

REVIEW ARTICLE

A₁ Adenosine Receptor Signaling and Therapeutic Target in Diabetes

Hiba Shabir, L.A. Khan, Shamama Nishat, HR. Ansari*

Dept. of Biosciences, Jamia Millia Islamia (Central University), New Delhi-110025, India

Received 23 May 2011; Revised 07 Aug 2011; Accepted 11 Aug 2011

ABSTRACT

Diabetes is one of the risk factors to human health which progressively leads to cardiovascular complications such as ischemic heart disease, renal nephropathy, hypertension, endothelial dysfunction, and atherosclerosis. The biochemical and morphological abnormalities in various animal models has been reported in the literatures. These changes may be attributed to altered action of adenosine receptors and these receptors are named as A₁, A_{2A}, A_{2B} and A₃, mediate their effector functions through a G-protein signalling. Among these A₁AR, couples to adenylate cyclase through Gi-protein and leads to vasoconstriction and fatty acid metabolism. Endothelial dysfunction has been known to be one of the factors being responsible for pathogenesis of vascular disease in diabetes. This review will give a general overview of the adenosine receptor and focuses on the role of A₁AR in diabetes. The insight into the signaling pathway through A₁AR could be helpful in developing a novel therapeutic tool to regulate the pathophysiological conditions that arises progressively in diabetes.

Keywords: Adenosine receptors, heart, endothelium, cell signaling pathway, diabetes.

INTRODUCTION

Adenosine is a potent endogenous nucleoside that is released from cells into the extracellular space at sites of inflammation and tissue injury which regulate many physiological functions in mammalian tissues. Its actions are mediated by interaction with specific cell membrane receptors. Four subtypes of adenosine receptors (ARs) have been identified and cloned namely A₁, A_{2A}, A_{2B}, and A₃. The significant advancement has been made in the understanding of the pharmacological and physiological relevance of ARs, but the knowledge of A₁ AR receptor still remains unclear in relation to diabetes in comparison to other receptor subtypes^[1]. The intracellular formation of adenosine, increases with increasing cellular workload and this increase is related to oxygen consumption and excitatory transmit release^[2,3].

These adenosine receptors belong to a family of G-protein coupled receptors (GPCRs) composed of a hepta-helical structure. All four receptors bind to adenosine with varying affinity and activate various signaling mechanism(s). Among these, A₁AR and A₃AR are coupled to adenylate cyclase in an inhibitory manner (being coupled to G_i protein) and A₂AR and A_{2B}AR in a

stimulatory manner (being coupled to G_s protein). Apart from this general concept, it is also known that one adenosine receptor may be coupled to more than one G proteins^[4,5]. Genetically engineered mice have played diverse physiological functions mediated by adenosine receptors, modulation of cardiovascular systems^[6,7].

The vasodilatory effects of adenosine and its analogues are mediated through adenosine A₂ receptors^[8,9]. Vasorelaxant responses to adenosine are partly mediated through adenosine induced release of endothelium derived relaxing factor (EDRF) in some blood vessels^[10,11].

Adenosine binds to its receptors; subsequently it initiates signaling cascades, most characterized mechanism being the effect on adenylate cyclase^[12]. Adenylate cyclases comprise a family of transmembrane proteins catalysing the formation of cAMP from ATP and exist in nine different isoforms, which are differentially activated by G_α, βγ subunits and intracellular calcium^[13]. Adenosine receptors couple with mitogen activated protein (MAPs) kinases and activate various downstream signaling molecules, mediating their effect of vasoconstriction or vasodilation, each receptor stimulating a specific

pathway and play major roles in cardiovascular and other systems, (Fig 1) [14,15]. On the other hand the antilipolyte effects in adipocytes, the A₁AR inhibits adenylate cyclase activity through Gi protein reduces cAMP formation and consequently inhibits protein kinase A (PKA) which ultimately reduces the hormone-sensitive lipase and/or adipose triglyceride lipase activity. These result in inhibition of the breakdown of triglycerides to free fatty acids, (Fig 2) [16,17]. There are reports which suggest organ damages such as kidney, liver and adipocytes in diabetes [18,19,20]. The direct inhibition of hormone-sensitive lipase by A₁AR agonists has not been demonstrated, because of the well-established role of hormone-sensitive lipase and more recently adipose triglyceride lipase in lipolysis. It is assumed that inhibition of lipolysis by adenosine and its analogs is due to the activation of A₁ARs, resulting in the inhibition of hormone-sensitive lipase and or adipose triglyceride lipase.

A₁AR signaling causes contraction in smooth muscle cells. The βγ subunit of A₁AR activates PLC β which causes contraction through MAPK pathway and an increase in intracellular Ca²⁺ as well as ROS. ROS causes phosphorylation in some specific domain of PKC which also plays a role in its activation.

Fig. 2: A₁AR cellular signaling in adipocytes

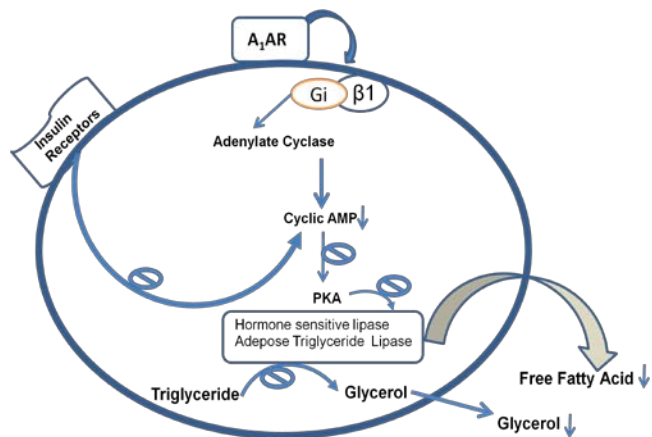


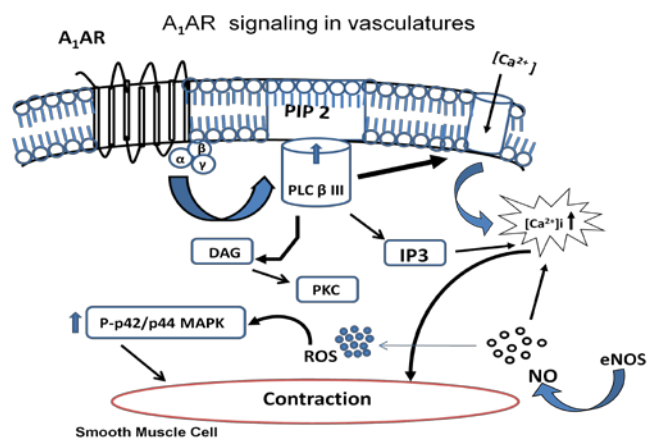
Fig. 2: A₁ARs signaling: The antilipolyte effects in adipocytes. The A₁AR inhibits adenylate cyclase activity through Gi protein reduces cAMP formation and consequently inhibits protein kinase A (PKA) which ultimately reduces the hormone-sensitive lipase and/or adipose triglyceride lipase activity. These result in inhibition of the breakdown of triglycerides to free fatty acids. Indicates inhibition in the signaling pathway.

Table 1: Agonists and antagonists of adenosine receptors

Adenosine receptor subtype	Agonists	Antagonists
A ₁ AR	CPA	DPCPX
	CCPA	WRC-0571
	CHA	BG9719
	S-ENBA	KW3902
	ADAC	FK453
		FK194921
A ₂ AR	CGS- 21680	KW6002
	HE-NECA	SCH58261
	CV-1674	VER6947
		SCH442416
A _{2B} R	LUF5853	MRS1754 MRE2029-F20
A ₃ AR	2-Cl-IB-MECA	MRS1292
		PSB-11
		MRS3777
		MRS1334
		MRE3008-F20
		MRS1523

Fredholm BB. Adenosine, an endogenous distress signal, modulates tissue damage and repair. *Cell Death and Differentiation* (2007) 14, 1315–1323.

Fig 1. A₁AR signaling in vasculatures



A_{2A} Adenosine Receptors: A_{2A} receptors are highly expressed in neurons and regulate a variety of cardiovascular functions such as vascular conductance, blood platelet aggregation and vascular relaxation [21,22]. These receptors are coupled to Gs proteins. A_{2A} receptor agonists act on endothelium as well as vascular smooth muscles to cause vasodilatation [23]. It has also been reported that A_{2A} AR activation causes vasorelaxation through cytochrome P-450 (CYP) epoxygenases and endothelium-derived hyperpolarizing factors, whereas lack of A_{2A} AR activation promotes vasoconstriction through Cyp4a in the mouse aorta [24]. The activation of A_{2A} receptor, increases MAPK activity and exert a mitogenic effect on endothelial cells by activating ERK1/2 using cAMP-ras-MEK1 pathway [25]. However, the signaling pathways by A_{2A} receptor varies with cell types and signaling machinery possessed by it. A_{2A} receptor activation in some cell lines has been known to activate PKC, Ras and SOS but not Gs, cAMP or PKA. In humans, A_{2A} receptors are present on the GABAergic output neurons in highest abundance [26]. In general, the responses produced by A_{2A} receptors can be classified as anti inflammatory and suppresses the release of inflammatory

mediators, primarily by inhibiting lymphoid or myeloid cells, including neutrophils, macrophages, lymphocytes and platelets^[27,28].

A_{2B} Receptor: A_{2B} receptors are coupled to intracellular pathways different from those of A_{2A} receptors, a finding that may provide the basis for their distinct physiological role. A_{2B} receptors have been implicated in mast cell activation and asthma, vasodilation, regulation of cell growth, intestinal function, and modulation of neurosecretion. The A_{2B} receptor subtype is coupled to both adenylyl cyclase and PLC^[29,30]. It is also known to couple to G_s protein, but recent studies have shown that these receptors may couple to G_q and produce Ca²⁺ mobilization and MAPK activation and mediates many of the important functions. In vascular endothelial cells these receptors have been found to cause vasodilatation mediated by Ca²⁺ dependent NO synthase activation^[31]. The A_{2B} AR is found to be upregulated by hypoxia and antagonists of this receptor effectively neutralize ATP-elicited reduction in post-hypoxic endothelial permeability^[32]. A_{2B} ARs are also important for adenosine-mediated inhibition of cardiac fibroblast functions and the stimulation of NO production during Na²⁺ linked absorption of glucose^[33]. It is seen that activation of A_{2B} AR causes an increase in the

release of angiogenic factors thus promoting angiogenesis^[34].

A₃ Adenosine Receptors: A₃AR is the last member of the adenosine receptor family to have been cloned and has got 40% sequence homology with A₁ and A_{2A} receptor subtypes^[35]. This receptor couples to classical second messenger system where adenylyl cyclase activity is inhibited and PLC is stimulated through G_i and G_q protein coupling^[36]. Activation of PLC is responsible for inositol triphosphate (IP3) and intracellular calcium (Ca²⁺) elevation in a variety of cellular models. These receptors are susceptible to phosphorylation by G protein coupled receptor kinases (GIRKs) which in turn leads to rapid desensitization of A₃ receptors. A₃AR is the receptor subtype that facilitates the degranulation of mast cells^[37]. In cardiac cells, A₃AR agonists activate K²⁺ channels and induce protection. RhoA-phospholipase D1 signaling mediates anti-ischemic effects of A₃AR^[38]. Like other ARs, A₃AR also couples to MAPK giving it a role in cell growth, survival, death and differentiation^[39]. A number of biological functions have been attributed to A₃AR in ischemic and inflammatory pathologies. It plays a major role in adenosine induced cardioprotection during and following ischemia-reperfusion.

Table 2: G-protein coupling of adenosine receptor subtypes

Adenosine receptor subtype	G protein	Effect of G protein coupling
A ₁	Gi 1/2/3, Go	↓ cAMP, ↑IP3/DAG, ↑arachidonate(PLA2), ↑cAMP, ↑IP3
A _{2A}	Gs, Golf, G15,16	↑cAMP, ↑IP3
A _{2B}	Gs, Gq/11	↑cAMP, ↑IP3/DAG (PLC)
A ₃	Gi/2,3 Gq/11	↓ cAMP, ↑IP3/DAG

Ref. #7 Fredholm BB, IJzerman AP, et al. International Union of Pharmacology. LXXXI. Nomenclature and Classification of Adenosine Receptors- an update. *Pharmacol Rev* 2011; 63:1-34.

A₁ Adenosine receptors

A₁ AR is a member of P1 family of seven transmembrane adenosine receptors, and is the best characterized of the widely distributed purinergic receptor family, which couples to G_i to decrease the second messenger cAMP, activates K⁺ channels or inhibits certain N, P and Q type Ca²⁺ channels in various cells^[40]. However, these receptors can stimulate Ca²⁺ mobilization through PLC β with G protein βγ subunit^[41]. This signaling pathway is synergistic with other receptors activating PLC via G_q subunit of G protein. A₁ receptors and a heterotrimeric G protein G_o, have been found to be abundant in brain suggesting a possible role of A₁ AR in

regulating neurite growth in the CNS^[42]. High expression of A₁AR is seen in brain, spinal cord, eye, atria and adrenal gland. It is intermediately expressed in skeletal muscle, liver, kidney, adipose tissue, salivary glands, esophagus, colon, antrum and testis. However, a very low expression is seen in lungs and pancreas^[43]. The molecular mass of purified A₁AR characterized from brain is 35-36 kD.

A₁AR Mediated signaling

A₁ARs have been found to produce cellular responses through a number of effector systems such as inhibition of adenylyl cyclase^[44], enhanced glucose uptake^[45], increased inositol phosphates^[46] etc. In general, these responses

have been found to be mediated by pertussis toxin sensitive G-proteins^[47]. However, the first known effector of A₁AR was inhibition of adenylate cyclase. Adenosine is found to activate the same K⁺ channel in cardiac muscle as well as other tissues, as does acetylcholine and that acetylcholine and adenosine responses are mediated by cholinergic muscarinic acid and A₁AR respectively. However, myocardial adenosine receptors and adenosine receptors in coronary arteries have also been found to be coupled via G proteins to ATP-sensitive K²⁺ channel which can be blocked by sulphonylureas^[48]. An increase in K⁺ conductance indirectly decreases Ca²⁺ entry through voltage-sensitive channels by hyperpolarizing the membrane potential. A₁ARs are also directly involved in reducing [Ca²⁺]_i by activating G proteins directly inhibitory to Ca²⁺ channels^[49].

Experiments conducted on mice have shown that the activation of A₁ARs causes contraction through PLC in the A₁AR wild-type (A₁WT) mice aorta and a decrease in coronary flow in the A₁WT mouse heart^[50]. The A₁AR coupled to Gi/o protein is known to regulate signaling pathways in various tissues, including the modulation of PLC activity, inhibition of PLA₂ and adenylate cyclase, activation of K⁺ channels, and inhibition of Ca²⁺ channels^[51-54]. Among the various PLC isoforms, PLCβIII is seen to be the predominantly activated isoform. The activation of A₁AR in turn activates PKCα which leads to p42/p44 MAPK phosphorylation in CASMCs as well as contraction of vascular smooth muscle^[55]. A₁ receptor activation can directly activate K⁺ channels and inhibit Q-, P- and N-type Ca²⁺ channels. A₁AR has been specifically shown to activate p42/44 MAPK (ERK 2) in different cells^[56]. It has been seen in COS-7 cells that A₁ receptors activate ERK1/2 via βγ subunits released from pertussis toxin-sensitive G proteins Gi/o. However, the importance of p42/44 MAPK (ERK 1/2) signaling and its relationship with PKC in causing A₁AR-mediated contraction in vasculature is still unknown. However, A₁AR may be constitutively activated at basal adenosine level of 30-300nM. In general, the activation of A₁AR mediates its effect via the following signaling pathway:

A₁AR → PLC-βIII → PKC-α → p42/p44 MAPK phosphorylation → contraction^[57].

Though the cells of the immune system express adenosine receptors and are responsive to the modulatory effects of adenosine in an inflammatory environment, still most of the

signalling pathways were uncovered in non-immune cell types, and A₁ receptor signalling mechanisms in cells of the immune system are not known.

A₁ adenosine receptor and Diabetes

Adenosine A₁ receptor (A₁-AR) activation can lower plasma glucose in diabetic rats lacking insulin and the change in A₁-AR gene expression in diabetic rats has also been reported^[20]. They concluded that the gene expression of A₁-AR in the liver is increased in insulin deficient diabetic rats. Correction of hyperglycemia by insulin or phlorizin reversed the gene expression of A₁-AR in the liver of diabetic rats, suggesting the major role of hyperglycemia in causing the change in gene expression.

Diabetes is an epidemic of the 21st century. Rare in the past diabetes has grown into an increasingly common disease both in developed countries and in the third world. It has been reported that the most important factor for this unforeseen trend appears to be the increase in body weight around the world attributable to the changes in lifestyle over the last decade. Among other complications of diabetes, cardiovascular and renal vascular diseases are among the most costly in terms of human suffering and national healthcare costs^[58-60]. It is likely that the increasing prevalence of diabetes will greatly affect the cardiovascular disease burden in the future. Although the morbidity and mortality of cardiovascular diseases has fallen over the last three decades, this trend may flatten or even reverse. Thus, a better understanding of the consequences of diabetes in the vasculature and the heart is of great importance. Indeed, diabetes markedly affects the function of the cardiovascular system, both in the microcirculation as well as in large conduit arteries supplying vital organs such as the heart, brain and kidney.

Role of adenosine receptors in ameliorating the course of diabetes has been studied. Adenosine was found to increase vascular conductance and flow in nondiabetic and diabetic rats^[61]. Vasorelaxation response to adenosine and its analogues is attenuated in certain pathological conditions affecting the blood vessels, e.g., hypertension and diabetes^[62]. Development of diabetes leads to dysfunction of many tissues including heart, there being an increased risk of congestive heart failure in patients, structural, functional and biochemical changes as well in diabetic heart^[63,64]. One of the most common structural abnormality being noted in diabetic heart is cardiomyopathy arising from

microangiopathic changes in small vessels, a few others being ventricular hypertrophy, microvascular constriction, increased collagen deposition, atherogenesis, etc^[65]. Biochemical modifications such as non-enzymatic glycation, sorbitol-myoinositol mediated changes, redox potential alterations, PKC activation and free fatty Acid metabolism have been observed in the cells of endothelium as well as myocytes^[66-68]. Among other biochemical changes, an elevated level of adenosine in diabetic heart has also been observed in animal models^[69].

In heart, activation of A₁ receptor has been found to attenuate β- adrenoceptor stimulation^[70], delay ischemic contracture^[71], and stimulate anaerobic glycolysis^[72]. Adenosine receptors are key elements in mediating cardioprotective functions of adenosine. Traditionally, A₁ receptor has been found to be the most important of all in cardioprotection. Activation of A₁AR reduces the cardiac work and myocardial oxygen consumption. Anti-ischemic effects of adenosine mediated by A₁AR have been pointed out in a number of clinical and experimental data^[73]. Though, as observed in diabetic rat, the mRNA level for A₁AR in whole heart or isolated cardiomyocytes does not change, however, A₁AR protein level increase significantly in diabetic cardiomyocytes. Over expression of A₁AR however leads to increased protection against ischemia-induced myocyte injury and enhanced pre conditioning effect^[74]. It has also been reported that A₁AR activation following treatment with sildenafil plays an integral role in the signaling cascade responsible for delayed protection against global ischemia reperfusion injury whereas adenosine A₁ receptor mediates delayed cardioprotective effect with sildenafil in mouse^[75]. A₁AR/A_{2a} AR ratio increases in cardiomyocytes of diabetic rat which may have an important physiological consequence, since both receptors exist on the same cell and have similar affinity for adenosine^[76], but activation of A_{2A} AR counteracts the anti adrenergic effect of A₁ AR^[77]. It can be assumed that an increase in A₁AR/A_{2a}AR ratio may alter physiological balance between pro and anti-adrenergic action of adenosine, this may have important consequence for failing heart.

A major complication of diabetes is the development of cardiovascular disorders, especially, hypertension. Patients with diabetes show an impairment of endothelium dependent vasodilatation. This happens partly due to the induction of ROS production by circulating free

fatty acids in diabetics^[78]. There are various enzyme systems in mammalian cells responsible for ROS generation; however, the major source of ROS generation is NADPH oxidase system^[79]. NADPH oxidase in the cells is activated by growth factors, cytokines, stress, hypoxia and G-Protein coupled agonists^[80], wherein a role of adenosine receptor can be ruled out. NADPH oxidase complex is known to be activated by PKC via Nox2^[81], PKC inturn being activated by A₁AR^[56]. The involvement of the A₁-AR in NADPH oxidase activation and in cocaine-induced LV dysfunction and suggesting the A₁-AR stimulation, at least in part via NADPH oxidase induction, plays a critical role in the events leading to the cardiomyopathy observed after cocaine abuse^[82]. Oxidative stress has been seen to upregulate A₁AR in smooth muscle cells in an experimental model. Oxidative stress and adenosine A₁ receptor activation differentially modulate subcellular cardiomyocyte mitogen actiated protein kinases and A₁ AR expression by activating nuclear factor kB^[83-85].

Under normal conditions, A₁ receptor mediated contraction is not evoked. However, in hypertensive aorta, endothelium derived contractile response to adenosine appear to be A₁AR mediated, also involving free radicles which are possibly generated through increased release of cyclo oxygenases products from endothelium of hypertensive aorta^[86]. Although evidence has been provided in a number of studies for the involvement of nitric oxide pathway in insulin induced vasodilation, alternative pathways may also exist for relaxation of vascular tone in response to insulin. One of the pathways involves Na²⁺- K²⁺ ATPases and opening of ATP sensitive potassium channels in vascular smooth muscle cells^[87]. Involvement of adenosine has also been reported that A₁AR activation negatively modulates coronary vasodilation^[88].

FUTURE PROSPECTUS

In the present time known that A₁AR, works through various signaling pathways leads to aortic constriction in mice, diabetes in rats and thus open an area in the cardiovascular research. A better understanding of above pathway will help in its pharmacological modulations by using various agonists and antagonists. This will also allow understanding the pharmacology of A₁ receptor agonists that can be utilized in the development drugs with a number of reported diseases. This may present opportunities for the treatment of cardiovascular disorders and could be therapeutically targeted.

ACKNOWLEDGEMENT

Miss Hiba Shabir, Department of Biosciences acknowledges her Research Fellowship from Jamia Millia Islamia (Central University) for the preparation of this article.

REFERENCES

1. Fredholm BB. Adenosine receptors and drug targets. *Exp Cell Res* 2010;316:1284-8.
2. Deussen A, Schrader J. Cardiac adenosine production is linked to myocardial pO₂. *J Mol Cell Cardiol* 1991; 23: 495-504.
3. Jonzon B, Fredholm BB. Release of purines, noradrenaline, and GABA from rat hippocampal slices by field stimulation. *J Neurochem* 1985; 44: 217-224.
4. Manjunath S, Pranavkumar M. Sakhare. Adenosine and adenosine receptors: newer therapeutic perspective. *Indian J Pharmacol* 2009; 41: 97-105.
5. Auchampach JA, Jin X, Wan TC, Caughey GH, and Linden J. Canine mast cell adenosine receptors: cloning and expression of the A₃ receptor and evidence that degranulation is mediated by the A_{2B} receptor. *Mol Pharmacol* 1997; 52:846-860.
6. Marzena W, Andrzej Z, Lucyna AW. New insight in to A₁ adenosine receptors in diabetes treatment. *Current Pharmaceutical Design* 2010; 16: 4237-4242.
7. Fredholm BB, IJzerman AP, Jacobson KA, Klotz KN, Linden J, Muller CE. International Union of Pharmacology. LXXXI. Nomenclature and classification of adenosine receptors-an update. *Pharmacol Rev* 2011; 63:1-34.
8. Cushing DJ, Brown GL, Sabouni, MH, Mustafa SJ. Adenosine receptor-mediated coronary artery relaxation and cyclic nucleotide production. *Am J Physiol* 1991; H343-H348.
9. Fahim M, Hussain T, Mustafa SJ. Relaxation of rat aorta by adenosine in diabetes with and without hypertension; role of endothelium. *EurJ Pharmacol* 2001; 412: 51 -59.
10. Luscher TF, Aarchus LL, Vanhoutte PM. Indomethacin improves the impaired endothelium-dependent relaxations in small mesenteric arteries of the spontaneously hypertensive rats. *Am. J. Hypertension* 1990; 3: 55-8.
11. Fredholm BB, Arslan G, Halldner L, Kull B, Schulte G, Wasserman W. Structure and function of adenosine receptors and their genes. *Naunyn-Schmiedeberg's Arch Pharmacol* 2000; 362: 364-374.
12. Tang WJ, Hurley JH. Catalytic mechanism and regulation of mammalian adenylyl cyclases. *Mol pharmacol* 1998; 54: 231-240.
13. Baines CP, Cohen MV, Downy JM. Signal transduction in ischemic preconditioning: The role of kinases and mitochondrial KATP channels. *Journal of Cardiovascular Electrophysiology* 1998; 10:741-745.
14. Linden J. Molecular approach to adenosine receptors: receptor mediated mechanisms of tissue protection. *Annu Rev Pharmacol Toxicol* 2001; 41:775-87.
15. Dobson JG Jr. Reduction by adenosine of the isoproterenol-induced increase in cyclic adenosine 3,5-monophosphate formation and glycogen phosphorylase activity in rat heart muscle. *Circ Res* 1978; 43:785-792.
16. Fain JN, Malbon CC. Regulation of adenylate cyclase by adenosine. *Mol Cell Biochem* 1979; 25: 143-169.
17. Fain JN, Pointer RH, Ward WF. Effects of adenosine nucleosides on adenylate cyclase, phosphodiesterase, cyclic adenosine monophosphate accumulation, and lipolysis in fat cells. *J Biol Chem* 1972; 247:6866-6872.
18. Londos C, Wolff J. Two distinct adenosine-sensitive sites on adenylate cyclase. *Proc Natl Acad Sci USA* 1977; 74: 5482-5486.
19. Pawelczyk T, Grden M, Rzepko R, Sakowicz M, Szutowicz A. Region-specific alterations of adenosine receptors: Expression level in kidney of diabetic rat. *American Journal of Pathology* 2005;167:
20. Liu IM, Tzen TF, Tsai, Lai TY, Chang CT, Cheng JT. Increase in adenosine A₁ receptor gene expression in the liver of streptozotocin-induced diabetic rats. *Diabetes Metab Res Rev* 2003; 19: 209-215.
21. Ongini E, Fredholm BB. Pharmacology of adenosine A_{2A} receptors. *Trends Pharmacol Sci* 1996; 17: 364-372.
22. Abebe W, Hussain T, Olanrewaju H, Mustafa SJ. Role of nitric oxide in adenosine receptor-mediated relaxation of porcine coronary artery. *Am J Physiol*

- Heart Circ Physiol 1995; 269: H1672-H1678.
23. Sexl V, Mancusi G, Holler C, Gloria-Maercker E, Schutz W, and Freissmuth M. Stimulation of the mitogen-activated protein kinase via the A_{2A}-adenosine receptor in primary human endothelial cells. *J Biol Chem* 1997; 272:5792-5799.
 24. Nayeem MA, Poloyac SM, Falck JR, Zeldin DC, Ledent C, Ponnoth DS *et al.* Role of CYP epoxygenases in A_{2A} AR-mediated relaxation using A_{2A} AR-null and wild-type mice. *Am J Physiol Heart Circ Physiol*. 2008 ;295: H2068-78.
 25. Seidel MG, Klinger M, Freissmuth M, Holler C. Activation of mitogenactivated protein kinase by the A_{2A}-adenosine receptor via a rap1-dependent and via a p21(ras)-dependent pathway. *J Biol Chem* 1999; 274: 25833-25841.
 26. Schiffmann SN, Jacobs O, Vanderhaeghen JJ. Striatal restricted adenosine A₂ receptor (RDC8) is expressed by enkephalin but not by substance P neurons: an in situ hybridization histochemistry study. *J Neurochem* 1991; 57:1062-1067.
 27. Cronstein BN. Adenosine, an endogenous anti-inflammatory agent. *J Appl Physiol*. 1994; 76:5-13.
 28. Palmer TM, Stiles GL. Identification of threonine residues controlling the agonist-dependent phosphorylation and desensitization of the rat A₃ adenosine receptor. *Mol Pharmacol*. 2000; 57:539-45.
 29. Linden J, Auchampach JA, Jin X, Figler RA. The structure and function of A₁ and A_{2B} adenosine receptors. *Life Sci*1998; 62:1519-24.
 30. Linden J, Thai T, Figler H, Jin X, Robeva AS. Characterization of human A_{2B} adenosine receptors: radioligand binding, western blotting, and coupling to G(q) in human embryonic kidney 293 cells and HMC-1 mast cells. *Mol Pharmacol* 1999; 56:705-13.
 31. Eltzschig HK, Ibla JC, Furuta JT, Leonard MO, Jakobson KA, Enjyoji K, Robson SC, Colgon SP. Coordinated adenine nucleotide phosphohydrolysis and nucleoside signaling in post hypoxic endothelium: role of ectonucleotidases and adenosine A_{2B} receptors. *J Exp Med* 2003; 198: 783-796.
 32. Matheson PJ, Spain DA, Harris PD, Garrison RN, Wilson MA. Glucose and glutamine gavage increase portal vein nitric oxide metabolite levels via adenosine A_{2B} activation. *J Surg Res* 1999; 84: 57-63.
 33. Linden J. Adenosine in tissue protection and tissue regeneration. *Mol Pharmacol* 2005; 67: 1385-1387.
 34. Meyerhof W, Muller-Brechlin R, Richter D. Molecular cloning of a novel putative G-Aprotein coupled receptor expressed during rat spermiogenesis. *FEBS Lett* 1991; 284:155-160.
 35. Abbracchio MP, Brambilla R, Ceruti S, Kim HO, von Lubitz DK, Jacobson KA, Cattabeni F. G protein-dependent activation of phospholipase C by adenosine A₃ receptors in rat brain. *Mol Pharmacol* 1995; 48: 1038-1045.
 36. Palmer TM, Benovic JL, Stiles GL. Molecular basis for subtype-specific desensitization of inhibitory molecular adenosine receptors-analysis of a chimeric A₁-A₃ adenosine receptor. *J Biol Chem* 1996; 271:15272-78.
 37. Fozard JR, Pfannkuche HJ, Schuurman HJ. Mast cell degranulation following adenosine A₃ receptor activation in rats. *Eur J Pharmacol* 1996; 298: 293-97.
 38. Mozzicato S, Joshi BV, Jacobson KA, Liang BT. Role of direct RhoA-phospholipase D1 interaction in mediating adenosine induced protection from cardiac ischemia. *FASEB J* 2004; 18: 406-408.
 39. Schulte G. and Fredholm BB. Signaling from adenosine receptors to mitogen activated protein kinases. *Cell signal*. 2003; 15: 813-827.
 40. Dickenson JM, Hill SJ. Involvement of G-protein beta-gamma subunits in coupling the adenosine A₁ receptor to phospholipase c in transfected cho cells. *Eur J Pharmacol*. 1998; 355:85-93.
 41. Chen LT, Gilman AG, Kozasa TA. Candidate target for G protein action in brain. *J Biol Chem* 1999; 274: 26931-38.
 42. Schulte G. Adenosine receptor signaling and activation of Mitogen Activated Protein Kinases. ISBN 2002; 91-7349-299-X.
 43. Linden J, Earl CQ, Patel A, Craig RH, Daluge SM. Agonist and antagonist radioligands and photoaffinity labels for the adenosine A₁ receptor. *Topics and*

- perspectives in adenosine research.1987 (Gerlach, E., and Becker, B. F., eds) pp. 3-14 Springer Verlag, Berlin.
44. Londos C, Wolff J, Cooper DMF. Adenosine as a regulator of adenylate cyclase. Purinergic receptors (Burnstock, G., ed) pp. 287-323, Chapman and Hall, London; 1981.
 45. Simpson IA, Cushman SW. Mechanism of insulin's stimulatory action on glucose transport in the rat adipose cell. Biochemical actions of hormones 1986 (Litwack, G., ed) pp. 1-31, Academic, Orlando, Florida.
 46. Alexander SPH, Hill SJ, Kendall DA. Is the adenosine receptor modulation of histamine-induced accumulation of inositol phosphates in cerebral cortical slices mediated by effects on calcium ion fluxes? *Neurochem* 1990; 55: 1138-1141.
 47. Dolphin AC, Prestwich SA. Pertussis toxin reverses adenosine inhibition of neuronal glutamate release. *Nature (London)* 1985; 316: 148-150.
 48. Kirsch GE, Codina J, Birnbaumer L and Brown AM. Coupling of ATP-sensitive K⁺ channels to Adenosine receptors by G proteins in rat ventricular myocytes. *Am J Physiol* 1990; 259, H820-H826.
 49. Dolphin AC, Forda SR, Scott RH. Calcium dependent currents in cultured rat dorsal root ganglion neurons are inhibited by an adenosine analogue. *J Physiol* 1986; 373: 47-61.
 50. Tawfik HE, Schnermann J, Oldenburg PJ, Mustafa SJ. Role of A₁ adenosine receptors in regulation of vascular tone. *Am J Physiol Heart Circ Physiol* 2005; 288: H1411-H1416.
 51. Tawfik HE, Teng B, Morrison RR, Schnermann J, Mustafa SJ. Role of A₁ adenosine receptor in the regulation of coronary flow. *Am J Physiol Heart Circ Physiol* 2006; 291: H467-H472.
 52. Ralevic V, Burnstock G. Receptors for purines and pyrimidines. *Pharmacol Rev* 1998; 50: 413-492.
 53. Shim JO, Shin CY, Lee TS, Yang SJ, An JY, Song HJ, Kim TH, Huh IH, Sohn UD. Signal transduction mechanism via adenosine A₁ receptor in the cat esophageal smooth muscle cells. *Cell Signal* 2002; 14: 365-372.
 54. Yang SJ, An JY, Shim JO, Park CH, Huh IH, Sohn UD. The mechanism of contraction by 2-chloroadenosine in cat detrusor muscle cells. *J Urol* 2000; 163: 652-658.
 55. Dickenson JM, Hill SJ. Involvement of g-protein beta-gamma subunits in coupling the adenosine A₁ receptor to phospholipase c in transfected CHO cells. *Eur. J. Pharmacol.* 1998; 355:85-93.
 56. Ansari HR, Teng B, Nadeem A, Roush KP, Martin KH, Schnermann J, Mustafa SJ. A₁ adenosine receptor-mediated PKC and p42/p44 MAPK signaling in mouse coronary artery smooth muscle cells. *Am J Physiol Heart Circ Physiol* 2009; 297: H1032-H1039.
 57. Faure M, Voyno-Yasenetskaya TA, Bourne HR. cAMP and beta gamma subunits of heterotrimeric G proteins stimulate the mitogen-activated protein kinase pathway in COS-7 cells. *J Biol Chem* 1994; 269:7851-7854.
 58. Hasko G, Linden J, Cronstein B, Pacher P. Adenosine receptors: therapeutic aspects for inflammatory and immune diseases. *Nat Rev Drug Discov.* 2008; 7: 759-770.
 59. Amos AF., McCarty DJ., Zimmet P. The rising global burden of diabetes and its complications: estimates and projections to the year 2010. *Diabet Med* 1997; 14 (suppl 5): S1-85.
 60. Bazzano LA, Serdula M, Liu M. Prevention of type 2 diabetes by diet and lifestyle modification. *J Am Coll Nutr.* 2005; 24: 92-310.
 61. Sowers JR, Epstein M, Frohlich ED. Diabetes, hypertension and cardiovascular disease: an update. *Hypertension* 2001; 37: 1053-1059.
 62. Olsson R, Jansson L, Andersson A, Carlsson PQ. Local blood flow regulation in transplanted rat pancreatic islets: influence of adenosine, angiotensin and nitric oxide inhibition. *Transplantation* 2000; 70, 280-287.
 63. Luscher TF, Aarchus LL, Vanhouttee PM. Indomethacin improves the impaired endothelium-dependent relaxations in small mesenteric arteries of the spontaneously hypertensive rats. *Am J Hypertension* 1990; 3: 55-58.
 64. Taegetmeyer H, Mc Nulty P, Young ME. Adaptation and maladaptation of heart in diabetes: Part I General concepts. *Circulation* 2002; 105: 1727-1733.

65. Hayat SA, Patel B, Khattar RS, Malik RA. Diabetic cardiomyopathy: Diagnosis and treatment. *Clin Science* 2004; 107: 539-557.
66. Brownlee M. Biochemical and molecular biology of diabetic complications. *Nature* 2001; 414: 813-820.
67. Sheetz MJ and King GL. Molecular understanding of hyperglycaemia's adverse effects for diabetic complications 2002; *JAMA* 288:2579-2588.
68. Khan ZA and Chakraborti S. endothelins in chronic diabetic complication. *Can J Physiol Pharmacol* 2003; 81: 622-634.
69. Pawelczyk T, Podgorska M, Sakowicz M. The effect of insulin on expression level of nucleoside transporters in diabetic rats. *Mol pharmacol* 2003; 63:81-88.
70. Headrick JP, Gauthier NS, Morrison RR, Matherine GP. Chronotropic and vasodilatory responses to adenosine and isoproterenol in mouse heart: effects of adenosine A₁ receptor overexpression. *Clin exp pharmacol physiol* 2000; 27:185-190.
71. Reichelt ME, Willams L, Molina JG, Sun C-X, Nobel JC, Ashton KJ, Schnermann J, Blackburn MR, Headrick JP. Genetic deletion of adenosine A₁ receptor limits myocardial ischemic tolerance. *Circ Res* 2005; 96: 363-367.
72. Finegan BA, Gandhi M, Lopaschuk GD, Clanachan AS. Antecedent ischemia reverses effects of adenosine on glycolysis and mechanical function of working hearts. *Am J Physiol* 1996; 271:H2116-H2125.
73. Headrick JP, Hack B, Ashton KJ. Acute adenosine synergic cardioprotection in ischemic reperfused hearts. *Am J Heart Circ Physiol* 2003; 285:H1797-H1818.
74. Dougherty C, Barucha J, Schofield PR, Jacobson KA, Liang BT. Cardiac myocytes rendered ischemia resistance by expressing human adenosine A₁ or A₃ receptor. *FASEB J* 1998; 12: 1785-1792.
75. Salloum FN, Das A, Thomas CS, Yin C, Kukreja RC. Adenosine A₁ receptor mediates delayed cardioprotective effect of sildenafil in mouse. *Journal of Molecular and Cellular Cardiology* 2007; 43:545-551.
76. Linden J. Molecular approach to adenosine receptors: receptor mediated mechanism of tissue protection. *Annu rev pharmacol Toxicol* 2001; 41:775-787.
77. Norton GR, Woodiwiss AJ, McGinn RJ, Lorbar M, Chungn ES, Honeyman TW, Fenton RA, Dobson JG Jr., Meyer TE. Adenosine A₁ receptor mediated antiadrenergic effects are modulated by A_{2A} receptor activation in rat heart. *Am J Physiol Heart Circ Physiol* 1999; 276: H341-349.
78. Tripathy D, Mohanty P, Dhindsa S, Syed T, Ghanim H, Aljada A, Dandona P. Elevation of free fatty acids induces inflammation and impairs vascular reactivity in healthy subjects. *Diabetes* 2003; 52:2882-2887.
79. Babior BM. The NADPH oxidase of endothelial cells. *IUBMB Life* 2000; 50: 267-269.
80. Griendling KK, Sorescu D, Ushio-Fukai M. NAD(P)H oxidase: role in cardiovascular biology and disease. *Circ Res* 2000; 86: 494-501.
81. Frey RS, Ushio-Fukai M, Malik AB. NADPH Oxidase-Dependent Signaling in Endothelial Cells: Role in Physiology and Pathophysiology. *Antioxidants and Redox Signal* 2009; 11:791-810.
82. Isabelle M, Monteil C, Moritz F, Dautreux B, Henry JP, Richard V, Mulder P, Thuillez C. involvement of the A₁-AR in NADPH oxidase activation and in cocaine-induced LV dysfunction *Cardiovas Res* 2005; 67:699-704.
83. Zhongzhen NIE, Mei Y, Ford M, Rybak L., Marcuzzi A., Ren H, Stiles GL, Ramkumar V. Oxidative stress increases A₁ adenosine receptor expression by activating nuclear factor kB, *Mol Pharmacol* 1998; 53:663-669.
84. Ramkumar V, Hallam DM and Nie Z. Adenosine oxidative stress and cytoprotection. *Jpn J Pharmacol.* 2001; 86, 265-274.
85. Croft CB, Locklar AC, Keith BJ, Mentzer Jr RM, Lasley RD Oxidative stress and adenosine A₁ receptor activation differentially modulate subcellular cardiomyocyte MAPKs, *Am J Physiol Heart Circ Physiol* 2008; 294:H263-H271.
86. Fahim M, Mustafa SJ. Evidence for presence of A₁AR in the aorta of spontaneously hypertensive rats. *British Journal of Pharmacology* 2001; 134, 1760-1766.
87. Tack CJJ, Lenders JWM, Goldstein DS, Lutterman JA, Smits P, Thien T.

Haemodynamic actions of insulin. *Curr Opin Nephrol Hypertens*; 7: 99-106.

88. Newby AC. Metabolic vasodilatation: the role of adenosine. *Biochem Soc Trans* 1988; 16:479-80.