Available Online at www.ijpba.info.



International Journal of Pharmaceutical & Biological Archives 2011; 2(2): 669-672

# **ORIGINAL RESEARCH ARTICLE**

# Changes In Protein Content In The Muscle Of *Mystus cavasius* (Ham) Exposed Electroplating Industrial Effluent Chromium

P Palanisamy\*, G Sasikala, D Mallikaraj, N Bhuvaneshwari and GM Natarajan

\*PG and Research Department, Department of Zoology, Government Arts College, Coimbatore, Tamil Nadu, India

Received 20 Jan 2011; Revised 13 Feb 2011; Accepted 10 Mar 2011;

#### ABSTRACT

Protein content in the muscle of *Mystus cavasius* (Ham) exposed to 1/3 of sublethal concentration of electroplating industrial effluent chromium (0.25%) for period of 30 days has been studied. After 24, 48, 72 hrs and 15 days of exposure the muscle showed significant depletion from -9.77%, -20.45%, -26.14% and -31.82% over the control.

Key words: Electroplating industry effluent, chromium, protein, Mystus cavasius, muscle

## INTRODUCTION

In India chromium is widely used in all electroplating industries. Potassium dichromate is an oxidizing agent in chrome tanning industries. Vast quantities of effluent containing chromium are dumped in inland water bodies. The chromium which is used in chrome tanneries is potential pollutant to Indian freshwater bodies for fish culture and public health. Natural water receives Cr from anthropogenic sources such as industrial effluent derived from the production of corrosion inhibitor and pigments (Galvin 1996), which then become a pollutant of aquatic ecosystem and thus harmful to aquatic organisms (Srivastava and Singh 1981). The toxicity of Cr is affected by species, body sizes and life stage of the organisms as well as the pH of the water and to a lesser extent, by harness, salinity and temperature (Holdway 1988).

The protein content in the tissues of animals plays a role in the metabolism of animals (Palanivelu *et al* 2005). Morthy and Priyamvada (1982) stated that the protein content of the cell may be considered as an important tool for evaluation of physiological standards. The soluble protein fraction represents the activity level of enzymes in general. The structural protein fraction forms the structural moiety of a cell (Lehninger 1978). Begam and Vijayaraghavan (1996) observed protein depletion in the fish indicates the physiological strategy in order to meet the energy demand and to adapt itself to the changed metabolic system which may lead to the degradative stimulation of processes like proteolysis and utilization of degraded products for increased energy metabolism. In general, organophosphorus and organochlorine pesticides are known to depress blood protein in fishes (Grant and Mehrle 1973; Mukhopadhyay and Dehadrai 1980). Depletion of tissue protein in fishes exposed to various pesticides toxicant has been reported by many workers (Eisler and Edmunds 1996; Mehrle et al 1971 and Kabar et al 1978; Mukhopadhy and Dehadrai 1980). Further it has been reported that acute or chronic treatment of pesticide cause biochemical alterations in the organs involved in detoxification mechanisms (Dishit et al 1975; Sastry and Sharma 1979; Avan Maruthi

*et al* 2000; Shobana Rani *et al* 2000 and Prabhakar *et al* 2002). The aim of present study has been to get information about disturbance in protein metabolism due to electroplating industrial effluent chromium exposed air-breathing cat fish *Mystus cavasius*.

## MATERIALS AND METHODS

Healthy adult fish *Mystus cavasius* (14 - 22 gm weight and 12 - 18 cm length) were and acclimated to the laboratory conditions with softened tap water under the following conditions: Ca, 0.725 mm; Mg, 0.135 mm; pH 7.1  $\pm$  0.4; D.O, 7.4  $\pm$  0.2 mg/ l. Water was checked daily for NH<sub>3</sub>, nitrite, and nitrate and replaced every 2 days for half of the volume. Water was filtered with a trickling filter and biological kits were used for NH<sub>3</sub>, nitrite and nitrate measurements to ensure that levels never exceeded 0.1. 1 and 20mg  $1^{-1}$ . respectively. The percentage survival of M.cavasius at various concentrations of waste water was determined by adopting the procedure laid down by Doudoroff and Katz (1953) for industrial waste waters. The test medium was changed daily (Sprague, 1971) to maintain the constant toxic concentration. The  $LC_{50}$  value was obtained by Finney (1971). The 1/3 of sublethal concentration of chromium (0.25%) was used to experimental fishes for 24, 48, 72 hours and 15 days. At these concentrations, the fishes survived indefinitely without any ill effect. The Protein content estimated by Lowry et al., (1951) and statistical significance of difference between control and treated groups of different exposure period were tested by using't' test (Zar 1984).

# **RESULT AND DISCUSSION**

Alteration in physiological and biochemical parameters of toxicant treated fish has recently emerged as an important tool for the water quality assessment in pathological studies of fish in the field of environmental toxicology (Racicot et al 1975: Wieser and Hinterleitner 1980 and Kulshrestha et al 1995). Protein as one of the main sources of energy and it plays an important role in the maintenance of blood glucose (Jrueger et al 1968). It is the most fundamental and abundant biochemical constituent present in the animal body and the estimation of protein is considered to be important (Ravichandran et al 1994). Mule and Lomte (1995) have reported that the protein content of an animal is an important organic constituent, which plays a major role in cellular metabolism. After the exposure of electroplating effluent chromium protein content in the muscle of *M.cavasius* was significantly

affected. After 24, 48, 72 hr and 15 days of exposure the muscle showed constant depletion in amount protein from -9.77%. the of -20.45%, -26.14% and -31.82% over control. Gradual decrease of protein 24, 48, 72 hr and 7<sup>th</sup> day of exposure may be due to the influence of exogenous factors like toxic environment as has been suggested by Castell et al (1970). The loss of protein under effluent chromium to long period may attribute to the utilization of amino acid in various catabolic reactions. Another probability is that might have occurred blocking of the protein synthesis and proteolysis on exposure to chronic period of stress condition. This is in support with studies if Srinivas Rao (1987). Similar observations was made by Jana and Bandyo Padhay (1987) who noticed the reduction of total protein in the muscle of tissue C. punctatus after the treatment of heavy metals. Khillare and Wash (1989) recorded protein declined in muscle of Puntius stigma after exposure of endosulfan, malathion and sevin. Ravichandran et al (1994) observed the effluent of phenol, found decreased the protein in muscle Oreochromis mossambicus. Lomte and Sabhia Alam (1984) studied effect of Malathion on the biochemical components of prosobranch, Belamia bengalensis and reported that the decrease in glycogen, protein and lipid under pesticidal stress. Rajan (1990) reported the effect of textile mill effluent on Cyprinus carpio. There was a significant decrease in protein content of muscle, liver and intestine. Govindan et al (1994) observed the decrease protein in the of Gambusai affinis muscle exposed to Phosphoridon. Jone Nelson and Sunil kumar (1996) reported the decrease level of protein in the muscle of Etroplus maculates after exposure of Ekalux. Vijaya mohan (2000) noticed the decreased protein content of muscle in Orechromis mossambicus after the exposure of titanium dioxide.

 Table 14. Electro plating industrial effluent chromium (0.25 %) change the protein content in the Air-breathing fish

 Mystus cavasius (Ham)

Muscle	$8.80 \pm 1.71$	$7.94 \pm 1.60^{*}$ -9.77	$7.00 \pm 1.53^{*}$ -20.45	$6.50 \pm 1.08^{**}26.14$	$6.00 \pm 1.15^{**}\text{-}31.82$

**48 hrs** 

Each value is the mean of 6 individual determinations  $\pm$  indicates SE The signs + or – indicate percent increase or decrease over control Values significant at \*P<0.05, \*\*P<0.01,

24 hrs

#### References

Samples

- 1. Galvin RM. 1996.Occurrence of metals in water. An overview. *Water Sa.*, 22(1) : 7-18.
- 2. Srivastava AK and Singh NH. 1981. Effect of acute exposure of methyl parathion on carbohydrate metabolism of Indian catfish,

15 days

72 hrs

Controls

*Heteropneustes fossilis. Acta. Pharmacol. Toxicol.* **48** : pp 26.

- Holdway DA. 1988. The toxicity of chromium to fish. In : *Chromium in the Natural and Human Environments*. JO Nriagu and Niboer (eds). John Wiley and Sons. Inc., New York, USA. pp 369 – 397.
- 4. Palanivelu V, Vijayavel K, S Balasubramanian and MP 2005. Balasubramanian. Influence of insecticidal derivative (Cartap *hydrochloride*) from the marine polycheate on certain enzyme systems of the fresh water fish Oreochromis mossambicus. J. Environ. Biol., 26:191-195.
- Morthy AS and Priyamvada D. 1982. The effects of endosulfan and its isomers on tissue protein glycogen and lipids in the fish Channa punctatus. J. Pestic. Biochem. Physicl. 17: 280 286.
- 6. Lehninger AL. 1978. Biochemistry. Kalyani Publications, Ludhiana, India, 223 – 236.
- Begam G and Vijayaraghavan S. 1996. Alterations in protein metabolism of muscle tissue in the fish *Clarias batrachus* (Linn.) by commercial grade Dimethoate. *Bull. Environ. Contam. Toxicol.*, 57: 223-228.
- 8. Grant BF and Mehrle PM. 1973. Endrin toxicosis in rainbow trout (Salmo gairdneri). J. Fish. Res. Board. Can., 30, 31 – 40.
- Mukhopadhyay PK and Dehadrai, TV. 1980. Studies on air-breathing catfish *Clarias batrachus* (Linn.) under sublethal malathion exposure. *Ind. J. Exp. Biol.*, 18: 348 – 352.
- Eisler R and Edmunds PH. 1966. Effects of endrin on blood and tissue chemistry of a marine fish. *Trans. Amer. Fish. Soc.*, 95: 153 – 159.
- Merhrk PM Stalling DL and Bloomfiled RA. 1971. Serum amino acid in rainbow trout *Salmo gairdneri* as affected by DDT and aldrin. *Com. Biochem. Physiol.*, 38B: 373 – 377pp.
- Kabeer Ahammad Sahib I, Sambasiva Rao KRS and Ramana Rao KV. 1978. Effect of malathion on free amino acids to the protein, glycogen and some enzymes of *Pelypod Lamellidens marginalis* (Lamarck). *Proc. Ind. Acad. Sci.*, **87(12)**: 377 – 387.

- Dikshit TSS, Behari JF, Datta, FK and Mathur AK. 1975. Effects of diazinon in male rats "Histopathological and biochemical studies". *Environ. Physiol. Biochem.*, 51 : 293 – 299.
- Sastry KV and Sharma SK. 1979. Toxic effects of endrin on liver and kidney of a teleost fish. *Proc. Sym. Environ. Biol.*, 339 342.
- 15. Avsan Maruthi Y and Subha Rao MV.
  2000. Effects of distillery effluents on biochemical parameters of fish *Channa punctatus* (Bloch). *J. Environ. Poll.*, 12: 111 112.
- 16. Shobana Rani D, Sudhar san R, Reddy, TN, Reddy PV and Raju TN. 2001. Effects of arsenic on certain aspects of protein metabolism in fresh water teleost *Tilapia mossambica* (Peters). *J. Environ. Biol.*, 22(2): 101 – 104.
- 17. Prabhakara Rao and Prasad Rao PV. 2002. Pollution potential of sago industry. A case study. *J. Ecotoxicol. Environ. Monit.*, **12** (1): 53 – 56.
- Doudorff P and Katz M. 1953. A critical review of literature on the toxicity of industrial wastes and their components to fish. II. The metals as salts. II. Sewage Ind Waste. 25: 802 – 839.
- Sprague JB. 1971. Measurement of pollutant toxicity of fish – III. Sub-lethal effects and safe concentrations. *Water*. *Res.*, 5: 245 – 266.
- 20. Finney DJ. 1971. Probit Analysis, Cambridge University, 3<sup>rd</sup> Edition.
- Lowry OH, Rosebrough NJ, Farr AL and Randall, RJ. 1951. Protein measurement with folin phenol reagent. *J. Bio. Chem.*, 195: 265 – 275pp.
- 22. Zar JH. 1984. *Biostatistical Analysis* (2<sup>nd</sup> Ed.) Prentice Hall Inc., *New Jersey*, U.S.A.
- 23. Racicot CG, Gaudet M and Leray C. 1975. Blood and liver enzymes in raibow trout (Salmo gairdneri Rich) with emphasis on their diagnostic use study of CCl4 toxicity and a case of Aeromonas infection. J. Fish. Biol., 7: 825 – 835.
- 24. Wiser W and Hinterleitner, S. 1980. Serum enzymes in rainbow trout as tools in the diagnosis or water quality. *Bull. Environ. Contam. Toxicol.*, 25 : 188 193.
- 25. Kulshrestha UC. Sarkar AK Srivastava SS and Parashar DC. Wet-only and bulk deposition studies at New Delhi (India)...

Palanisamy *et al.* / Changes In Protein Content In The Muscle Of *Mystus Cavasius* (Ham) Exposed Electroplating Industrial Effluent Chromium

Water, Air, & Soil Pollution, 85(4) : 2137 – 2142.

- Jrueger HM, Saddler JB, Chapman GA, Tinesly, IJ and Lowry RR. 1968.
   Bioenergetics exercise and fatty acids of fish. *Am. Zool.*, 8: 19.
- 27. Ravichandran. 1994. Impact of phenol metabolism in the freshwater fish Oreochromis mossambicus. J. Ecotoxicol. Environ. Monit., 4(1): 33 37.
- 28. Mule and Lomte, 1995.
- 29. Castell CN, Smith B and Neal W. 1970. Effects of transition metal ions on the extractable protein of fish muscle. *J. Fish. Res. Bd. Can.*, **27** : 701 – 714.
- 30. Srinivasa Rao, C. 1987. Metabolic modulation of selected biochemical parameters in the fish *Tilapia mossambica* during ammonia sulphate exposure. M.Phil. dissertation submitted to S.U. University, Tirupathi. India.
- 31. Jana S and Bandyopadhyay. 1987. Effect of heavy metals on some biochemical parameters in the freshwater fish *Channa punctatus*. *Environ*. 5(3): 488 – 493.

- Khillare YK and Wash SB. 1989. Effects of endosulfan, malathion and sevein on biochemical constituents of freshwater *Puntius stigma. Environ. Ecology.* 7(1): 66 69.
- 33. Lomte VS and Sabiha Alam. 1984. Changes in the biochemical components of prosobranch, *Belmia bengalensis* on exposure to Malathion. *Proc. Att. India. Ymp.*, 69 – 72.
- 34. Rajan, MR. 1990. Sublethal effects of textile mill effluents on protein, carbohydrate and lipid content of different tissue of fish *Cyprinus carpio*. *Environ*. *Ecol.*, 8(1): 54 – 58.
- 35. Govindan VS, Jacob L and Devika R. 1994. Toxicity and metabolic changes in *Gambusia affinis* exposed to Phosphamidon. *Toxicol. Environ. Monit.*, 4(1): 001 – 006.
- 36. Jones Nelson D and Sunil Kumar G. 1996. Effect of Ekalux on biochemical parameters in the freshwater fish *Etrophes malulatus*. J. Ecotoxical. Environ. Monit., 6(1): 65 – 67.