also pump toxic substances out of the cell in an ATP-dependent manner ^[79]. The drugs transported by ABCC1 are similar to Pgp substrates (except taxanes). Unlike ABCB1, ABCC1 can also export drugs modified by glycosylation, sulfation and glutathione ^[80, 81]. The other members of MRP family, MRP2/ABCC2, MRP3/ABCC3, MRP6/ABCC6 and MRP7/ABCC10 have been also evident to develop resistance to anticancer agents ^[82].

BCRP in anticancer drug resistance

The other ABC transporter involved in MDR is breast cancer resistance protein (BCRP/ABCG2). In contrast to Pgp and MRP1, it contains only one NBD precedes one TMD with six membranes spanning helices ^[83]. BCRP has a potential role in drug resistance in hematologic malignancies like AML and CML due to its frequent expression on malignant hematopoietic and lymphoid cells ^[84].

Some BCRP inhibitors, transported out of the cell at low concentrations, at high concentrations are capable to inhibit ABC transporters.

LRP in anticancer drug resistance

Lung resistance related protein or major vault protein (LRP/MVP) is not localized to the cell membrane like MDR1 and MRP1. LRP, found in the cytoplasm is a main component of multimeric vaults and associated with cytoskeletal elements as well as nuclear membrane that help to transport drugs from the nucleus to the cytoplasm *via* vesicular trafficking ^[79]. LRP can cause anticancer drug resistance in some cancer cells ^[85]. For an example, Zhang et al stated that LRP-1 as well as MRP-1 were increased in NDRG1 over expressing cells, implying NDRG1-mediated pathways in multidrug resistance of neuroblastoma ^[86].

Sphingosine 1-phosphate in anticancer drug resistance

Sphingosine 1-phosphate (S1P) is another important factor for developing anticancer drug resistance. Generally, S1P favors proliferation, angiogenesis and cell survival. A shift in the balance toward S1P was seen in glioblastoma (GBM) and other cancers, and resulted in tumor cell survival and resistance to anticancer chemotherapy ^[87]. Sphingosine kinase-1 (SK1-1), which is responsible for the synthesis of S1P, was reported to decrease the apoptotic effects of anticancer drugs ^[88].

Glucosylceramide in anticancer drug resistance

Glucosylceramide (GC) is also found to be increased in drug resistant cancer cells ^[89]. Suppression of glucosylceramide synthase (GCS) activity results in a decrease in MDR1 expression levels, and this reverses anticancer drug resistance ^[90]. The glycosylation by GCS is a critical step regulating the modulation of cellular activities by controlling ceramide and glycosphingolipids (GSLs). Chemotherapy or other stresses can increase in ceramide, which drives cells to proliferation arrest and apoptosis or autophagy. Furthermore, ceramide glycosylation promptly eliminates ceramide and its induced processes, leads protecting cancer cells. An enhanced ceramide glycosylation can increase GSLs, participating in selecting cancer cells to drug resistance. Ceramide glycosylation by GCS is a rate-limiting step in GSL synthesis, thus inhibition of GCS sensitizes cancer cells to anticancer drugs and eradicates CSCs. If ceramide glycosylation

remains uncontrolled it can modulate gene expression and decrease MDR1 through the cSrc/ β -catenin pathway and restoring p53 expression *via* RNA splicing [91]. GCS is also known to de-activate in the regulation of apoptosis progression in a number of cancer cells [92], it may be due to the regulation of apoptosis-related proteins such as Bcl-xL [93].

Bcl-2 family genes in anticancer drug resistance

Bcl-2 is an oncogene, which enhances cancer cell proliferation and suppresses apoptosis. Bcl-2 is evident to express in an uncontrolled manner in various cancer cells. The family of Bcl-2 also comprises the genes encoding proapoptotic Bax, Bad, Bim and antiapoptotic Bcl-xL and Bcl-2 proteins [94]. The presence of CSCs is important in the prevention of therapy failure and tumor recurrence. Upregulation of Nrf2 leads to the overexpression of drug efflux proteins such as ABCG2 in CSCs, resulting in cancer treatment failure and cancer relapse. In cervical CSCs, an aberrant upregulation of Nrf2 along with an elevated transcriptional regulation of ABCG2, Bcl-2 and Bmi-1 was seen, that resulted prolonged cell survival, infinite cell proliferation and highly resistant apoptosis [95].

PTEN in anticancer drug resistance

The phosphatase and tensin homolog, PTEN, is a tumor suppressor which has phosphatase activity, preventing PI3K/Akt signaling pathway, known as an important cancer promoting pathway [96], as it plays a key role in cancer development and maintenance of CSCs [97], thus results development of drug resistant cancer cells.

Resistance mutations

Resistance mutations are of two types (a) mutations that interfere binding of the drug and (b) mutations that make the drug target perform their biological function more efficiently. In the first case, it occurs by altering the active domain of the protein (e.g.- the catalytic domain of an enzyme) directly or indirectly. Direct interference happens by altering residues at the binding site, while indirect interactions modify the dynamics of the protein, such as by destabilizing an inactive state that binds to the drug, and hence efficiency of the whole protein [98].

Epigenetics-mediated cancer drug resistance

The two main types of epigenetic changes are DNA methylation and histone modification *via*

acetylation or methylation [99]. Epigenetic mechanisms can also influence DNA damage repair. DNA mismatch repair processes can be lost due to hypermethylation of the human mutL homolog 1 (hMLH1) gene promoter, and this can lead to cancer development [100]. The DNA repair enzyme MGMT inhibits the killing of tumor cells by alkylating chemotherapy agents. Methylation of *MGMT* causes gene silencing and decreased *MGMT* production. Epigenetic alteration of *MGMT* expression has been associated with a modified chromatin configuration [101].

Cancer stem cell and drug resistance

The cancer cell population is very heterogeneous, as the cells within a tumor are not equivalent, neither structurally nor genetically, which is an obstacle to targeted treatment. Heterogeneous populations have stem cell properties and are usually drug resistant. However, another small fraction of adult cancer cells also possesses drug resistance capabilities [102,103]. The bulk tumor cells lack the capacity for self-renew. Only the CSCs are capable of self reproduction, which are insensitive to chemotherapy and even to radiation therapy. CSCs can reside in a dormant state for a long duration and have metastatic property [104], supporting the three hallmarks of cancer [58] such as evading apoptosis, limitless replicative potential and tissue invasion as well as metastasis. If all cells within a tumor are able to divide and inherit genetic changes to the next generation, there is a strong selection pressure of cancer therapy, thus leading to develop resistance [105]. Source of cancer stem cells and possible control by the tumor microenvironment of them have been shown in **Figure 2**.

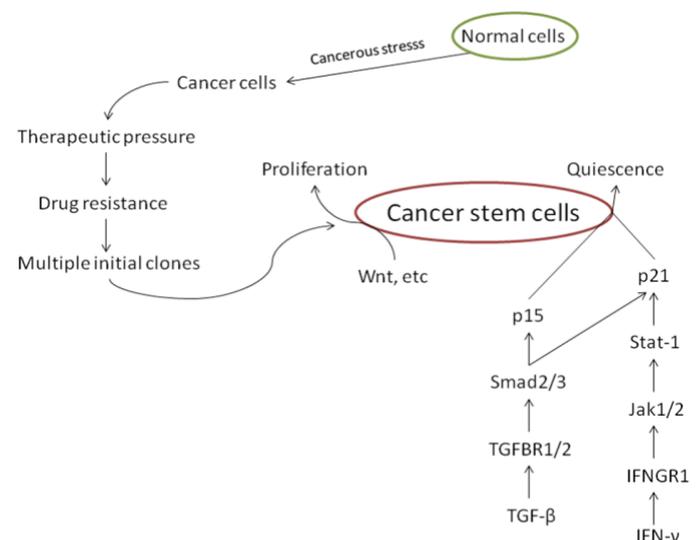


Figure 2. Occurrence of tumor stem cells and hypothetical control by the microenvironment.

Tumour heterogeneity in resistance

Tumor heterogeneity having stem cell property can arise resistance and even MDR in a number of cancer cells. The generation heterogeneity through mutations or epigenetic changes, affect not only cell cycle entry, but also centrosome replication [106], and centrosome defects lead during mitosis to chromosome instability, tetraploidy and aneuploidy. Moreover, microsatellite instability and defective DNA repair can also contribute to a so-called mutator phenotype, while tumor cells may develop large numbers of genetic changes [107].

Drug resistance is a consequence of endogenous defense mechanisms

Drug resistance can be seen as a necessary evolutionary consequence of the body's need to get rid of toxins [108] or xenobiotics. Protection mechanisms involving proteins that pump the drugs out of the cells (e.g.- Pgp), mutation of the target of a particular toxin, and activation of alternate biological pathways instead of the one hit by a toxin. Host defense peptides have been demonstrated to exhibit prominent advantages in cancer therapy that can overcome the limitations of traditional chemotherapy agents, such as toxicity on non-malignant cells and the emergence of drug resistance [109].

Evolutionary ways of overcoming anticancer drug resistance**MDR mechanisms controlling drug efflux from cells**

At least, 48 structurally related transporters, known collectively as the ABC family, are known [110], and the three subfamilies concerned with drug transport are the 'B' subfamily that includes Pgp, the 'C' subfamily that includes MRP-related (multi-resistance protein related) transporters and the 'G' subfamily that includes the ABCG2, MXR and ABCP proteins. Some of these transporters act directly on the drug while others act on conjugates concerting with the cellular conjugating enzymes that first link the drug to glutathione, glucuronide or sulphate. Expression of ABCG2 and MRP1 has been reported inside populations of tumor cells lines with some of the characteristics of stem cells [111], consistent with the hypothesis that CSCs commonly express these drug efflux proteins [112]. To overcome this type of resistance, GST inhibitors and substrates for GSH conjugation are used.

Overcoming of transport MDR

There are two main approaches to overcoming transport MDR [80]. The first is to co-administer a drug that inhibits the action of the transporter [113] and the second is to design or utilize anticancer drugs whose activity is not significantly altered by the presence of transporters [114]. First, second and third generation inhibitors are the best choices to overcome this kind to resistance. Drugs that are insensitive to transport resistance mechanisms, requires they should be taken up efficiently into the cells and is relatively resistant to conjugation with glutathione, glucuronide or sulphate. In this way, the rate of uptake rate greatly exceeds the potential rate of transporter-mediated efflux. Cellular uptake of drugs by diffusion is generally controlled by diffusion; lipophilic drugs can enter into the cells rapidly by diffusion. In a recent study, Jin et al suggested that MUC1 has an important role in the induction of drug resistance [23]. Through stimulation of EGFR activation and nuclear translocation, MUC1 increased the expression of ATP-binding cassette transporter B1 (ABCB1). Thus, a targeted suppression of EGFR or ABCB1 may effectively reverse chemoresistance.

MDR involving host immune responses

Interleukins (e.g.- IL-6 and -8) induce senescence of tumor cells [115]. As a consequence of cell turnover, tumor apoptotic is released continuously from the tumor and taken up by tumor macrophages and dendritic cells with a resultant immunosuppressive response [116]. T-cell mediated release of interferon-gamma (IFN- γ) potentially prevent the proliferation of CSCs, and therapy reduces interferon release could allow the continuation and even with stimulation of tumor growth. HMGB1, a high mobility group protein associated with chromatin, and calreticulin [117], a protein associated with the endoplasmic reticulum. Substances that facilitate the production or effects of such proteins can be used to augment responses of tumor cells to anticancer drugs. Release of chemokines within the tumor leads to the recruitment of immature macrophages to tumor tissue, but until they are appropriately activated these may lead cancer resistance to immunotherapy. Adoptive transfer of NK cells may be helpful in cancer therapy, as they have natural cytotoxicity against tumor cells and safety upon adoptive transfer of patients [118]. Partial or complete loss of HLA class I expression is evident in a wide spectrum of human tumor types, which may result from immune selection of escape variants by tumor-specific CD8 T cells, that

generally links to acquired resistance to checkpoint inhibition therapy [119].

MDR modulators

Numerous therapeutic agents, classified as first, second and third generation are MDR modulators. Pgp and LRP-mediated drug resistance is vastly studied in MDR modulatory resistance. Moreover, the membrane transporter proteins ABCG2 (BCRP) and ABCC1 (MRP1) are also involved in the formation of MDR in cancer chemotherapy [120]. Recently, many promising strategies have emerged using natural resources such as plants, fungi, and even marine organisms to overcome MDR. These modulators are low in toxicity and are well tolerated in the human body. One of the approaches to reverse MDR in cancer treatment is the inhibition of Pgp, while LRP-mediated drug resistance could be reversed by a pyrimidine analog.

Multifunctional nanoparticles for targeted chemotherapy

The rapid growth of solid tumor results in the altered physiology of the tumor leads to leaky and defective vasculatures. The increased vascular permeability coupled with impaired lymphatic drainage in the tumor produces an enhanced permeability and retention (EPR) effect, which is often referred to as passive targeting [121]. Targeted nanomedicines can overcome various limitations of conventional chemotherapy; enhance selectivity, early and more precise cancer diagnosis, individualized treatment as well as overcoming drug resistance [122,123].

RNA interference (iRNA) therapy

RNA interference (RNAi) therapeutics (e.g. siRNA, miRNA, etc.) is going to be a popular medicinal remedy for a variety of ailments, including cancer [124]. iRNA, a biological process in which cells use to inhibit or silence specific gene expression through the destruction of specific messenger RNA (mRNA) molecules triggered by RNA molecules. Typically, iRNA can be achieved through two different pathways: (a) an RNA-based approach where effector small interfering RNA (siRNAs) is delivered to the target cells, and (b) a DNA-based approach in which effective siRNAs are generated by the intracellular processing of RNA hairpin transcripts [125]. However, delivering siRNAs to targeted tumor sites still a challenge. Naked siRNAs can be rapidly degraded by serum ribonucleases and can hardly cross the cell membrane due to their

polyanionic nature and relatively large molecular weight [126]. Nanocarriers designed for iRNA therapy can be synthesized from a variety of materials, including polymers (e.g. - biodegradable polymers), metal oxides, carbon nanotubes, and modified nanoparticles (e.g. - lipid-modified dextran nanoparticles) are helpful to manage MDR cancers [127,128].

Combinatorial nanoparticles loaded with both anticancer drugs and MDR modulators/siRNAs

Combinatorial nanoparticles, formulated with both MDR modulators and chemotherapeutic agents, have been used successfully to restore the sensitivity of tumor cells and to enhance the therapeutic efficacy of cancer treatment. Co-administration of anticancer drugs and siRNAs is also another effective strategy to enhance the therapeutic efficacy of cancer treatment [129]. Moreover, a combination of epigenetic drugs with conventional chemotherapy should be more effective in treating tumors and drug resistant cancers.

Miscellaneous

Membrane lipids-mediated MDR can be overcome by using lipophilic cationic agents. Drug resistance, including MDR arisen from p53 function can be overcome by targeting p53 expression, while drug resistance arising from the over expression of GCS can be reversed by inhibiting GCS activity [130]. In order to overcome Bcl-2-related resistance, agents targeting Bcl-2 family members can be used, while resistance arisen by PTEN mutation could be overcome *via* targeting PI3K/Akt signaling, and also *via* increasing intracellular levels of PTEN [131].

Major obstacles to success in reversing clinical MDR

Early termination or clinical failure of MDR modulators are two major problems in mutli-drug therapy in cancer patients. Unexpected and undesired pharmacokinetic interactions between the modulators and the anti-cancer drugs used for the treatment of patients, reduces doses of anticancer drugs, thus the inefficient benefit [132]. The multifactorial nature of MDR is another obstacle, causing inefficient therapy of patients. However, not only ABC transporter but also different other transporters could be responsible for removal of an anticancer drug [133]. Several properties of membrane such as the fluidity and lipid density are also interested in MDR.

However, MDR could be prevented by targeting both pH changes and transporters.

Some challenges yet to resolved related ocancer therapy

Not only anticancer drug resistance, but also inevitable recurrences, cancer progression and metastasis are also still-remained challenging to be solved ^[134]. The combine efficacy, safety of the healthy cells, convenience and narrow therapeutic index of some drugs are also some potential challenges in the development of anticancer drugs ^[135]. Doubtless, an inherited/acquired drug resistance is one of the major challenges of the chemotherapy ^[136]. A complete molecular mechanisms underlying combination or multi-drug treatment is often unclear ^[137]. Incomplete elucidation of anticancer drug resistance is another important problem ^[138]. The side effects, toxicity, lack of selectivity and rapid resistance rate of most of the chemotherapeutic agents limiting their use in anticancer treatments ^[139]. Correction of the host factors, mainly the pathophysiology and related other factors are a great challenge in cancer therapy.

CONCLUSION

Undoubtedly, cancer remains one of the main sources of research in the world. Along with some effective treatments, a number of ways have been identified to cause resistance in cancer, stimulating scientists to search more effective, safer as well as compatible treatment strategies in this fearful disease. Existing problems must be solved before getting others. From this viewpoint, existing challenges are the crucial facts in an effective cancer treatment.

Conflicts of interest

None declared.

References

1. Friedman R. Drug resistance in cancer: molecular evolution and compensatory proliferation. *Oncotarget* 2016; 7: 11746-11755.
2. Shaffer BC, Gillet JP, Patel C, Baer MR, Bates SE, Gottesman MM. Drug resistance: still a daunting challenge to the successful treatment of AML. *Drug Resist Updat* 2012;15: 62-9.
3. Bolhuis H, Van Veen HW, Poolman B, Driessen AJ, Konings WN. Mechanisms of multidrug transporters. *FEMS Microbiol Rev* 1997;21:55-84.
4. Glaysher S, Gabriel FG, Johnson P, Polak M, Knight LA, Parker K, Poole M, Narayanan A, Cree IA; NHS Collaborative Research Programme for Predictive Oncology. Molecular basis of chemosensitivity of platinum pre-treated ovarian cancer to chemotherapy. *Br J Cancer* 2010;103:656-62.
5. Yoshino H, Nohata N, Miyamoto K, Yonemori M, Sakaguchi T, Sugita S, Itesako T, Kofuji S, Nakagawa M, Dahiya R, Enokida H. PHGDH as a key enzyme for serine biosynthesis in HIF2 α targeting therapy for renal cell carcinoma. *Cancer Res* 2017. doi: 10.1158/0008-5472.CAN-17-1589.
6. Caulin AF, Maley CC. Peto's Paradox: evolution's prescription for cancer prevention. *Trends Ecol Evol* 2011;26:175-82.
7. Greaves M. Darwinian medicine: a case for cancer. *Nat Rev Cancer* 2007;7:213-21.
8. Ricci-Vitiani L, Lombardi DG, Pilozzi E, Biffoni M, Todaro M, Peschle C, De Maria R. Identification and expansion of human colon- cancer-initiating cells. *Nature* 2007;445:111-5.
9. Negrini S, Gorgoulis VG, Halazonetis TD. Genomic instability - an evolving hallmark of cancer. *Nat Rev Mol Cell Biol* 2010; 11:220-8.
10. Zhao F, Zhang R, Wang J, Wu D, Pan M, Li M, Guo M, Dou J. Effective tumor immunity to melanoma mediated by B16F10 cancer stem cell vaccine. *Int Immunopharmacol.* 2017;52:238-44. doi: 10.1016/j.intimp.2017.09.019.
11. Cree IA, Charlton P. Molecular chess? Hallmarks of anti-cancer drug resistance. Cree and Charlton *BMC Cancer* 2017;17:10. doi: 10.1186/s12885-016-2999-1.
12. Ullah MF. Cancer multidrug resistance (MDR): a major impediment to effective chemotherapy. *Asian Pac J Cancer Prev* 2008;9:1-6.
13. Bardenheuer W, Lehmborg K, Rattmann I, Brueckner A, Schneider A, Sorg UR, Seeber S, Moritz T, Flasshove M. Resistance to cytarabine and gemcitabine and in vitro selection of transduced cells a after retroviral expression of cytidine deaminase in human hematopoietic

- progenitor cells. *Leukemia* 2005;19:2281-8.
14. Sampath D, Cortes J, Estrov Z, Du M, Shi Z, Andreeff M, Gandhi V, Plunkett W. Pharmacodynamics of cytarabine alone and in combination with 7-hydroxystaurosporine(UCN- 01) in AML blasts *in vitro* and during a clinical trial. *Blood* 2006;107:2517-24.
 15. Wilson TR, Longley DB, Johnston PG. Chemoresistance in solid tumours. *Ann Oncol* 2006;17:x315-24.
 16. Sharma SV, Settleman J. Oncogene addiction: setting the stage for molecularly targeted cancer therapy. *Genes Dev* 2007;21:3214-31.
 17. Wong AL, Lee SC. Mechanisms of resistance to trastuzumab and novel therapeutic strategies in HER2-positive breast cancer. *Int J Breast Cancer* 2012;2012:415170.
 18. Engelman JA, Zejnullahu K, Mitsudomi T, Song Y, Hyland C, Park JO, Lindeman N, Gale CM, Zhao X, Christensen J, Kosaka T, Holmes AJ, Rogers AM, Cappuzzo F, Mok T, Lee C, Johnson BE, Cantley LC, Jänne PA. MET amplification leads to gefitinib resistance in lung cancer by activating ERBB3 signaling. *Sci* 2007;316:1039-43.
 19. Tucker CL, Fields S. Lethal combinations. *Nat Genet* 2003;35:204-5.
 20. Nijman SM. Synthetic lethality: general principles, utility and detection using genetic screens in human cells. *FEBS Lett* 2011;585:1-6.
 21. Chang KH, Ercole CE, Sharifi N. Androgen metabolism in prostate cancer: from molecular mechanisms to clinical consequences. *Br J Cancer* 2014;111:1249-54.
 22. Antonarakis ES, Lu C, Wang H, Lubner B, Nakazawa M, Roeser JC, et al. AR-V7 and resistance to enzalutamide and abiraterone in prostate cancer. *N Engl J Med* 2014;371:1028-38.
 23. Jin W, Liao X, Lv Y, Pang Z, Wang Y, Li Q, Liao Y, Ye Q, Chen G, Zhao K, Huang L. MUC1 induces acquired chemoresistance by upregulating ABCB1 in EGFR-dependent manner. *Cell Death Dis* 2017;8:e2980. doi: 10.1038/cddis.2017.378.
 24. Nobili S, Landini I, Mazzei T, Mini E. Overcoming tumor multidrug resistance using drugs able to evade P-glycoprotein or to exploit its expression. *Med Res Rev* 2012;32:1220-62.
 25. Van Schaeybroeck S, Kelly DM, Kyula J, Stokesberry S, Fennell DA, Johnston PG, Longley DB. Src and ADAM-17-mediated shedding of transforming growth factor- α is a mechanism of acute resistance to TRAIL. *Cancer Res* 2008;68:8312-21.
 26. Wheeler DL, Huang S, Kruser TJ, Nechrebecki MM, Armstrong EA, Benavente S, Gondi V, Hsu KT, Harari PM. Mechanisms of acquired resistance to cetuximab: role of HER (ErbB) family members. *Oncogene* 2008;27:3944-56.
 27. Yao Z, Fenoglio S, Gao DC, Camiolo M, Stiles B, Lindsted T, Schleder M, Johns C, Altorki N, Mittal V, Kenner L, Sordella R. TGF- β IL-6 axis mediates selective and adaptive mechanisms of resistance to molecular targeted therapy in lung cancer. *Proc Natl Acad Sci USA* 2010;107:15535-40.
 28. Shang Y, Cai X, Fan D. Roles of epithelial-mesenchymal transition in cancer drug resistance. *Curr Cancer Drug Targets* 2013;13:915-29.
 29. Bates RC, Mercurio AM. The epithelial-mesenchymal transition (EMT) and colorectal cancer progression. *Cancer Biol Ther* 2005;4:365-70.
 30. Shiao SL, Ganesan AP, Rugo HS, Coussens LM. Immune microenvironments in solid tumors: new targets for therapy. *Genes Dev* 2011;25:2559-72.
 31. Bhatia R, McGlave PB, Dewald GW, Blazar BR, Verfaillie CM. Abnormal function of the bone marrow microenvironment in chronic myelogenous leukemia: role of malignant stromal macrophages. *Blood* 1995;85:3636-45.
 32. Ruoslahti E, Pierschbacher MD. New perspectives in cell adhesion: RGD and integrins. *Science* 1987;238:491-7.
 33. Hoyt K, Castaneda B, Zhang M, Nigwekar P, di Sant'agnese PA, Joseph JV, Strang J, Rubens DJ, Parker KJ. Tissue elasticity properties as biomarkers for prostate cancer. *Cancer Biomark* 2008;4:213-25.
 34. Damiano JS. Integrins as novel drug targets for overcoming innate drug resistance. *Curr Cancer Drug Targets* 2002;2:37-43.

35. Danen EH. Integrins: regulators of tissue function and cancer progression. *Curr Pharm Des* 2005;11:881-91.
36. Di Nicolantonio F, Mercer SJ, Knight LA, Gabriel FG, Whitehouse PA, Sharma S, Fernando A, Glaysher S, Di Palma S, Johnson P, Somers SS, Toh S, Higgins B, Lamont A, Gulliford T, Hurren J, Yiangou C, Cree IA. Cancer cell adaptation to chemotherapy. *BMC Cancer* 2005;5:78.
37. Gawryluk JW, Wang JF, Andrezza AC, Shao L, Young LT. Decreased levels of glutathione, the major brain antioxidant, in post-mortem prefrontal cortex from patients with psychiatric disorders. *Int J Neuropsychopharmacol* 2011;14:123-30.
38. Michael M, Doherty MM. Tumoral drug metabolism: Overview and its implications for cancer therapy. *J Clin Oncol* 2005;23:205-29.
39. Dean M, Rzhetsky A, Alliknets R. The human ATP-binding cassette (ABC) transporter superfamily. *Genome Res* 2001;11:1156-66.
40. Gottesman MM. Mechanisms of cancer drug resistance. *Ann Rev Med* 2002;53:615-27.
41. Luqmani YA. Mechanisms of drug resistance in cancer chemotherapy. *Med Princ Pract* 2005;14:35-48.
42. Hilgendorf C, Ahlin G, Seithel A, Artursson P, Ungell A, Karlsson J. Expression of thirty-six drug transporter genes in human intestine, liver, kidney, and organotypic cell lines. *Drug Metab Dispos* 2007;35:1333-40.
43. Mutoh K, Tsukahara S, Mitsuhashi J, Katayama K, Sugimoto Y. Estrogen-mediated post transcriptional downregulation of P-glycoprotein in *MDR1*-transduced human breast cancer cells. *Cancer Sci* 2006;97:1198-204.
44. Duvvuri M, Krise JP. Intracellular drug sequestration events associated with the emergence of multidrug resistance: a mechanistic review. *Front Biosci* 2005;10:1499-509.
45. Sancar A, Lindsey-Boltz LA, Unsal-Kacmaz K, Linn S. Molecular mechanisms of mammalian DNA repair and the DNA damage checkpoints. *Annu Rev Biochem* 2004;73:39-85.
46. Su Z, Yang Z, Xu Y, Chen Y, Yu Q. Apoptosis, autophagy, necroptosis, and cancer metastasis. *Mol Cancer* 2015;14:48.
47. Yu Y, Xu L, Qi L, Wang C, Xu N, Liu S, Li S, Tian H, Liu W, Xu Y, Li Z. ABT737 induces mitochondrial pathway apoptosis and mitophagy by regulating DRP1-dependent mitochondrial fission in human ovarian cancer cells. *Biomed Pharmacother* 2017;96:22-9. doi:10.1016/j.biopha.2017.09.111.
48. Zhou QM, Sun Y, Lu YY, Zhang H, Chen QL, Su SB. Curcumin reduces mitomycin C resistance in breast cancer stem cells by regulating Bcl-2 family-mediated apoptosis. *Cancer Cell Int* 2017;17:84. doi: 10.1186/s12935-017-0453-3.
49. Martinez-Outschoorn UE, Whitaker-Menezes D, Pavlides S, Chiavarina B, Bonuccelli G, Casey T, Tsigos A, Migneco G, Witkiewicz A, Balliet R, Mercier I, Wang C, Flomenberg N, Howell A, Lin Z, Caro J, Pestell RG, Sotgia F, Lisanti MP. The autophagic tumor stroma model of cancer or “battery-operated tumor growth”: A simple solution to the autophagy paradox. *Cell Cycle* 2010;9:4297-306.
50. Cheng CY, Liu JC, Wang JJ, Li YH, Pan J, Zhang YR. Autophagy inhibition increased the anti-tumor effect of cisplatin on drug-resistant esophageal cancer cells. *J Biol Regul Homeost Agents* 2017;31:645-52.
51. Wei MF, Chen MW, Chen KC, Lou PJ, Lin SY, Hung SC, Hsiao M, Yao CJ, Shieh MJ. Autophagy promotes resistance to photodynamic therapy-induced apoptosis selectively in colorectal cancer stem-like cells. *Autophagy* 2014;10:1179-92. doi: 10.4161/auto.28679.
52. Zhang Z, Wang A, Li H, Zhi H, Lu F. STAT3-dependent TXNDC17 expression mediates Taxol resistance through inducing autophagy in human colorectal cancer cells. *Gene* 2016;584:75-82. doi: 10.1016/j.gene.2016.03.012.
53. Bergers G, Hanahan D. Modes of resistance to anti-angiogenic therapy. *Nat Rev Cancer* 2008;8:592-603.
54. Tomida C, Nagano H, Yamagishi N, Uchida T, Ohno A, Hirasaka K, Nikawa T, Teshima-Kondo S. Regorafenib induces adaptive resistance of colorectal cancer cells via inhibition of vascular endothelial growth factor receptor. *J Med Invest* 2017;64:262-5. doi: 10.2152/jmi.64.262.

55. Murakami A, Takahashi F, Nurwidya F, Kobayashi I, Minakata K, Hashimoto M, Nara T, Kato M, Tajima K, Shimada N, Iwakami S, Moriyama M, Moriyama H, Koizumi F, Takahashi K. Hypoxia increases gefitinib-resistant lung cancer stem cells through the activation of insulin-like growth factor 1 receptor. *PLoS One* 2014;9:e86459.
56. Vencken SF, Sethupathy P, Blackshields G, Spillane C, Elbaruni S, Sheils O, Gallagher MF, O'Leary JJ. An integrated analysis of the SOX2 microRNA response program in human pluripotent and nullipotent stem cell lines. *BMC Genomics* 2014;15:711.
57. Abdel-Hafiz HA. Epigenetic Mechanisms of Tamoxifen Resistance in Luminal Breast Cancer. *Diseases*. 2017;5. doi: 10.3390/diseases5030016.
58. Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell* 2000;100:57-70.
59. Sharma SV, Gajowniczek P, Way IP, Lee DY, Jiang J, Yuza Y, Classon M, Haber DA, Settleman J. A common signaling cascade may underlie "addiction" to the Src, BCR-ABL, and EGF receptor oncogenes. *Cancer Cell* 2006;10:425-35.
60. Danial NN. BCL-2 family proteins: Critical checkpoints of apoptotic cell death. *Clin Cancer Res* 2007;13:7254-63.
61. Pham CG, Bubici C, Zazzeroni F, Knabb JR, Papa S, Kuntzen C, Franzoso G. Upregulation of twist-1 by NF-kappaB blocks cytotoxicity induced by chemotherapeutic drugs. *Molecul Cell Biol* 2007;27:3920-35.
62. Osford SM, Dallman CL, Johnson PW, Ganesan A, Packham G. Current strategies to target the anti-apoptotic Bcl-2 protein in cancer cells. *Curr Med Chem* 2004;11:1031-9.
63. Harrington EA, Bennett MR, Fanidi A, Evan GI. c-Myc-induced apoptosis in fibroblasts is inhibited by specific cytokines. *EMBO J*. 1994;13:3286-95.
64. Beckta JM, Ahmad SF, Yang H, Valerie K. Revisiting p53 for cancer-specific chemo- and radiotherapy: ten years after. *Cell Cycle* 2014;13:710-3.
65. Geretto M, Pulliero A, Rosano C, Zhabayeva D, Bersimbaev R, Izzotti A. Resistance to cancer chemotherapeutic drugs is determined by pivotal microRNA regulators. *Am J Cancer Res* 2017;7:1350-71.
66. Bonanno L, Favaretto A, Rosell R. Platinum drugs and DNA repair mechanism in lung cancer. *Anticancer Res* 2014;34:493-502.
67. Cieślak-Boczula K, Świętek P, Jaszczyszyn A, Zawilska P, Gąsiorowski K, Malinka W, Köhler G. Phase separation in phosphatidylcholine membrane caused by the presence of a pyrimidine analogue of fluphenazine with high anti-multidrug-resistance activity. *J Phys Chem B* 2014;118:3605-15. doi: 10.1021/jp410882r.
68. Wang W, Rastinejad F, El-Deiry WS. Restoring p53-dependent tumor suppression. *Cancer Biol Ther* 2003;2:S55-63.
69. Pluquet O, Hainaut P. Genotoxic and non-genotoxic pathways of p53 induction. *Cancer Lett* 2001;174:1-15.
70. Smith ML, Seo YR. p53 regulation of DNA excision repair pathways. *Mutagenesis* 2002;17:149-56.
71. Bode AM, Dong Z. Post-translational modification of p53 in tumorigenesis. *Nat Rev Cancer* 2004;4:793-805.
72. Honda R, Tanaka H, Yasuda H. Oncoprotein MDM2 is a ubiquitin ligase E3 for tumor suppressor p53. *FEBS Lett* 1997;420:25-7.
73. Pirollo KF, Bouker KB, Chang EH. Does p53 status influence tumor response to anticancer therapies? *Anticancer Drugs* 2000;11:419-32.
74. Chang EH, Pirollo KF, Bouker KB. Tp53 gene therapy: a key to modulating resistance to anticancer therapies? *Mol Med Today* 2000;6:358-64.
75. Riordan JR, Ling V. Purification of P-glycoprotein from plasma membrane vesicles of Chinese hamster ovary cell mutants with reduced colchicine permeability. *J Biol Chem* 1979;254:12701-5.
76. Prajapati R, Sangamwar AT. Translocation mechanism of Pglycoprotein and conformational changes occurring at drug-binding site: insights from multi-targeted molecular dynamics. *Biochim Biophys Acta* 2014;1838:2882-98.
77. Liu T, Peng Y, Li X, Liu L, Liu F, He L. A novel delocalized lipophilic cation-chlorambucil conjugate inhibits P-

- glycoprotein in HepG2/ADM cells. *Bioorg Med Chem* 2017. doi: 10.1016/j.bmc.2017.08.003.
78. Kruh GD, Belinsky MG. The MRP family of drug efflux pumps. *Oncogene* 2003;22:7537-52.
 79. Stavrovskaya AA. Cellular mechanisms of multidrug resistance of tumor cells. *Biochem* 2000;65:95-106.
 80. Ozben T. Mechanisms and strategies to overcome multiple drug resistance in cancer. *FEBS Lett* 2006;580:2903-9.
 81. Szaka'cs G, Paterson JK, Ludwig JA, Booth-Genthe C, Gottesman MM. Targeting multidrug resistance in cancer. *Nat Rev* 2006;3:219-34.
 82. Borst P, Evers R, Kool M, Wijnholds J. A family of drug transporters: the multidrug resistance-associated proteins. *J Natl Cancer Inst* 2000;92:1295-301.
 83. Ni Z, Bikadi Z, Rosenberg MF, Mao Q. Structure and function of the human breast cancer resistance protein (BCRP/ABCG2). *Curr Drug Metab* 2010;11:603-17.
 84. Natarajan K, Xie Y, Baer RMT, Ross DR. Role of breast cancer resistance protein (BCRP/ABCG2) in cancer drug resistance. *Biochem Pharmacol* 2012;83:1084-103.
 85. Chauhan PS, Bhushan B, Singh LC, Mishra AK, Saluja S, Mittal V, Gupta DK, Kapur S. Expression of genes related to multiple drug resistance and apoptosis in acute leukemia: response to induction chemotherapy. *Exp Mol Pathol* 2012;92:44-9.
 86. Zhang D, Jia J, Zhao G, Yue M, Yang H, Wang J. NDRG1 promotes the multidrug resistance of neuroblastoma cells with upregulated expression of drug resistant proteins. *Biomed Pharmacother* 2015;76:46-51. doi: 10.1016/j.biopha.2015.10.015.
 87. Sordillo LA, Sordillo PP, Helson L. Sphingosine Kinase Inhibitors as Maintenance Therapy of Glioblastoma After Ceramide-Induced Response. *Anticancer Res* 2016;36:2085-95.
 88. Pchejetski D, Golzio M, Bonhoure E, Calvet C, Doumerc N, Garcia V, Mazerolles C, Rischmann P, Teissié J, Malavaud B, Cuvillier O. Sphingosine kinase-1 as a chemotherapy sensor in prostate adenocarcinoma cell and mouse models. *Cancer Res* 2005;65:11667-75.
 89. Ponnusamy S, Meyers-Needham M, Senkal CE, Saddoughi SA, Sentelle D, Selvam SP, Salas A, Ogretmen B. Sphingolipids and cancer: ceramide and sphingosine-1-phosphate in the regulation of cell death and drug resistance. *Future Oncol* 2010;6:1603-24.
 90. Gouaze' V, Liu YY, Prickett CS, Yu JY, Giuliano AE, Cabot MC. Glucosylceramide synthase blockade down-regulates P-glycoprotein and resensitizes multidrugresistant breast cancer cells to anticancer drugs. *Cancer Res* 2005;65:3861-7.
 91. Liu YY, Hill RA, Li YT. Ceramide glycosylation catalyzed by glucosylceramide synthase and cancer drug resistance. *Adv Cancer Res* 2013;117:59-89. doi: 10.1016/B978-0-12-394274-6.00003-0.
 92. Turáková K, Pavlíková L, Messingerová L, Lakatoš B, Breier A, Sulová Z. Reduced UDP-glucose Levels Are Associated with P-glycoprotein Over-expression in L1210 Cells and Limit Glucosylceramide Synthase Activity. *Anticancer Res* 2015;35:2627-34.
 93. Chiu WH, Su WC, Li CL, Chen CL, Lin CF. An increase in glucosylceramide synthase induces Bcl-xL-mediated cell survival in vinorelbine-resistant lung adenocarcinoma cells. *Oncotarget* 2015;6:20513-24.
 94. Reed JC. Bcl-2: prevention of apoptosis as a mechanism of drug resistance. *Hematol Oncol Clin North Am* 1995;9:451-73.
 95. Jia Y, Chen J, Zhu H, Jia ZH, Cui MH. Aberrantly elevated redox sensing factor Nrf2 promotes cancer stem cell survival via enhanced transcriptional regulation of ABCG2 and Bcl-2/Bmi-1 genes. *Oncol Rep* 2015;34:2296-304. doi: 10.3892/or.2015.4214.
 96. Stambolic V, Suzuki A, de la Pompa JL, Brothers GM, Mirtsos C, Sasaki T, Ruland J, Penninger JM, Siderovski DP, Mak TW. Negative regulation of PKB/Akt-dependent cell survival by the tumor suppressor PTEN. *Cell* 1998;95:29-39.
 97. Uygur B, Abramo K, Leikina E, Vary C, Liaw L, Wu WS. SLUG is a direct transcriptional repressor of PTEN tumor suppressor. *Prostate* 2015;75:907-16. doi: 10.1002/pros.22974.

98. Friedman R. Drug resistance missense mutations in cancer are subject to evolutionary constraint. *PLoS One* 2013;8:e82059.
99. Islam MT. Crucial challenges in epigenetic cancer therapeutic strategy yet to be resolved. *International Journal of Pharmacy and Pharmaceutical Sciences* 2016; 8: 1-6. doi: 10.22159/ijpps.2016v8i12.14510.
100. Plumb JA, Strathdee G, Sludden J, Kaye SB, Brown R. Reversal of drug resistance in human tumor xenografts by 2'-deoxy-5-azacytidine-induced demethylation of the hMLH1 gene promoter. *Cancer Res* 2000; 60:6039-44.
101. Bearzatto A, Szadkowski M, Macpherson P, Jiricny J, Karran P. Epigenetic regulation of the MGMT and hMSH6 DNA repair genes in cells resistant to methylating agents. *Cancer Res* 2000;60:3262-70.
102. Almendro V, Marusyk A, Polyak K. Cellular heterogeneity and molecular evolution in cancer. *Annu Rev Pathol* 2013;8:277-302.
103. Meacham CE, Morrison SJ. Tumour heterogeneity and cancer cell plasticity. *Nature* 2013;501:328-37.
104. Clevers H. The cancer stem cell: premises, promises and challenges. *Nat Med* 2011;17:313-9.
105. Kreso A, Dick JE. Evolution of the cancer stem cell model. *Cell Stem Cell* 2014;14:275-91.
106. McDermott KM, Zhang J, Holst CR, Kozakiewicz BK, Singla V, Tlsty TD. p16(INK4a) prevents centrosome dysfunction and genomic instability in primary cells. *PLoS Biol* 2006;4:e51.
107. Loeb LA, Bielas JH, Beckman RA. Cancers exhibit a mutator phenotype: Clinical implications. *Cancer Res* 2008;68:3551-7.
108. Blagosklonny MV. Oncogenic resistance to growth-limiting conditions. *Nat Rev Cancer* 2002;2:221-225.
109. Dai Y, Cai X, Shi W, Bi X, Su X, Pan M, Li H, Lin H, Huang W, Qian H. Pro-apoptotic cationic host defense peptides rich in lysine or arginine to reverse drug resistance by disrupting tumor cell membrane. *Amino Acids* 2017. doi: 10.1007/s00726-017-2453-y.
110. Gillet JP, Efferth T, Remacle J. Chemotherapy-induced resistance by ATP-binding cassette transporter genes. *Biochim Biophys Acta* 2007;1775:237-62.
111. Ejendal KF, Hrycyna CA. Multidrug resistance and cancer: The role of the human ABC transporter ABCG2. *Curr Protein Peptide Sci* 2002;3:503-11.
112. Loebinger MR, Giangreco A, Groot KR, Prichard L, Allen K, Simpson C, Bazley L, Navani N, Tibrewal S, Davies D, Janes SM. Squamous cell cancers contain a side population of stem-like cells that are made chemosensitive by ABC transporter blockade. *British J Cancer* 2008;98:380-7.
113. Szakacs G, Paterson JK, Ludwig JA, Booth-Gentle C, Gottesman MM. Targeting multidrug resistance in cancer. *Nat Rev Drug Discov* 2006;5:219-34.
114. Baguley BC. Novel strategies for overcoming multidrug resistance in cancer. *BioDrugs* 2002;16:97-103.
115. Kuilman T, Michaloglou C, Vredeveld LC, Douma S, van Doorn R, Desmet CJ, Aarden LA, Mooi WJ, Peeper DS. Oncogene-induced senescence relayed by an interleukin-dependent inflammatory network. *Cell* 2008;133:1019-31.
116. Fonseca C, Dranoff G. Capitalizing on the immunogenicity of dying tumor cells. *Clin Cancer Res* 2008;14:1603-8.
117. Panaretakis T, Joza N, Modjtahedi N, Tesniere A, Vitale I, Durchschlag M, Fimia GM, Kepp O, Piacentini M, Froehlich KU, van Endert P, Zitvogel L, Madeo F, Kroemer G. The co-translocation of ERp57 and calreticulin determines the immunogenicity of cell death. *Cell Death Differentiation* 2008;15:1499-509.
118. Vahedi F, Nham T, Poznanski SM, Chew MV, Shenouda MM, Lee D, Ashkar AA. *Ex Vivo* Expanded Human NK Cells Survive and Proliferate in Humanized Mice with Autologous Human Immune Cells. *Sci Rep* 2017;7:12083. doi: 10.1038/s41598-017-12223-8.
119. Malmberg KJ, Sohlberg E, Goodridge JP, Ljunggren HG. Immune selection during tumor checkpoint inhibition therapy paves way for NK-cell "missing self" recognition. *Immunogenetics*. 2017;69:547-56. doi: 10.1007/s00251-017-1011-9.

120. Schäfer A, Köhler SC, Lohe M, Wiese M, Hiersemann M. Synthesis of Homoverrucosanoid-Derived Esters and Evaluation as MDR Modulators. *J Org Chem* 2017. doi: 10.1021/acs.joc.7b02012.
121. Maeda H. The enhanced permeability and retention (EPR) effect in tumor vasculature: the key role of tumor-selective macromolecular drug targeting. *Adv Enzyme Regul* 2001;41:189-207.
122. Palakurthi S, Yellepeddi VK, Vangara KK. Recent trends in cancer drug resistance reversal strategies using nanoparticles. *Expert Opin Drug Deliv* 2012;9:287-301.
123. Bar-Zeev M, Livney YD, Assaraf YG. Targeted nanomedicine for cancer therapeutics: Towards precision medicine overcoming drug resistance. *Drug Resist Updat* 2017;31:15-30. doi: 10.1016/j.drug.2017.05.002.
124. Maheshwari R, Tekade M, Gondaliya P, Kalia K, D'Emanuele A, Tekade RK. Recent advances in exosome-based nanovehicles as RNA interference therapeutic carriers. *Nanomedicine (Lond)* 2017. doi: 10.2217/nmm-2017-0210.
125. Aagaard L, Rossi JJ. RNAi therapeutics: principles, prospects and challenges. *Adv Drug Deliv Rev* 2007;59:75-86.
126. Chaturvedi K, Ganguly K, Kulkarni AR, Kulkarni VH, Nadagouda MN, Rudzinski WE, Aminabhavi TM. Cyclodextrin-based siRNA delivery nanocarriers: a state-of-the-art review. *Expert Opin Drug Deliv* 2011;8:1455-68.
127. Xiong XB, Uludag H, Lavasanifar A. Novel biodegradable amphiphilic poly(ethylene oxide)-block-polyesters with grafted polyamines as supramolecular nanocarrier for efficient siRNA delivery. *Biomaterials* 2009;30:242-53.
128. Susa M, Iyer AK, Ryu K, Choy E, Hornicek FJ, Mankin H, Milane L, Amiji MM, Duan Z. Inhibition of ABCB1 (MDR1) expression by an siRNA nanoparticulate delivery system to overcome drug resistance in osteosarcoma. *PLoS One* 2010;5:e10764.
129. Creixell M, Peppas NA. Co-delivery of siRNA and therapeutic agents using nanocarriers to overcome cancer resistance. *Nano Today* 2012;7:367-79.
130. Maurer BJ, Melton L, Billups C, Cabot MC, Reynolds CP. Synergistic cytotoxicity in solid tumor cell lines between N-(4-hydroxyphenyl)retinamide and modulators of ceramide metabolism. *J Natl Cancer Inst* 2000;92:1897-909.
131. Chun-Guang W, Liang Z, Yong-L L, Xue-Jun S, Long-Qin S, Li Z, Bei-Zhong L. Emodin Exerts an Antiapoptotic Effect on Human Chronic Myelocytic Leukemia K562 Cell Lines by Targeting the PTEN/PI3K-AKT Signaling Pathway and Deleting BCR-ABL. *Integr Cancer Ther* 2016. pii: 1534735416664784.
132. Patel J, Mitra AK. Strategies to overcome simultaneous P-glycoprotein mediated efflux and CYP3A4 mediated metabolism of drugs. *Pharmacogenomics* 2001;2:401-15.
133. Li J, Li ZN, Du YJ, Li XQ, Bao Q, Chen P. Expression of MRP1, BCRP, LRP, and ERCC1 in advanced non-small-cell lung cancer: correlation with response to chemotherapy and survival. *Clin Lung Cancer* 2009;10:414-21.
134. Fu Y, Liu S, Yin S, Niu W, Xiong W, Tan M, Li G, Zhou M. The reverse Warburg effect is likely to be an Achilles' heel of cancer that can be exploited for cancer therapy. *Oncotarget* 2017;8:57813-25. doi: 10.18632/oncotarget.18175.
135. Ismael GF, Rosa DD, Mano MS, Awada A. Novel cytotoxic drugs: old challenges, new solutions. *Cancer Treat Rev* 2008;34:81-91.
136. Pan C, Wang X, Shi K, Zheng Y, Li J, Chen Y, Jin L, Pan Z. MiR-122 Reverses the Doxorubicin-Resistance in Hepatocellular Carcinoma Cells through Regulating the Tumor Metabolism. *PLoS One* 2016;11:e0152090. doi: 10.1371/journal.pone.0152090.
137. Madani Tonekaboni SA, Soltan Ghorai L, Manem VS, Haibe-Kains B. Predictive approaches for drug combination discovery in cancer. *Brief Bioinform* 2016. PMID: 27881431.
138. Yang W, Ma J, Zhou W, Zhou X, Cao B, Zhang H, Zhao Q, Fan D, Hong L. Molecular mechanisms and clinical implications of miRNAs in drug resistance of esophageal cancer. *Expert Rev Gastroenterol Hepatol* 2017;30:1-13. doi: 10.1080/17474124.2017.1372189.

139. Sun G, Fan T, Zhao L, Zhou Y, Zhong R. The potential of combi-molecules with DNA-damaging function as anticancer agents. *Future Med Chem* 2017;9:403-35. doi: 10.4155/fmc-2016-0229.