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RESEARCH ARTICLE

Biopotential Features and Pesticidal Study of Cascabela peruviana, Nerium oleander, and Mimusops elengi against Armyworm Spodoptera litura (Noctuidae: Lepidoptera) and Pod Borer Larvae of Helicoverpa armigera (Noctuidae: Lepidoptera)

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ABSTRACT

To study the report, the different solvents of methanol, ethyl acetate, chloroform, and acetone for *Cascabela peruviana* (*C. peruviana*), *Nerium oleander* (*N. oleander*), and *Mimusops elengi* (*M. elengi*) were used the experimental analysis in pest control of most dangerous notorious *Lepidopteran* pests of *Spodoptera litura* (*S. litura*) and *Helicoverpa armigera* (*H. armigera*). The antifeedant activity of *C. peruviana* against *S. litura* 98.6%, *H. armigera* 94.6%, *N. oleander* against *S. litura* 94.2%, *H. armigera* 90.8% and *M. elengi* against *S. litura* 92.8%, *H. armigera* 86.4%. Lethal concentration 50 (L_{cso}) and L_{cso} 0 and L_{cso} 1 and L_{cso} 2 and L_{cso} 3 and L_{cso} 4 and L_{cso} 4 and L_{cso} 5 and L_{cso} 6 and L_{cso} 6 and L_{cso} 7 and L_{cso} 7 and L_{cso} 8 and L_{cso} 8 and L_{cso} 9 and a

Keywords: Cascabela peruviana, Helicoverpa armigera, Mimusops elengi, Nerium oleander, notorious, Spodoptera litura

INTRODUCTION

One possible way to reduce the high consumption of synthetic insecticides is through the application botanical pesticides commonly considered to be environmentally and medically safe.^[1,2] Botanical properties are highly toxic to many insect species and more than 2000 plant verities are known to possess some medical properties.^[3] Biopesticides are alternative to synthetic pesticides due to their generally low environmental pollution, low toxicity to humans, and other applications. Essential oils and their constituents have been reported to be an effective source of botanical pesticides.^[4,5]

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Cascabela peruviana, family: Apocynaceae, is a most toxicant plant motherland of Southern Mexico and Central America plantation as an ornamental. C. peruviana is commonly used as domestic medicine in Tropical America and also in Tropical Asia. These toxic substances have experiments for uses in the biopest control. C. peruviana seed oil was used to antifungal. antibacterial, and anti-termite products. [6] The clinical advantage of antidotes, atropine, and digitoxin antibodies and treatment include oral management activities of charcoal.^[7] C. peruviana Plant materials reduced the toxic fragment that produces antibody of ovine polyclonal as used for many countries.^[8] The plant latex is applied to decayed teeth to recover toothache, it is used to treat chronic sores and ulcers, and it is applied to soften corns and calluses. The chemical

constituents of flavonol glycoside have been isolate from the *C. peruviana* leaves. Particular chemical constituents found to the inhibitory effects against HIV-1 reverse transcriptase HIV-1 integrase. *Nerium oleander*, family Apocynaceae, is an evergreen shrub growing up to 4 m in height. It is one of the ornamental plants distributed in Tropical Asia. Oleander is one of the most toxicness plants in the world and contains numeric toxic contents, many of which can be died human beings, especially young children. *N. oleander* is considered extremely high and it has conclusions of some cases that only a small amount had lethal effects. The most significant of these toxins is oleandrin, which are cardiac glycosides. [9]

In addition to their antioxidant and their very marked antimicrobial activity, the polyphenols have other activities such as vasculoprotectrice activity anti-inflammatory and antitumor.[10] The aqueous extract of N. oleander L. has been undergoing clinical investigations as an anticancerous agent. Oleandrin and its aglycone oleandrigenin are the active compounds that are isolated from this plant which shown to have anticancerous properties, and the glycosides are beneficial in lowering blood pressure.[31] Anvirzel has also revealed cytotoxicity in human tumor cell lines with evidence of apoptosis as a principal mode of cell death.[11] Mimusops elengi Linn, family Sapotaceae, generally known as bakul is a small to large evergreen tree showed all over the different parts of India. M. elengi plant materials were used as anthelmintic, anti-anxiety, antihyperlipidemic, antiulcer, anti-inflammatory, analgesic, antipyretic, and anticonvulsant to be used. Natural antioxidants such as flavonoids, tannins, and phenols are increasingly attracting because they are disease preventing, health promoting, and antiaging substances.[32] The traditional medicinal valuable reported many plant parts used as cardio tonic, alexipharmic, stomachic, anthelmintic, tonic astringent which curable biliousness, disease of the gums and teeth.[33] The flower is good aroma and cooling, astringent to the bowels are used to curable disease to the blood, liver complaints, disease of the nose, headache and then curable to the asthma disease.[12] The developmental practices of traditional medicine are based on centuries of belief and observations and noted, which need help in the development of innovation medicine. In recent days, there is worldwide

interest in herbal drugs. The natural flora has been used in a number of systems of medicines in our country as well as in other countries. India is well known as the 'Emporium of Medicinal Plants'.[13] Spodoptera litura, the armyworm, is a major devastating pest which is the host varieties of 180 plant specious and 45 families of worldwide.[14,15] This pest of armyworm plays a major role in infestation of agriculture forming. It causes commercial economic losses to various vegetables and field crops from 10% to 30%.[16] The neonate larvae earlier attack the foliage of the plants and later stage feed on matured seeds in the pod. The selected pest is considered as the serious pests of various commercial important crops such as cotton, peanut, chilly, tobacco, castor bean, legumes, and teases.[17]

Helicoverpa armigera is another devastating pest of worldwide occurrence inflicting crop damage in India to the sum of one billion dollars at the end of the year attacks over 200 variety species of field crop belonging to 45 families.[18] This pest was damaged potential of average infestation single larvae can be destroyed 30-40 pods per plant in cotton the field.[19] H. armigera is a cosmopolitan insect and has gained importance as a major devastating pest due to its capacity to feed on many verities of plant species, some of which are important agricultural crops. [20] Therefore, extensive studies are carried out to screen plants as insect growth control agents. Over the past decades, better attention has been focused on the bioactivity of phytochemicals for their potential as pesticides against phytophagous insects. [21] Hence, the present study has been made on the important medicinal plant of C. peruviana, N. oleander, and M. elengi against S. litura and H. armigera to study of eco-friendly approaches of agriculture pest control.

MATERIALS AND METHODS

Collection and extraction of plant material

C. peruviana, N. oleander, and M. elengi the mature flowers are collected from Naduvalur village of Salem district, Tamil Nadu, India. The bulk plant raw material was dried in the shade at room temperature. The dried leaves 150 g were extracted with hexane, ethyl acetate, chloroform, and methanol (750 ml), in a Soxhlet apparatus individually until

exhaustion. The extract was concentrated under reduced pressure of 22–26 mm Hg at 45°C by "Rotovapor" and the residue obtained was stored at 4°C by in an amber vial. Then, the vials labeled with silver foil and transported to the laboratory.

Rearing of insects

The taro caterpillar *S. litura* and *H. armigera* were collected from the field in Kumaramangalam village, Cuddalore district of Tamil Nadu, India, and the collected larvae were reared individually in plastic container vials and fed usually peanut leaf (*Arachis hypogea*) until the larvae became pupae under the laboratory condition $(27 \pm 2^{\circ}\text{C})$ and $75 \pm 5\%$ relative humidity. Usually, hale and healthy uniform sized fourth instar larvae, the recently emerged matured eggs, and adult moths of genteel species were used in the pesticidal activity.

Antifeedant activity

Antifeedant activity of the methanol extracts is used leaf disc method. The fresh peanut (A. hypogea) plant leaf was used the experimental studies. Leaf disc of 4.0 cm diameter was punched using leaf eater and was dipped individually 25, 75, 125, 175, and 225 ppm. The leaf disc dipped in hexane, ethyl acetate, methanol, and chloroform was used to extracts. In each plastic Petri dish ($40 \text{ cm} \times 90 \text{ cm}$), wet filter paper was placed to avoid early drying of the tested leaves. The fourth instar larvae were pioneered in each and every jurisidiction Petridis. The consumption of leaf disc in the treated and control S. litura and H. armigera larvae after 48 h of the experience was measured using leaf area meter. Leaf discs consumed by the larvae in the test were corrected from the negative jurisdiction. Five replicates were maintained for each treatment with 25 larvae. The investigation was conducted at laboratory condition (27.0°C ± 2°C) with 14:10 h illumination, and dark photoperiod and $75 \pm 5\%$ relative humidity activity were calculated according to the formula.[22]

Larvicidal activity

Larvicidal activity was studied using leaf nochoice method. Peanut leaf disc was used; they were dipped in various concentrations of 25, 75, 125, 175, and 225 ppm plant extracts as used for the larvicidal activity. After 48 h experiment, the larvae *S. litura* and *H. armigera* were continuously maintained on untreated fresh peanut and castor leaves. Diet was changed every 48 h. Larval mortality was recorded up to 24 h experiment. The number of larvae 25 replicates used and laboratory conditions were the same as the percentage of larval mortality was calculated using Abbot's formula.^[23]

Mortality% =
$$\frac{\%MT - MT}{100 - \%MC} \times 100$$

Oviposition-deterrent activity

Twenty-five individual eggs of *S. litura* and *H. armigera* were separated and immersed in 25, 75, 125, 175, and 225 ppm concentrations. Five replicates were maintained (n = 100); number of eggs hatched in the control and the treatments was recorded. The laboratory conditions were the same as the antifeedant activity treatment.

$$\%OA = \frac{\%EHC - \%EHT}{\%EHC} \times 100$$

Statistical analysis

The normal mortality information was focused to probit^[24] investigation for ascertaining lethal concentration 50 (LC₅₀), LC₉₀, and Chi-square values which were figured by utilizing the product utilizing Statistical Package of the Social Sciences rendition 16.0 for windows, and the significant level was set at P < 0.05.

RESULTS

Antifeedant activity of various solvent extracts of *C. Peruviana*, *N. Oleander*, and *M. Elengi*

Antifeedant activity observed against fourth instar larvae of *S. litura* and *H. armigera* is shown in Table 1. The present study results showed that the antifeedant activity was assessed based on antifeedant index which normally indicates the decreased rate of feeding. The experiment study maintained three different most important medicinal plants and controlled the two notorious *Lepidopteran* pests. Plant extract used as both

Table 1: Antifeedant activity of C. peruviana, N. oleander, and M. elengi against S. litura and H. armigera

Plant	Pest	Solvent	Concentration % ppm						
			25	75	125	175	225		
C. peruviana	S. litura	Methanol	23.6±1.94ab	38.2±1.30°	60.2±2.48 ^d	82.4±1.81°	99.2±1.94 ^f		
		Ethyl acetate	21.4 ± 3.57^{ab}	36.4±1.51°	57.4 ± 2.07^{d}	76.2±2.68e	$97.2 \pm 2.48^{\rm f}$		
		Chloroform	19.6 ± 1.94^{ab}	33.2 ± 2.48^{bc}	54.2 ± 2.28^{cd}	72.6±1.81e	$94.4 \pm 0.89^{\rm f}$		
		Acetone	17.2±1.78 ^a	30.6 ± 1.94^{b}	52.4±1.51 ^{cd}	69.4±2.19 ^{de}	90.8 ± 2.38^{ef}		
	H. armigera	Methanol	21.4 ± 4.27^{ab}	$35.6 \pm 1.30^{\circ}$	56.8 ± 3.56^d	77.6±3.13e	$96.6 \pm 2.60^{\mathrm{f}}$		
		Ethyl acetate	18.8 ± 2.38^a	32.6 ± 2.60^{bc}	53.4 ± 2.88^{cd}	75.4±2.96e	$93.2 \pm 2.48^{\rm f}$		
		Chloroform	15.4±2.19a	30.2 ± 1.78^{b}	52.6 ± 0.89^{cd}	69.2 ± 2.86^{de}	$91.6 \pm 1.94^{\mathrm{f}}$		
		Acetone	12.8 ± 2.38^a	26.4 ± 2.50^{b}	43.2±1.64°	55.2±1.92 ^{cd}	82.2±2.04e		
N. oleander	S. litura	Methanol	22.4 ± 2.19^{ab}	37.4±2.88°	58.2 ± 2.48^d	79.6±2.88e	$96.6 \pm 2.30^{\rm f}$		
		Ethyl acetate	$20.2{\pm}1.09^{ab}$	34.6 ± 2.60^{bc}	55.4±2.19 ^{cd}	73.2±2.48e	$92.2 \pm 2.48^{\rm f}$		
		Chloroform	18.4 ± 2.19^{ab}	32.4 ± 1.67^{bc}	50.6 ± 2.60^{c}	69.8±2.19 ^{de}	87.6 ± 1.94^{ef}		
		Acetone	16.6 ± 1.94^{a}	27.4 ± 2.88^{b}	48.4±1.67°	65.4±2.19 ^{de}	$84.6 \pm 1.94^{\rm ef}$		
	H. armigera	Methanol	20.2 ± 0.83^{ab}	32.2 ± 1.78^{bc}	53.8 ± 1.64^{cd}	74.6±3.04°	$94.8 \pm 1.09^{\rm f}$		
		Ethyl acetate	17.4±2.88a	30.6 ± 1.94^{b}	50.2±2.68°	71.4±1.51e	$91.4 \pm 2.70^{\rm f}$		
		Chloroform	14.8 ± 2.38^a	28.2 ± 2.86^{b}	46.2 ± 1.09^{c}	65.8 ± 1.92^{de}	88.6 ± 1.14^{ef}		
		Acetone	11.2±3.19a	23.6 ± 1.94^{ab}	39.8±1.92°	51.6±2.50 ^{cd}	79.6±1.51e		
M. elengi	S. litura	Methanol	$21.8{\pm}1.78^{ab}$	35.8 ± 1.30^{bc}	56.6 ± 2.60^d	77.2±2.48e	$94.8 \pm 1.92^{\rm f}$		
		Ethyl acetate	19.4 ± 1.01^{ab}	31.6 ± 1.94^{b}	53.4 ± 2.07^{cd}	70.2 ± 1.64^{de}	$89.2 \pm 3.70^{\rm ef}$		
		Chloroform	17.2±1.78 ^a	30.4 ± 2.88^{b}	48.2 ± 2.48^{c}	64.6 ± 1.94^{de}	87.4 ± 2.19^{ef}		
		Acetone	15.4 ± 1.78^a	24.2 ± 2.68^{ab}	46.4 ± 2.88^{c}	62.4 ± 2.30^{de}	81.2 ± 1.14^{ef}		
	H. armigera	Methanol	18.6 ± 1.34^{a}	30.8 ± 1.64^{b}	51.6 ± 2.60^{cd}	72.6±1.94°	$92.4 \pm 1.51^{\rm f}$		
		Ethyl acetate	16.2±1.64 ^a	28.4±1.81b	48.2 ± 2.48^{c}	68.4 ± 1.14^{de}	87.2 ± 4.49^{ef}		
		Chloroform	13.8 ± 2.16^a	25.2 ± 1.92^{ab}	42.4±1.81°	66.2 ± 1.92^{de}	85.6 ± 1.94^{ef}		
		Acetone	10.4 ± 2.88^{a}	19.4 ± 2.19^{ab}	37.6±1.51°	48.4 ± 2.07^{c}	70.2 ± 2.38^{de}		

Within the column, means±SD followed by the same letter indicates different significantly (ANOVA, Tukey's HSD test, P<0.05 levels), C. peruviana: Cascabela peruviana, S. litura: Spodoptera litura, H. armigera: Helicoverpa armigera, M. elengi: Mimusops elengi, N. oleander: Nerium oleander

larvicidal activity and ovicidal activity at the same control methods plants and pests. The maximum antifeedant activities were recorded in methanol extract on C. peruviana against S. litura and H. armigera, and the values are S. litura 98.6% and H. armigera 94.6%. The antifeedant activity of N. oleander against S. litura and H. armigera, and the highest values are represented S. litura 94.2% and H. armigera 90.8%. The antifeedant activity N. oleander against S. litura and H. armigera and the highest values are noted at the values are S. litura 92.8% and H. armigera 86.4%. The maximum antifeedant activity was observed in the methanol extract of C. peruviana plant extract. The higher concentration limit such as 225 ppm followed by four solvent.

Lc₅₀ of various solvent extracts of *C. Peruviana*, *N. Oleander*, and *M. Elengi*

LC₅₀ observed against fourth instar larvae of *S. litura* and *H. armigera* is shown in Table 3.

LC₅₀ and LC₉₀ values of *C. peruviana* against *S. litura* values are LC₅₀ = 88.803 and LC₉₀ = 204.91 and *H. armigera* values are LC₅₀ = 103.19 and LC₉₀ = 232.10, respectively. LC₅₀ and LC₉₀ values of *N. oleander* against *S. litura* and *H. armigera* the values are *S. litura* (LC₅₀ = 102.10 and LC₉₀ = 228.01) and *H. armigera* (LC₅₀ = 121.55 and LC₉₀ = 254.69). LC₅₀ and LC₉₀ values of *M. elengi* against *S. litura* and *H. armigera* the values are *S. litura* (LC₅₀ = 120.55 and LC₉₀ = 250.43) and *H. armigera* (LC₅₀ = 137.60 and LC₉₀ = 279.85) [Table 2]. The maximum lethal activity was observed in the methanol extract at *C. peruviana* plant extract. The higher concentration limit such as 225 ppm followed by four solvent.

Oviposition-deterrent activity of various solvent extracts of *C. Peruviana*, *N. Oleander*, and *M. Elengi*

Oviposition-deterrent activity observed against fourth instar larvae of *S. litura* and *H. armigera* is

Table 2: LC _{so} and LC _{so} values of *C. peruviana*, *N. oleander*, and *M. elengi* against *S. litura* and *H. armigera*

Plant	Pest	Solvent I	LC ₅₀ (ppm)	95% confidence limit		LC ₉₀ (ppm)	95% confidence limit		χ*
				LCL	UCL	70	LCL	UCL	
C. peruviana	S. litura	Methanol	88.803	76.046	100.18	204.91	184.17	229.83	4.860 n.s
		Ethyl acetate	104.70	91.159	117.25	239.38	216.75	271.16	4.818 n.s
		Chloroform	124.49	110.71	138.33	272.20	244.35	312.69	5.882 n.s
		Acetone	145.06	131.06	160.55	297.86	266.03	345.00	7.286 n.s
	H. armigera	Methanol	103.19	90.075	115.31	232.10	211.05	261.20	1.877 n.s
		Ethyl acetate	122.84	109.76	135.84	261.33	236.23	296.97	1.633 n.s
		Chloroform	145.92	133.46	160.72	291.74	262.00	335.07	2.023 n.s
		Acetone	166.77	153.12	182.97	307.01	276.11	352.04	3.621 n.s
N. oleander	S. litura	Methanol	102.10	89.240	114.05	228.01	207.73	255.85	1.366 n.s
		Ethyl acetate	120.94	107.54	134.11	262.23	236.67	298.73	0.481 n.s
		Chloroform	142.75	129.35	157.28	288.33	259.06	330.92	1.951 n.s
		Acetone	164.55	150.65	180.96	309.39	277.39	356.51	2.532 n.s
	H. armigera	Methanol	121.10	108.35	133.64	254.69	231.17	287.62	1.981 n.s
		Ethyl acetate	143.66	130.16	158.33	290.47	260.84	333.69	0.511 n.s
		Chloroform	163.86	149.66	180.70	312.96	279.79	362.20	0.389 n.s
		Acetone	184.33	169.35	203.29	327.78	293.11	379.49	1.439 n.s
M. elengi	S. litura	Methanol	120.55	108.09	132.78	250.43	227.85	281.80	0.535 n.s
		Ethyl acetate	140.04	127.52	153.32	274.52	248.78	310.98	0.512 n.s
		Chloroform	163.04	149.93	178.28	299.33	270.20	341.39	1.214 n.s
		Acetone	181.72	168.36	198.09	309.52	280.14	352.00	3.153 n.s
	H. armigera	Methanol	137.60	124.45	151.43	279.85	252.36	319.38	2.720 n.s
		Ethyl acetate	159.67	145.55	176.12	310.18	277.26	359.08	1.409 n.s
		Chloroform	183.41	167.93	203.14	333.78	297.00	389.49	1.803 n.s
		Acetone	202.96	187.10	224.02	338.57	302.94	392.30	2.287 n.s

 LC_{50} =Lethal concentration that kills 50% of the exposed larvae, LC_{90} =Lethal concentration that kills 50% of the exposed larvae. LCL=Lower confidence limit; UCL=Upper confi

shown in Table 3. Methanol extract of *C. peruviana* showed that *S. litura* 90.8% and *H. armigera* 86.2%. The oviposition deterrent activity of *N. oleander* the values are *S. litura* 88.6% and *H. armigera* 82.8%. The oviposition deterrent activity *M. elengi* the values are *S. litura* 78.2% and 73.6%. The maximum lethal activity was observed in the methanol extract of *C. peruviana* plant extract. The higher concentration limit such as 225 ppm followed by four solvent.

DISCUSSION

The maximum larvicidal activity was recorded from the highest concentration of methanol extract of *Abrus precatorius* at 500 ppm, and the least larvicidal activity was recorded from the 100 ppm concentration of hexane extract. Furthermore, the larval mortality observed from the 100, 200, 300, 400, and 500 ppm concentrations extracts showed 67.414 \pm 2.26, 78.73 \pm 2.63, 95.28 \pm 2.49, 100.00 \pm 0.00, and 100.00 \pm 0.00. LC₅₀ observed against

fourth instar larvae of S. litura with various solvent extracts of hexane, diethyl ether, dichloromethane, ethyl acetate, and methanol extract of A. precatorius was 255.91, 266.21, 265.98, 251.84, and 225.76 ppm, respectively. The methanol extract was responsible for strong lethal activity observed against selected pest species. [25] Asia has abundant species of medicinal and aromatic plants and traditional medicines have practiced in Asia since ancient times. India has made use of medicinal plants to cure ailments for thousands of years.[34] The results of the antifeedant potential of the solvent crude extracts of Duranta erecta were investigated against S. litura and H. armigera larvae. Antifeedant activity was assessed based on antifeedant index. Higher antifeedant index normally indicates decreased rate of feeding. In the current year report, irrespective of concentration and solvents used for extraction, the antifeedant activity varied significantly. Data pertaining to the above experiment clearly revealed that maximum antifeedant activity was recorded in ethyl acetate

Table 3: Oviposition deterrent activity of *C. peruviana*, *N. oleander* and *M. elengi* against *S. litura* and *H. armigera*

Plant	Pest	Solvent	Concentration ppm						
			25	75	125	175	225		
C. peruviana	S. litura	Methanol	19.2±2.48ab	38.4±1.14°	59.4±2.07 ^d	76.6±1.51°	98.8±1.92 ^f		
		Ethyl acetate	17.8 ± 2.07^a	35.2 ± 1.78^{bc}	57.2 ± 2.16^d	74.4±1.67e	$95.2 \pm 2.04^{\rm f}$		
		Chloroform	15.6 ± 1.14^{a}	32.8 ± 1.64^{bc}	54.6 ± 1.34^{cd}	71.8±1.92°	$93.4 \pm 1.51^{\rm f}$		
		Acetone	13.2 ± 1.30^a	30.2 ± 1.48^{b}	51.2 ± 0.83^{cd}	69.4 ± 1.81^{de}	$91.2 \pm 2.16^{\rm f}$		
	H. armigera	Methanol	16.4 ± 2.19^{a}	35.6 ± 0.89^{bc}	56.4 ± 2.30^d	73.2±0.83°	$96.2 \pm 1.64^{\rm f}$		
		Ethyl acetate	14.4 ± 2.16^{a}	33.2 ± 1.78^{bc}	54.4 ± 1.64^{cd}	71.6±1.34e	92.4±1.51 ^f		
		Chloroform	12.8 ± 2.58^a	29.6 ± 1.94^{b}	52.2 ± 2.61^{cd}	68.8 ± 1.64^{de}	90.2 ± 2.04^{ef}		
		Acetone	10.2 ± 1.30^a	$24.6{\pm}0.86^{ab}$	45.6±1.94°	63.4 ± 1.51^{de}	83.6±1.94°		
N. oleander	S. litura	Methanol	18.2 ± 2.68^{ab}	36.2±2.61°	56.6 ± 1.51^d	73.2±1.41°	$96.6 \pm 1.34^{\rm f}$		
		Ethyl acetate	16.8±1.81a	33.6 ± 1.51^{bc}	54.4 ± 2.30^{cd}	70.4 ± 1.81^{de}	$91.2 \pm 2.28^{\rm f}$		
		Chloroform	14.2±1.09a	30.4 ± 2.96^{b}	51.2 ± 2.48^{cd}	68.2 ± 2.61^{de}	89.4 ± 1.51^{ef}		
		Acetone	12.2 ± 1.14^a	27.2 ± 1.30^{b}	47.6±2.07°	65.4 ± 1.14^{de}	86.2 ± 2.38^{ef}		
	H. armigera	Methanol	15.8 ± 1.30^a	32.6 ± 1.14^{bc}	53.2 ± 1.09^{cd}	71.6±1.51°	$93.8 \pm 1.92^{\rm f}$		
		Ethyl acetate	13.6±1.51a	30.4 ± 1.18^{b}	51.4±3.04°	69.4 ± 1.51^{de}	88.4 ± 0.89^{ef}		
		Chloroform	11.6±1.31ª	27.2 ± 1.92^{b}	49.6±1.34°	65.6 ± 2.30^{de}	86.8 ± 0.86^{ef}		
		Acetone	9.4±1.14 ^a	21.4 ± 2.19^{ab}	41.2±0.83°	60.8 ± 1.92^d	80.8 ± 2.48^{e}		
M. elengi	S. litura	Methanol	17.2±2.58 ^a	33.4 ± 0.54^{bc}	52.8 ± 1.92^{cd}	71.2±2.68e	$93.2 \pm 2.86^{\rm f}$		
		Ethyl acetate	15.2±2.28a	31.2 ± 1.64^{b}	49.4±1.81°	$67.6{\pm}2.07^{\text{de}}$	86.6 ± 1.94^{ef}		
		Chloroform	13.6±2.40a	$26.8{\pm}2.28^{\rm bc}$	45.6 ± 0.89^{c}	64.4 ± 1.67^{de}	84.6±1.51e		
		Acetone	11.2±1.92ª	23.4 ± 0.89^{ab}	42.2±1.64°	62.2 ± 2.94^d	81.2±1.48e		
	H. armigera	Methanol	14.8 ± 1.64^a	30.6 ± 1.51^{b}	50.6±1.94°	68.6 ± 1.81^{de}	89.6 ± 2.40^{ef}		
		Ethyl acetate	12.8 ± 2.28^a	27.2 ± 1.78^{b}	48.2 ± 2.16^{c}	64.2 ± 1.78^{de}	85.4 ± 2.70^{ef}		
		Chloroform	10.8 ± 1.92^a	23.4 ± 1.51^{ab}	44.2±1.64°	61.4 ± 2.07^d	82.4±1.81e		
		Acetone	8.2±0.44 ^a	18.6 ± 1.94^{ab}	36.4±1.81°	58.8 ± 2.58^{d}	76.8±1.64e		

Values are expressed as mean±S.D of five replications. Within each row, different letters indicate significant differences (ANOVA, Tukey's HSD test, P<0.05),

C. peruviana: Cascabela peruviana, S. litura: Spodoptera litura, H. armigera: Helicoverpa armigera, M. elengi: Mimusops elengi, H. armigera: Helicoverpa armigera,

N. oleander: Nerium oleander

extract on S. litura 80.37% and H. armigera 78.18% at 5% concentration; high larval mortality normally indicates potential larvicidal activity of plant extracts. Irrespective of concentration and solvents used for extraction, the insecticidal activity varied significantly. Insecticidal activity data revealed clearly that maximum insecticidal activity was recorded in ethyl acetate extract on S. litura 69.88% and H. armigera 63.2%, followed by chloroform extract and petroleum ether extract at the same concentration. One-way analysis of variance followed by least significant difference test showed statistical significance P < 0.05. [26] The previous study reported that methanol extract of Rivina humilis showed $22.55 \pm 1.52\%$ feeding deterrency against the fourth instar larvae of S. litura at 100 mg/L concentration, whereas $36.49 \pm 1.68\%$ and $71.9 \pm 2.67\%$ of antifeedant activity were recorded in the methanol extract of R. humilis at 200 and 300 mg/L, respectively. Similarly, at higher concentrations such as 400 and 500 mg/L, $86.57 \pm 3.80\%$ and $98.54 \pm 3.24\%$

antifeedant activities were recorded, respectively. The maximum larvicidal activity was recorded from the highest concentration of methanol extract at 1000 mg/L and the least larvicidal activity was recorded from the 125 mg/L concentration of benzene extract. Furthermore, the larval mortality observed from the 250, 500 750, and 1000 mg/L concentration extracts showed $34.20 \pm 1.94\%$, $62.80 \pm 2.35\%$, $76.60 \pm 2.64\%$, and $98.60 \pm 2.22\%$, respectively with methanol extract.[17] The previous experiment reported that the toxicity of different extracts of Tinospora crispa and Psidium guajava was tested against S. litura. The antifeedant activity of T. crispa and P. guajava tested against S. litura fourth instar larvae was liable during a 24-h test period. All extracts are showed moderate antifeedant activity; however, very least antifeedant activity was noted in benzene and significant antifeedant activity was observed in methanol extract. Methanol extract of T. crispa and P. guajava showed 100% and 98.38% feeding deterrency against the fourth instar larvae

of S. litura at 500 ppm concentration value of benzene, diethyl ether, ethyl acetate, and methanol leaf extracts of *T. crispa* which were 92.64, 96.25, 94.67, and 84.94 ppm, respectively, and P. guajava shows the LC₅₀ values of 144.95, 164.22, 135.64, and 121.86ppm, respectively. The Chi-square values are statistically significant at P = 0.05level. Among five solvent extracts, the methanol extract was responsible for vigorous lethal activity observed against selected pest species. [27] The report showed that feeding deterrent activity of solvent extracts of Caesalpinia bonducella methanol extract of C. bonducella showed $19.67 \pm 1.93\%$ feeding deterrency against the fourth instar larvae of *H. armigera* at 100mg/l concentration, whereas $29.3 \pm 1.53\%$ and $78.52 \pm 2.86\%$ of antifeedent activity were recorded in the methanol extract of C. bonducella at 200 and 300 mg/l, respectively. Similarly, at higher concentration such as 400 and 500 mg/l, $86.87 \pm 2.82\%$ and $96.73 \pm 2.36\%$ antifeedant activities were recorded, respectively. The maximum larvicidal activity was recorded from the highest concentration of methanol extract at 1000 mg/l and the least larvicidal activity was recorded from 125 mg/l concentration of ethyl acetate extract. Furthermore, the larval mortality of the methanol extract at the concentration of 250, 500, 750, and 1000 mg/l was $29.6 \pm 1.69\%$, $57.4 \pm 1.94\%$, $81.2 \pm 1.48\%$, and $98.4 \pm 1.55\%$ respectively.[28]

The maximum results showed on the antifeedant activity of flindersine against *H. armigera* and *S.* litura. The maximum antifeedant activity of 84.24 and 78.07% was noted at 1000 ppm concentrations, respectively. All the experiment concentrations showed >50% antifeedant activity against both insects. Flindesines showed concentrationdependent activity against dual pests. LC50 value of hexane, diethyl ether, dichloromethane, ethyl acetate, and methanol extract of A. precatorius was 255.91, 266.21, 265.98, 251.84, and 225.76 ppm, respectively, against fourth instar larvae of S. litura. [32] Report the flindersines the maximum larvicidal activity of 79.11 and 69.33% at 1000 ppm concentration against H. armigera and S. litura respectively. At 125 ppm concentration, flindersine exhibited 24.88 and 22.44% larvicidal activity against H. armigera and S. litura, respectively. Flindesine showed different kinds of growth inhibitory activities against H. armigera and S. litura. It exhibited maximum larval duration of 14.66 and 15.10 days for H. armigera and S. litura, respectively, at 1000 ppm concentration. At 125 ppm concentration, it also significantly increased the larval duration to 10.87 and 10.49 for *H. armigera* and *S. litura*, respectively.^[29] The results showed that leaf extract of Catharanthus roseus was less effective in inhibiting feeding in fourth instar larvae of S. litura, both in choice and no-choice conditions, when compared to Ocimum sanctum. In choice experiment, feeding was 45.06% at the highest concentration of 5% extract differentiate to 82.47% in control. Antifeedant index was highest in 48.42% at 5% extract and lowest in 12.84% at 1%. At 2, 3, and 4% extract, antifeedant indices were 18.76, 28.44, and 31.06%, respectively.[30]

CONCLUSION

It is proof that methanol extract of *C. peruviana*, *N. oleander*, and *M. elengi* plant flower extracts was most effective insecticidal activity of armyworm *S. litura* and pod borer larvae of *H. armigera*. The flower extract experimented in the present study showed the oviposition-deterrent activity. The three different plants are useful to the active principles for the management of field crop pest and protection of agricultural ecosystem.

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