

ORIGINAL RESEARCH ARTICLE

Antimicrobial Activity of Sponges Endemic to Uchipully Coastal Line, Near Rameswaram, TamilnaduNarayanaswamy Tamilselvan¹ and Ernest David^{*2}

¹Post graduate and Research Department of Zoology, Physiology wing, Voorhees College, Vellore – 632001, Tamilnadu, India

²Department of Biotechnology, Thiruvalluvar University, Serkadu, Vellore-632115

Received 16 Sep 2012; Revised 01 Dec 2012; Accepted 09 Dec 2012

ABSTRACT

The study reports the *in-vitro* screening of methanolic extracts of nine marine sponges (porifera) collected from Uchipully coastal line, near Rameswaram, Tamilnadu, India, in search for novel pharmaceuticals. In vitro antimicrobial activity for nine sponge species against pathogenic strains of bacteria viz *Escherichia coli*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Bacillus subtilis*, *Staphylococcus epidermis*, *Staphylococcus aureus* and pathogenic strains of fungi *Aspergillus niger*, *Aspergillus flavus*, *Aspergillus fumigatus* was tested using agar well diffusion assay method. The results revealed that the sponge extracts inhibited the growth of majority of the bacterial as well as fungal pathogens. The maximum growth inhibition of *Escherichia coli* was recorded with all the extracts of sponges.

Keywords: Antimicrobial activity, Bacteria, Sponges.

INTRODUCTION

Marine sponges constitute an important component of benthic communities throughout the world, with regard to its biomass as well as their potential to influence benthic or pelagic processes (Dayton 1974; Dayton 1989; Gili 1998; Maldonado 2005). Sponges exist in oceans for about 580 million years, and diversified with 8,000 species today (Bergquist 1978; Van Soest *et al.*, 2008). They inhabit a wide variety of marine and freshwater ecosystems and are found throughout tropical, temperate and polar regions (Hooper *et al.*, 2002). Marine sponges, especially those found in tropical ocean areas, continue to represent the single most prolific source of structurally novel natural products of marine origin (Blunt *et al.*, 2009). Sponges (Phylum: Porifera) are among the oldest multicellular animals (Metazoa) and show relatively little differentiation and tissue coordination (Bergquist, 1978; Simpson, 1984; Leys and Meech, 2006). Sponges, which are the most primitive invertebrates, represent an important constitutive group of the coral reef fauna with a wide range of species (Van Soest, 1994). Sponges are known to

produce large number and vast variety of secondary metabolites.

Sponge-derived secondary metabolites have also attracted considerable attention because of their relevant and diverse pharmacological actions (Keyzers and Davies-Coleman, 2005). Some of the products showing antiviral, antitumor, antimicrobial or general cytotoxic properties. Certain marine sponges also contain amino acid derivatives. Biologically active peptides are the axinellins (Maldonado *et al.*, 2005), kapakahines (Bergquist, 1978), microsclerodermins (Simpson, 1984), polytheonamides (Leys and Meech, 2006), papuamides (Hooper *et al.*, 2002), stylisins (Blunt *et al.*, 2009), hymenamides (Kobayashi *et al.*, 1993), wainunuamide and dominicin (Williams *et al.*, 2005). Additional noteworthy examples include the laxaphycins (Bonnard *et al.*, 2007) and discodermins (Matsunaga *et al.*, 1984; Matsunaga *et al.*, 1985) they inhibit tumor promotion), calyculin¹⁵ (tumor promoting), and the antithrombin cyclotheonamides. Some of the isolated substances from sponges have striking structural similarities to metabolites of microbial origin (Proksch *et al.*, 2002). Over 5000

***Corresponding Author:** Dr. Ernest David, **Email:** ernestdavid2002@yahoo.com, **Phone No:** +91-9345300236

secondary metabolites were isolated from 500 species of marine sponges (Rifai *et al.*, 2005). Over 800 antibiotics have been isolated from marine sponges (Touati *et al.*, 2007). Many of these compounds likely to serve as chemical weapons that protect the immobile animals from being overgrown or ingested (Pawlik, 1992; Paul and Ritson-Williams, 2008). The first report of antimicrobial activity of sponge extract was by Nigrelli *et al.* (Newbold *et al.*, 1999). The present study was carried out to test the antimicrobial activity of the methanol extracts of sponges, against bacterial and fungal species.

MATERIALS AND METHODS

Sample Collection

Sponges sample were collected from Uchipully near Mandapam coast, Rameswaram, Tamilnadu, India. Samples were collected through normal diving at the depth of 5-6 m during September – October 2010 and immediately kept in methanol and transferred to lab for further study. Nine sponge types (A, B, C, D, E, F, G, H, I) were collected for this study.

Extract Preparation

The shade dried sponges powdered in an electric blender and was extracted separately to exhaustion in a Soxhlet apparatus using methanol solvent system. Sponges extracts were filtered through a cotton plug followed by Whatmann filter paper No.1 and then concentrated by using a rotary evaporator at low temperature (40-50°C). Extracts were preserved in air tight container and kept at 4-5°C until further use. The dried extracts were dissolved in dimethyl sulphoxide (DMSO) and subjected to study the antimicrobial activity.

Test organisms

The bacterial spp used for the test were *Escherichia coli* (*E.coli*), *Pseudomonas aeruginosa* (*P.aeruginosa*), *Klebsiella pneumoniae* (*K. pneumonia*), *Bacillus subtilis* (*B.subtilis*), *Staphylococcus epidermis* (*S.epidermis*), *Staphylococcus aureus* (*S.aureus*) and of the Fungal spp were *Aspergillus niger* (*A.niger*), *Aspergillus flavus* (*A.flavus*), *Aspergillus fumigates* (*A.Fumigatus*)

Antimicrobial susceptibility test by Agar well diffusion method

In order to determine the antimicrobial spectrum, the antibacterial activity and antifungal activity

was performed by agar well diffusion method. The inoculum suspension of each bacterial strain was swabbed on the entire surface of Mueller Hinton Agar (MHA). On the surface of the medium, wells were made by using sterile cork borer (6 mm size). Each well was filled with 100 µl of sponges extracts. The diameter of inhibition zones were measured in mm after incubation at 37°C for 24 hours.

RESULTS AND DISCUSSION

The results revealed potential antimicrobial activity in extract sample B, C, D, E, F, G, H and I (**Table 1 & 2**). The antimicrobial effect of sample D was recorded higher on *P.aeruginosa*, *E.coli*, *K. pneumoniae*, *B. subtilis*, *A. niger*, *A. fumigatus*, *S. aureus* and *A.flavus* when compared to that of other samples. In case of sample B, *E.coli* inhibited maximally compared to that of *P. aeruginosa*, *B.subtilis*, *K. pneumoniae* and *S. aureus*. Sample E revealed significant zone inhibition for *E.coli* to that of *P. aeruginosa*, *B.subtilis*, *K. pneumoniae* and *S. aureus*. In case of antifungal effect, sample D showed maximum activity on *A.fumigatus*, *A.niger* and *A.flavus* compared to that of other samples.

Similar studies revealing potent antimicrobial activity of marine sponges were recorded elsewhere, Rifai *et al.*, 2005, reported that ten marine sponges extract collected from Atlantic coast of Morocco and from the Gulf of Thailand were tested against four bacterial pathogens and five fungal pathogens. Touati *et al.*, 2007, reported that marine sponge extracts collected from Tunisian coast was tested against eight human pathogenic bacteria and six human pathogenic fungi using the agar disk diffusion method. Galeano and Martnez, 2007, reported that twenty-four sponge species (Poriferae), collected from the Uraba Gulf reefs (Colombian Caribbean region), against certified strains of bacteria (*Staphylococcus aureus* ATCC 25923 and *Escherichia coli* 25922) and yeast (*Candida albicans* 10231). Safaeian *et al.*, 2009 studied that, *in vitro* antimicrobial activity of six sponge species (Porifera), collected from offshore zone of Nay Band Bay, Iran, against pathogenic strains of bacteria (two Gram-positive: *Bacillus subtilis* ATCC 6633, *Staphylococcus aureus* ATCC 25923 and two Gram negative: *Pseudomonas aeruginosa*

ATCC 27853, *Escherichia coli* ATCC 25922) and pathogenic fungi (*Candida albicans* ATCC 10231, *Aspergillus spp.* PTCC 5266, *Penicillium spp.* PTCC 5251).

Table 1: Antibacterial activity of methanolic extracts (100 µl sample) of various marine sponge types

Sample ID	<i>E.coli</i>	<i>B.subtilis</i>	<i>S.aureus</i>	<i>P.aeruginosa</i>	<i>K.pneumoniae</i>
A	-	-	-	9 mm	-
B	16 mm	14 mm	13 mm	15 mm	14 mm
C	-	-	10 mm	-	-
D	18 mm	17 mm	15 mm	19 mm	18 mm
E	15 mm	13 mm	12 mm	14 mm	13 mm
F	14 mm	12 mm	10 mm	11 mm	14 mm
G	12 mm	10 mm	10 mm	12 mm	11 mm
H	-	-	-	-	10 mm
I	-	-	-	10 mm	-

*(Zone of inhibition in mm)

Table 2: Antifungal activity of methanolic extracts (100 µl sample) of various marine sponge types

Sample ID	<i>A.niger</i>	<i>A.fumigatus</i>	<i>A.flavus</i>
A	-	-	-
B	13 mm	15 mm	13 mm
C	-	9 mm	-
D	17 mm	19 mm	16 mm
E	12 mm	13 mm	14 mm
F	10 mm	12 mm	10 mm
G	9 mm	10 mm	9 mm
H	-	-	-
I	-	10 mm	-

*(Zone of inhibition in mm)

Figure 1: Different sponges

Figure 1: A

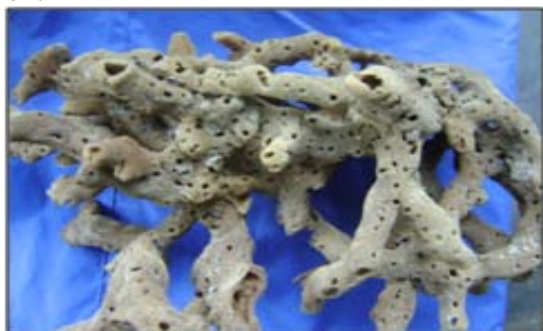


Figure 1: B



Figure 1: C



Figure 1: D



Figure 1: E



Figure 1: F



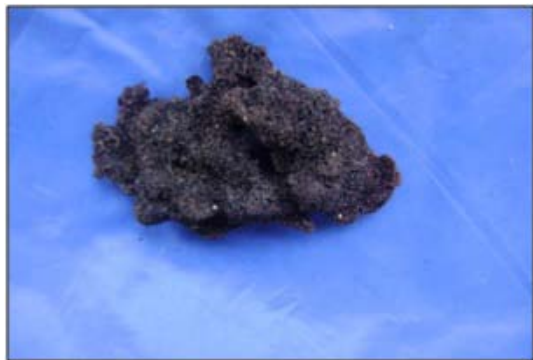
Figure 1: G



Figure 1: H



Figure 1: I



CONCLUSION

The present studies envisage exploiting potent antimicrobial agents from marine sponges of Uchipully coastal line.

ACKNOWLEDGEMENT

Authors are thankful to Voorhees College, Vellore, Tamilnadu for providing laboratory facilities to carry out this study

REFERENCE

- Bergquist PR. (1978) in *Sponges*. Hutchinson and Co. Ltd., London,UK. 268.
- Blunt, J W Copp BR, Hu WP, Munro, M. H. G.; Northcote, P. T.; Prinsep, M. R. *Nat. Prod. Rep.* 2009, 26, 170–244.
- Dayton, P.K. (1974) *Ecol. Monogr.*, 44, 105-128.
- Dayton, P.K. (1989) *Science*, 245, 1484-1486.
- Gili, J.-M. and Coma, R. (1998) *Trends Ecol. Evol.*, 13, 316-321.
- Malonado M, Carmona C, Velasquez Z, Puig A, Cruzado A, Lopez A. and Young, C.M. (2005) *Limnol. Oceanogr.*, 50, 799-809
- Simpson, T.L. (1984) *The cell biology of sponges*. Springer- Verlag, New York, NY, p. 662.
- Leys S P and Meech R W. (2006) *Can. J. Zool.*, 84(2), 288-306.
- Hooper, J.N.A. and van Soest, R.W.M. (2002) in *Systema Porifera: a guide to the classification of sponges*. Kluwer Academic/Plenum Publishers, New York, NY, p. 1810.
- Van Soest RWM. Demosponge distribution patterns. In: Van Soest RWM, Van Kempen TMG, Braekman JC, editors. *Sponges in time and space*. Rotterdam: Balkema; 1994. p. 213–23.
- Rifai S, Fassouane A, El-Abbouyi A, Wardani A, Kijjoa A, Van Soest R. Screening of antimicrobial activity of marine sponge extracts. *J Mycol Med* 2005; 15:33–8.
- Touati I, Chaieb K, Bakhrouf A, Gaddour K. Screening of antimicrobial activity of marine sponge extracts collected from Tunisian coast. *J Mycol Med.* 17 (2007) 183–7.
- Newbold RW, Jensen PR, Fenical W, Pawlik JR. Antimicrobial activity of Caribbean sponge extracts. *Aquat Microb Ecol* 19 (1999) 279–84.
- Proksch P, Edrada RA, Ebel R. *Appl Microbiol Biotechnol* 59 (2002) 125- 34.
- Blunt J W, Copp B R, Hu W P, Munro M H G, Northcote P T, Prinsep M R. 26 (2009). *Marine natural products*. *Nat. Prod. Rep.* 26, 170–244.
- Paul V J, Ritson-W R. *Marine chemical ecology*. *Nat. Prod. Rep.* 25 (2008) 662–695.
- Pawlik J.R. Chemical ecology of the settlement of benthic marine invertebrates. *Oceanogr. Mar. Biol.* 30 (1992) 273–335.
- Keyzers R A, Davies C M T. *Chem. Soc. Rev.* 34 (2005) 355–365. (Introduction protein peptide reference)
- Randazzo A, Dal P F, Orru S, Debitus C, Roussakis C, Pucci P, Gomez P L. *Eur. J. Org. Chem.* 1998 2659–2665.
- Nakao Y, Yeung B K S, Yoshida W Y, Scheuer P J J. *Am. Chem. Soc.* 117 (1995) 8271–8272;
- Yeung B K S, Nakao Y, Kinnel R B, Carney J R, Yoshida W Y, Scheuer P J, Kelly B M J. *Org. Chem.* 61 (1996) 7168–7173.
- Qureshi A, Colin P L, Faulkner D J. *Tetrahedron* 56 (2000) 3679–3685.
- Hamada T, Matsunaga S, Yano G, Fusetani N J. *Am. Chem. Soc.* 127 (2005) 110–118.
- Ford, P. W.; Gustafson, K. R.; McKee, T. C.; Shigematsu, N.; Maurizi, L. K.; Pannell, L. K.; Williams, D. E.; de Silva, E. D.; Lassota, P.; Allen, T. M.; Van Soest, R.; Andersen, R. J.; Boyd, M. R. J. *Am. Chem. Soc.* 1999, 121, 5899–5909.
- Mohammed R, Peng J, Kelly M, Hamann M. T. J. *Nat. Prod.* 69 (2006) 1739–1744.
- Kobayashi J, Tsuda M, Nakamura T, Mikami Y, Shigemori H. *Tetrahedron* 49 (1993) 2391–2402;

27. Tabudravu J, Morris L A, Van D, Jaspars M. *Tetrahedron Lett.* 42 (2001) 9273–9276.
28. Williams D E, Patrick B O, Behrisch H W, Van S R, Roberge M, Andersen R J. *J. Nat. Prod.* 68 (2005) 327–330.
29. Bonnard I, Rolland M, Salmon, J M. Debiton E, Barthomeuf C, Banaigs B. *J. Med. Chem.* 50 (2007) 1266–1279.
30. Matsunaga S, Fusetani N, Konosu S. *J. Nat. Prod.* 48 (1985) 236–241.
31. Matsunaga S, Fusetani N, Konosu S. *Tetrahedron Lett.* 25(1984) 5165–5168.
32. Kato Y, Fusetani N, Matsunaga S, Hashimoto K, Koseki K. *J. Org. Chem.* 1988, 53, 3930–3932.
33. Rifai S, Fassouane A, El-Abbouyi A, Wardani A, Kijjo A, Van SR. Screening of antimicrobial activity of marine sponge extracts. *Journal de Mycologie Médicale* 15 (2005) 33–38
34. Touati I, Chaieb K, Bakhrouf A, Gaddour K. Screening of antimicrobial activity of marine sponge extracts collected from Tunisian coast. *Journal de Mycologie Médicale* 17 (2007) 183–187.
35. Galeano E, Martinez A. Antimicrobial activity of marine sponges from Uraba Gulf, Colombian Caribbean region. *Journal de Mycologie Médicale.*, 17 (2007) 21–24.
36. Safaeian S, Hosseini H, Abbas PAA, Farmohamadi S. Antimicrobial activity of marine sponge extracts of offshore zone from Nay Band Bay, Iran. *Journal de Mycologie Médicale.*, 19 (2009) 11–16