

ORIGINAL RESEARCH ARTICLE

**Biological Treatment of Pulp and Paper Industry Effluent by White Rot Fungi
Schizophyllum commune and *Lenzites eximia***

K.Selvam* and M.Shanmuga Priya

Department of Biotechnology, Dr.N.G.P. Arts and Science College, Coimbatore -48, Tamilnadu, India

Received 03 Nov 2011; Revised 24 Jan 2012; Accepted 30 Jan 2012

ABSTRACT

White rot fungi, *Schizophyllum commune* and *Lenzites eximia* were collected from the Western Ghats region of Tamil Nadu, India from the living tree of *Tamarindus indica* and burnt tree respectively. The fungi were isolated using 2% malt extract agar medium and the fungal growth were sub cultured and incubated for 6 day at 37°C and maintained on Malt agar slants. Then, the spores were harvested without disturbing the mycelial growth using a camel hairbrush and filter sterilized. The spore concentration was adjusted to 10⁵ spores/ml and used as inoculums to treat pulp and paper industry effluents on a laboratory and pilot scales. In laboratory scale a maximum decolourization of 73.9% and 69.5% was achieved by *Schizophyllum commune* and *Lenzites eximia* on 6th day respectively. Inorganic chloride at a concentration of 539 mg/l and 534mg/l was liberated by *Schizophyllum commune* and *Lenzites eximia* respectively on 6th day of treatment. The chemical oxygen demand (COD) was reduced to 3996mg/l (67.0%) by *Schizophyllum commune* and 4317mg/l (65.0%) by *Lenzites eximia*. In Pilot scale, a maximum decolorization of 62.3% was obtained on 6th day of incubation by *Schizophyllum commune* and 56.20% by *Lenzites eximia* on 6th day. Inorganic chloride at a concentration of 477 mg/l and 469 mg/l was liberated by *Schizophyllum commune* and *Lenzites eximia* respectively on 6th day. The chemical oxygen demand (COD) was reduced to 4798mg/l (61.5%) by *Schizophyllum commune* and 5196mg/l (56.6%) by *Lenzites eximia* on 6th day treatment. These results revealed that *Schizophyllum commune* proved to be more efficient for the treatment of pulp and paper industry effluent in lab scale when compared to pilot scale.

Key words: White Rot Fungi, COD, pulp and paper industry effluent.

1. INTRODUCTION

In India there are 380 paper industries which produces a variety of different paper, paperboard as well as newsprint products. The pulp and paper industry is one of the major industries in India causing water pollution. It is estimated that 273-450m³ of water is required to produce 1 ton of paper and about 60-300m³ of wastewater is discharged.^[1] Each pulp and paper industry utilizes large amounts of water, which reappear in the form of an effluent containing large amounts of organic compounds. These higher molecular weight compounds are biologically inactive since they cannot penetrate inside the cellular membrane of living organisms and the degradation of such compounds results in lower molecular weight compounds which could be active and toxic to living organisms.^[10] The toxic solvents and chlorine compounds lead to the

formation of certain xenobiotic compounds like dioxin, biphenyls and polybrominated diphenyl ethers.^[11] When these compounds are introduced in aquatic life as well as humans the Adsorbable organic halides (AOXs) exhibit toxicity and it may bioaccumulate in fish tissue causing clastogenic, carcinogenic, endocrine and mutagenic effects.^[25] This is a major risk to human health if large amounts of fish exposed to these substances are consumed. So, the effluent generated by pulping industries is a major threat to the environment as well as human health.^[29] The effluent colour may increase in temperature and decrease photosynthesis, both of which may probably lead to decreased concentration of dissolved oxygen.^[17] The wastewaters from pulp and paper industry are toxic and cause extensive pollution. A survey within the United Kingdom

paper industry has found that the chemical oxygen demand (COD) of the pulp and paper effluent can be very high.^[32] Biological oxygen demand (BOD) and Total suspended solids (TSS) are the other two key parameters for the measurement of pollution.^[19] The physico-chemical methods are efficient only for removal of high molecular weight compounds and also the treatment cost is quiet high. So there arises a need to develop biological methods which can solve both the problem of time and cost thereby treating the effluent in effective manner.^[29] Biological methods for treating pulp and paper industry effluent are of great concern over physico-chemical methods due to their economical and ecofriendly impact.^[23] Biological treatment is categorized into aerobic treatment, anerobic treatment and fungal treatment. The aerobic treatment includes activated sludge processes, aerated lagoons and biological reactors. The microorganisms reported for aerobic treatment are *Pseudomonas putida*, *Citrobacter sp.* and *Enterobacter sp.*^[6] In aerated lagoons, the chlorinated effluents stream are mixed with general mill wastewaters and 70% of AOX removal was reported.^[30] In bioreactors moving bed biofilm reactor and suspended carrier biofilm reactor were the recent development and fungal bioremediation by *Phanerochaete chrysosporium*, *Lentinus edodes*, *Trametes versicolor*, has also been reported.^[31,15,11] White rot fungi are used for bioremediation processes because they have the ability to degrade a wide range of environmental pollutants.^[28] Bioremediation of pulp and paper industry effluent by *pleurotus sp.* has been widely reported.^[22] White rot fungi are the ideal organisms for decolourization, reduction of AOX and chemical oxygen demand (COD) and this can be achieved either by adsorption or oxidative degradation by the enzymes.^[12] Several strains of white rot fungi have been found to decolourize wood processing wastewater.^[18] The MYCOR (Mycelial Colour Removal) process for the treatment of spent chlorine bleaching liquor using *Phanerochaete chrysosporium* in rotating biological contractor was developed and this process efficiently reduces the amount of toxic low molecular weight compounds in the effluent.^[7] Extensive studies for waste water bioremediation by *Phanerochaete chrysosporium* in pulp and paper industry were reported.^[9] The decolourization of pulp mill wastewater using

thermotolerant white rot fungus *Daedaleopsis sp* was also studied.^[21]

The biological treatment of pulp and paper industry effluent by the white rot fungi *Fomes livdus* and *Thelephora sp* in both lab scale and pilot scale was reported.^[26] In the present study two newly isolated fungi *Schizophyllum commune* and *Lenzites eximia* were examined for their potential in decolourization, the reduction of COD and increase in organic chloride in both laboratory scale and pilot scale.

2. Materials and methods

2.1 Microorganism and media

The fungi, *Schizophyllum commune* and *Lenzites eximia* were collected from Western Ghats region in Tamil Nadu, India. The fungi were isolated from living tree of *Tamarindus indica*, burnt tree and used for the treatment of pulp and paper industry effluent. The fungi were identified based on the key provided previously.^[3,14] The fungal growth was cut and then sterilized with 1% mercuric chloride solution, repeatedly washed with sterile distilled water as described previously.^[33] and inoculated on 2% Malt agar medium. The fungal growth was sub cultured and incubated for 6 day at 37°C and maintained on Malt agar slants. Then, the spores were harvested without disturbing the mycelial growth using a camel hairbrush and filter sterilized. The spore concentration was adjusted to 10⁵ spores/ml and used as inoculum for further studies.

2.2 Effluent source and Treatment using a rotating biological contractor

The pulp and paper industry effluent was collected from Seshasayee paper mills, Pallipalayam, Erode, Tamilnadu, India and utilizing *Eucalyptus grandis* wood chips as a main raw material. The effluent was stored at 4°C and filtered through a 0.5mm sieve to remove large suspended particles. To analyze the efficiency of the waste water treatment, the selected fungi were grown in media by the method^[20]. In a rotating biological contractor, (890ml) was mixed with 10g of glucose and 60 ml of aqueous nutrient solution containing KH₂PO₄ 2g, MgSO₄.7H₂O-5g, CaCl₂-0.1g, NH₄Cl-0.116g, thiamine HCl-0.001g. The solution was sterilized and the pH was adjusted to 4.5 with concentrated H₂SO₄. The reactor was inoculated with 50ml of spore suspension (10⁵spores/ml) and maintained at 39°C for 4 day. On day 5, the medium was replaced by effluent of 820ml, nutrient solution without NH₄Cl 60 ml, NH₄Cl 35.3mg, benzyl alcohol 0.84ml, tween80

1.0 and 90ml of mineral solution containing nitriloacetic acid 1.5g, $MnSO_4 \cdot H_2O$ - 0.5g,

$FeSO_4 \cdot 7H_2O$ - 0.1 g, $CoSO_4$ - 0.1 g, $ZnSO_4$ - 0.1 g, $CuSO_4 \cdot 5H_2O$ - 0.01 g, $AlK(SO_4)_2$ - 0.01 g, H_3BO_3 - 0.01 g, $NaMoO_4$ - 0.01 g. The pH of the solution was adjusted to 4.5 with concentrated H_2SO_4 and the reactor was maintained at 39°C and continuously flushed with oxygen. After treatment the mycelia were harvested and their efficiency for reducing the colour, increasing the inorganic chloride content and reducing the Chemical Oxygen Demand were analyzed according to the methods reported previously.^[2] Samples were withdrawn at regular intervals every day.

3. RESULTS AND DISCUSSION

The potential of two white rot fungi namely *Schizophyllum commune* and *Lenzites eximia* were assessed when the paper mill effluent was treated with these selected fungi on two scales namely laboratory scale and pilot scale because different scales can show different efficiencies in the treatment. The colour, the chloride content and the COD in effluents are regarded as important factors to evaluate its quality. In laboratory scale experiments with, *Schizophyllum commune* the colour was reduced at a maximum by 73.9 % when compared with untreated effluent by 6 day incubation. The liberation of inorganic chloride was increased upto 282.0% (539 mg/l) of that in the untreated effluent on the 6th day of treatment and the COD was reduced to 3996mg/l (67.0%). In *Lenzites eximia* the percent of colour removed was at a maximum of 65.0% when compared to untreated effluent by 6 day incubation. The liberation of inorganic chloride was increased upto 534 mg/l (278.1%) when compared with untreated effluent by the 6th day incubation and the COD was reduced to 4317 mg/l (65.0%) (Fig 1). In pilot scale treatment with *Schizophyllum commune* the colour was reduced at a maximum by 62.3% of untreated effluent by 6 day incubation. The liberation of inorganic chloride was increased upto 477 mg/l (238.0%) of that in the untreated effluent by the 6 day incubation and the COD was reduced to 4798mg/l (61.3%). In *Lenzites eximia* the colour was removed at a maximum by 56.2% when compared to untreated effluent on 6 day incubation. The liberation of inorganic chloride was increased upto 477mg/l (231.2%) of that in the untreated effluent by the 6 day incubation and the COD was reduced to 4798 mg/l (61.3%). (Fig 2). These results revealed that laboratory scale experiments was more efficient

when compared to pilot scale experiments. *Pleurotus ostreatus* removed the

colour of kraft mill effluent by 69.0% and COD was reduced to 66.9% after fed batch treatment of kraft mill effluent.^[8] *Pleurotus sajor caju* decolourized the paper mill effluent by 66.7% on day 6 of incubation. Inorganic chloride liberated by *Pleurotus sajor caju* was 230.9% and chemical oxygen demand (COD) was reduced by 61.3% on 10 day treatment. In pilot scale treatment maximum decolourization was obtained by *Pleurotus sajor caju* 60.1% on 6 day of incubation. Inorganic chloride was increased by 524.0 mg/l and the COD was reduced by 1442.0mg/l (57.2%) by *Pleurotus sajor caju* on day 7 of incubation^[22]. *Trametes versicolor* on the fourth day of treatment showed a maximum decolourization of 63.9% in laboratory scale, Inorganic chloride at a concentration of 765mg/l, was liberated by *Fomes lividus* on the 10th day. The Chemical oxygen demand was also reduced to 1984mg/l by *Fomes lividus*. On the pilot scale, a maximum decolourization of 68% was obtained with the 6 day incubation by *Trametes versicolor*, inorganic chloride 475mg/l (103%) was liberated on the 7th day by *Trametes versicolor* and the COD was reduced by 1984 mg/l by *Fomes lividus*.^[26] *Daedaleopsis sp* and *Phanerochaete chrysosporium* exhibited the highest ability to decolourize waste water by 52% and 86% respectively, COD was reduced by 59-71% and 66-83%.^[21] The treatment of paper mill effluent in laboratory scale with *Thelephora sp.* a, maximum decolourization of 43.1% was observed on 4th day treatment. Inorganic chloride at the concentration of 751mg/l was liberated 10th day. The chemical oxygen demand was also reduced to 1840 mg/l in laboratory scale. In pilot scale, a maximum decolourization by 23.6% was obtained on 10th day incubation, inorganic chloride 361mg/l was liberated on the 6th day and the chemical oxygen demand was reduced to 2,000mg/l.^[27] *Trametes versicolor* reduced biological oxygen demand and chemical oxygen demand of paper mill effluent by 52% and 42% respectively.^[24] A maximum colour removal of 57% and 67% reduction of chemical oxygen demand after 14th day of incubation when treated with the white rot fungus *pleurotus sajor caju* was reported^[5]. *Ceriopsis subvermispora* could decolourize kraft-bleaching effluent at 90% and also resulted in reduction of COD of up to 45%.^[13] In the present study newly isolated white

rot fungi *Schizophyllum commune* and *Lenzites eximia* lignin and or lignin derivatives when compared to previous reports (Table 1).
eximia have superior potential to dechlorinate

Table 1: Comparison of the efficiencies of the treatment in lab scale and Pilot scale

Treatment	Content removed during treatment		
	Colour ^a (%)	Chloride (mg/l)	COD (mg/l)
Control	0.69	141	12000
<i>Schizophyllum commune</i>			
Lab scale	0.18(73.9)	539(282.0)	3996(67.0)
Pilot Scale	0.26 (62.3)	477(238.0)	4798(61.5)
<i>Lenzites eximia</i>			
Lab scale	0.21(69.50)	534(278.1)	4317(65.0)
Pilot Scale	0.30(56.20)	469(232.0)	5196(56.6)

Colour-% decrease over control, Chloride content-% increase over control, COD-% decrease over control.

^a Absorbance at 465 nm

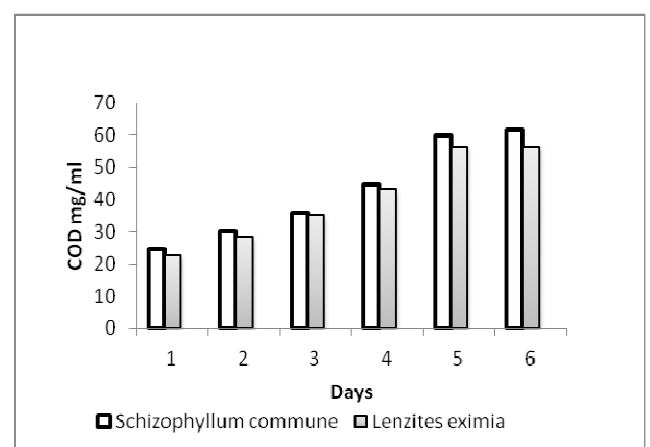
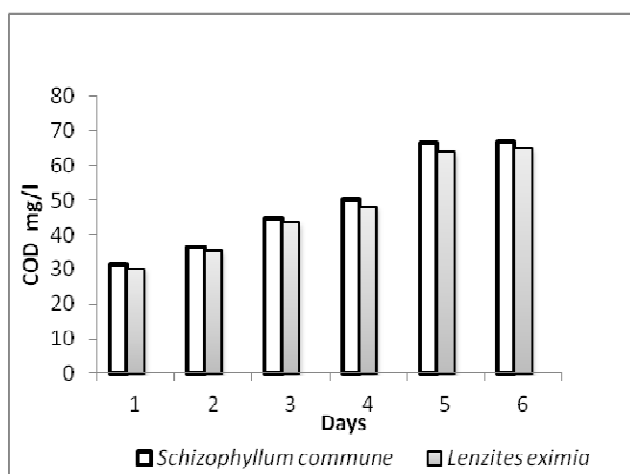
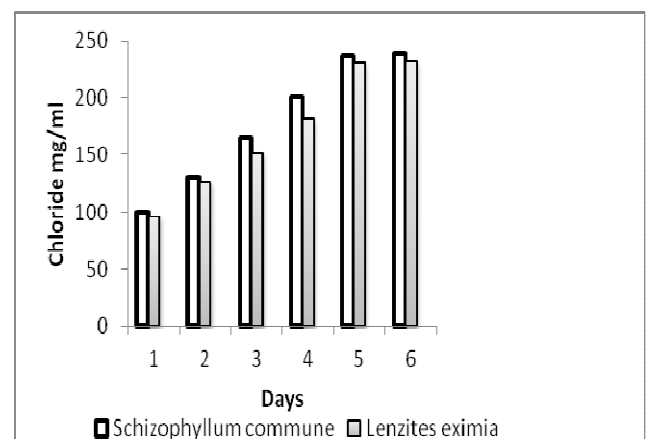
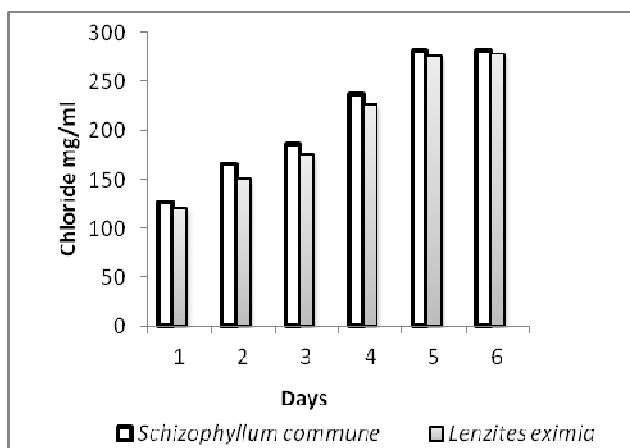
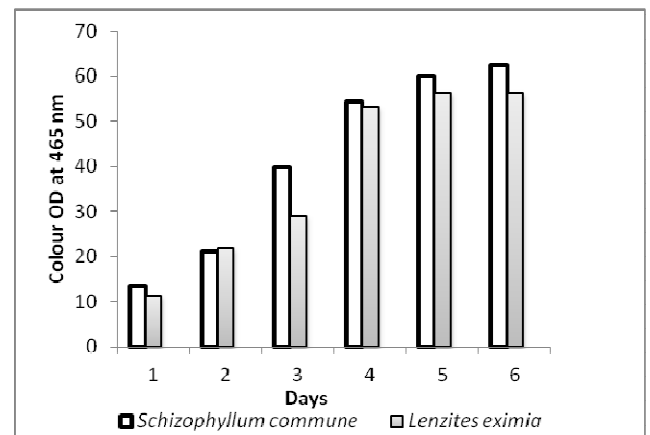
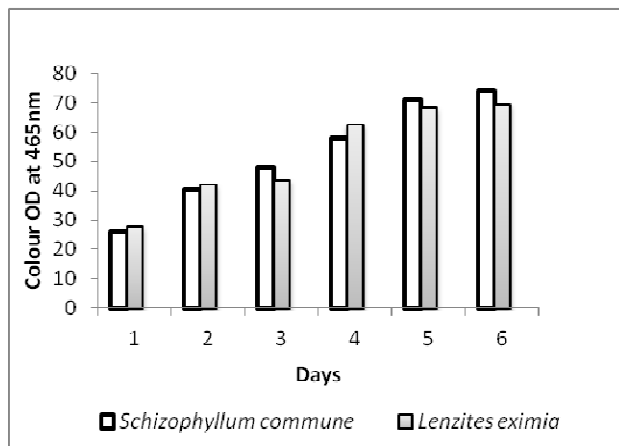


Fig 1: Colour removal of pulp and paper mill effluent by *Schizophyllum commune* and *Lenzites eximia* on laboratory scale. Colour (OD at 465nm)-values decrease over control, chloride content (mg/l)-values increase over control. COD (mg/l)- values decrease over control.

Fig: 2 Treatment of pulp and paper mill effluent by *Schizophyllum commune* and *Lenzites eximia* on pilot scale. Colour (OD at 465nm)-values decrease over control, chloride content (mg/l)-values increase over control. COD (mg/l)- values decrease over control.

ACKNOWLEDGEMENT

This research work was funded by University Grant Commission (UGC) major research project, No.F.35-31/2009 (SR). The authors thank University Grant Commission (UGC) for their constant financial support and guidance rendered throughout the period of study.

REFERENCES

1. Addison. R., Ikonomou. M., Smith.T, 2005, PCDD/F and PCB in harbor seals (*Phoca vitulina*) from British Columbia: response to exposure to pulp mill effluents, *Journal of Marine Environmental research* 59, 165-176.
2. APHA, 1976, Standard Methods for the Examination of Water and Wastewater, American Public Health Association, New York : ISBN 0-9662376-0.
3. Bakshi. B,K, 1971. Indian polyporaceae – on Trees and Timbers, New Delhi: Indian Council for Agricultural Research (ICAR) publications, 80–81.
4. Battaglia. A, N., Calace. E., Nardi. B,M., Petronio. M, Pietroletti, 2003, Paper mill sludge-soil mixture: Kinetic and thermodynamic tests of cadmium and lead sorption capability, *Microchemistry, Journal* 75: 97-102,
5. Belem.A, Panteleitchouk. A,V, Durate. A,C, Rochasantos. T,A,P, Freitas, A,C 2008 Treatment of the effluent from a kraft bleach plant with white rot fungi *pleurotus sajor caju* and *pleurotus ostreatus*, *Global NEST Journal* 10: 426-431,
6. Chandra. R, 2001, Microbial decolourization of pulp mill effluent in presence of nitrogen and phosphorous by activated sludge process, *Journal of environmental biology*, 22, 23-27.
7. Chang. H,T, W., Joyce, T,K, Krik and V,B, Huynh, 1985, Process of degrading chloroorganics by white rot fungi, U,S, Patent No,4,554,675.
8. Choudhury.S, RohellaManthan, M, and Sahoo. N, 1998, Decolourization of kraft mill effluent by white rot fungi, *Indian Journal of Microbiology*, 38, 221–224.
9. Christov. L, Driessel, B,V, 2003, Waste water bioremediation in the pulp and paper industry, *Indian Journal of Biotechnology*, 2, 444-450.
10. Coulibaly. L, Gourene. G, and Agathos N,S, 2003, Utilization of fungi for biotreatment of rawwastewaters, *African journal, Biotechnology*, 2, 620-630.
11. Couto.S, and Herrera, j, 2006, Industrial and biotechnological applications of laccases: A review, *Journal of biotechnology advances*, 24: 500-513.
12. Fu.Y, Viraraghavan, T, 2001, Fungal decolourization of Dye wastewaters, A review, *Bioresource Technol.,* 79, 3,251-262.
13. Ghoreishi. S, M, Haghghi. R 2007 Chromophores removal in pulp and paper mill effluent via hydrogenation-biological batch reactors, *Chem Eng JOURNAL*, 127, 59–70.
14. Gilbertson, R, L, and Ryvarden, L, 1986, North American Polypores,Oslo: Fungiflora, 1, p, 33.
15. Hofrichter, M, 2002, Review: lignin conversion by manganese peroxidase (MnP), *journal of enzyme and microbial technology* 30, 454-466.
16. Janasekhar.H, and Fietchter, A, 1988 Cultivation of *Phanerochaetechrysosporium* and production of lignin peroxidase in submerged stirred tank reactors, *Journal,Biotechnol.,* 8,97-112.
17. Kingstad. K, P, and Lindstorm, K, 1984, Spent liquors from pulp bleaching, *Environmental science and technology* 18: 236-248.
18. Luciana. C, Germain. G, Spiros. A,N 2003, Utilization of fungi for biotreatment of raw wastewater, *African Journal Biotechnology*, 2: 620–30,
19. OFIA (Ontario Forest Research Association), Pulp and paper mill effluent on the environment, 2005.
20. Pellinen. J, Joyce, T,W., and Chang, H,M, 1988, Dechlorination of high molecular weight chlorolignin by the white-rot fungus *Phanerochaete chrysosporium*, *TAPPI Journal* 71, 191–194.
21. Prasongsuk. S., Lotrakul. P., Imai.T., Punnapayak. H, 2009, Decolourization of pulp and paper mill wastewater using thermotolerant white rot fungi, *Science Asia* 35: 37-41.
22. Ragunathan. R., Swaminathan, K., 2004, Biological treatment of pulp and paper industry effluent by *Pleurotus* sp, *World*

- Journal Microbiology Biotechnology, 20: 389-393.
23. Ruggaber, T., Talley, J., 2006, Enhancing bioremediation with enzymatic processes : a review, Practice periodical of Hazardous, toxic, and Radioactive waste management, 10: 73-85.
 24. Saetang, J., and Babel, S., 2009, Effect of leachate loading rate and incubation period on the treatment efficiency by *T. versicolor* immobilized on foam cubes Int, Journal, Environment Science Technology, 6 3: 457-466
 25. Savant, D., Abdul, R., and Ranade, D., 2006, Anaerobic degradation of adsorbable organic halides (AOX) from pulp and paper industry wastewater, Journal of Bioresource Technology, 97:1092-1104.
 26. Selvam, K., Swaminathan, K., Song, M, H., Chae, K, S, 2002, Biological treatment of pulp and paper industry effluent by *Fomes lividus* and *Trametes versicolor*, World journal Microbiology Biotechnology 18, 523-526.
 27. Selvam K, Swaminathan K, Rasappan K, Rajendran R, Pattabhi S (2006). Decolourization and dechlorination of a pulp and paper industry effluent by *Thelephora* sp. Ecology environment conservation. 12, 223-226.
 28. Shah, V, Nerud, F, 2002, Lignin degrading system of whiterot fungi and its exploitation for dye decolourization, Canadian Journal microbiology, 48: 857-870.
 29. Singh, Y, P., Dhall, R, M., Mathur, R, K., Jain, V, V., Kumar, V., Kumar, R., Kumar, A, 2011, Bioremediation of pulp and paper mill effluent by tannic acid degrading *Enterobacter* sp, Water Air Soil Pollution, 218:693-701.
 30. Stuthridge, T, R., and Mafarlane, P, N, 1994, Adsorbable organic halide removal mechanisms in a pulp and paper mill aerated lagoon treatment system, Water science and Technology, 29,195-208.
 31. Tang, L., Zeng, G., Wang, H., Shen, G., Huang, D, 2005, Amperometric detection of lignin degrading peroxidase activities from *Phanerochaete chrysosporium*, journal of enzyme and microbial technology, 36: 960-966.
 32. Thompson, J., Swain, M., Kay, C, F., Forster, 2001, The treatment of pulp and paper mill effluent, a review, Bioresource Technology , 77:275-286.
 33. Watling, R, 1971, Basidiomycetes: homobasidiomycetidae, In Methods in Microbiology, ed, Booth, C, pp, 219-236, London and New York: Academic Press, ISBN 12-521504-5.