

Available Online at www.ijpba.info

International Journal of Pharmaceutical & Biological Archives 2013; 4(1):1-8

REVIEW ARTICLE

Microalgae, Vitamins and Multi-Mineral Supplementation-Promising Protective Effects against Cadmium Toxicity

Ali Annabi*, Khaled Said, Imed Messaoudi

Laboratoire de Recherche (LR11 - ES41): Génétique, Biodiversité et Valorisation des Bio - ressources. Institut Supérieur de Biotechnologie de Monastir, Université de Monastir, Tunisie

Received 15 April 2013; Revised 07 May 2013; Accepted 08 May 2013

ABSTRACT

Heavy metals like Cd, Pb and Hg have no known biological functions and consequently detrimental to essential life processes. These metals in the form of inorganic compounds from natural and anthropogenic sources continuously enter the aquatic ecosystem where they pose a serious threat because of their toxicity. Pollution of the aquatic environment with metals has become a serious health concern in recent years. Although environmental concentrations of metals are rarely directly dangerous for fish survival, are known to accumulate in fish tissues reaching concentrations of up to thousands of times higher than in the surrounding water environment and becoming extremely harmful. Cadmium (Cd) is widely distributed in aquatic environments and is an extremely toxic metal commonly found in industrial settings. It has been demonstrated that the feed is the principal source of contamination by metals such as Cd that tends to accumulate in tissues, which in the end are ingested by consumers. However, it has been others parameters: water hardness, water minerals concentration (calcium, Zinc and Selenium), protective effects of supplements (microalgae, vitamins). In this way, this mini-review aimed to bring the main work on the promising effects of microalgae (e.g Spirulina), Vitamines (C and E) and multi-mineral supplementation against the toxicity of cadmium.

Key words: Cadmium, Heavy metal toxicity, protective effects.

1. INTRODUCTION

Water is one of the most valuable natural resources and a major environmental concern due to dispersal of industrial and urban wastes generated human activities is by the contamination of soil and water. A wide range of inorganic and organic compounds cause contamination includes heavy metals, combustible substances and hazardous wastes. Major component of inorganic contaminates are heavy metals. They have some different problems than organic contaminants (Ghosh et al., 2005; Jadhav et al., 2010).

Metal pollution of the sea is less visible and direct than other types of marine pollution but its effects on marine ecosystems and humans are very extensive. The presence of metals varies between fish species; depend on age, developmental stage and other physiological factors. Additionally, fish are the largest sources of contaminants (e.g heavy metal) for man. The bioaccumulation of metals e.g Cadmium, Arsenic, Manganese, Mercury, and Lead in the trophic food chain is cause of concern since they can have deleterious effects on human health (Ersoy and Celik, 2009; Jarup and Akesson, 2009). Heavy metals normally occur in nature and are essential to life but can become toxic through accumulation in organisms.

Arsenic, cadmium, chromium, copper, nickel, lead and mercury are the most common heavy metals which can pollute the environment. Mercury, lead and cadmium are of greatest concern because of their ability to travel long distances in the atmosphere. Cadmium is also toxic to plants and microorganisms. Cd accumulates mainly in the kidney and liver of vertebrates and in aquatic invertebrates and algae. Acute toxic effects on fish, birds and other animals may include death or fetal malformations. Aquatic organisms, such as fish. accumulate pollutants directly from contaminated water and indirectly via the food chain. Heavy metals in water are particularly dangerous for fish juveniles and may considerably reduce the size of fish populations or even cause extinction of entire fish population in polluted reservoirs (Stomiñska and Jezierska, 2000). Heavy metals have the tendency to accumulate in various organs of aquatic organisms, which in turn may enter into the human metabolism through consumption causing serious health hazards (Raja et al., 2009). It's known that fish tissue accumulate large amounts of toxic contaminants from their living environment (Suhaimi et al., 2005). Sub-lethal effects of heavy metals are of concern as they accumulate and are transferred through food-chains to humans (Yılmaz and Yılmaz, 2007). Every pollution in the aquatic environment which impacts physiology, development, growth or survival of fish, affects human that, at the top of the food chain, consume fish. Cadmium as a toxic element might be a stressor agent for fish. Cadmium exposure may lead to the results of some patho-physiological damages including growth rate reduction in fish (Kaviraj and Ghosal, 1997; Hansen et al., 2002) and also in other aquatic organisms (Das and Khangarot, 2010). For this purpose, one of the aims of aquatic toxicology is to elucidate the subtler and most pronounced alterations induced by pollutants on aquatic organisms and their environment. Environmental stress is further accentuated by a concomitant increase in metal toxicity. In this way, it has been reported that acute toxicological and physiological effects to aquatic organisms following waterborne Cd exposure can be altered by some others parameters: water hardness (Davies et al., 1993), water metal concentration e.g Calcium, Zinc and Selenium (Brando-Netro et al., 1995; Meyers, 1999; Lin and Shiau, 2007; Abdel-Tawwab et al., 2007; Messaoudi et al., 2009, 2010). In this way, the antioxidant therapy is considered an important approach to intervention of heavy metal toxicity.

2. MICROALGAE: IMPLICATION IN HEAVY METAL DETOX

Land plants, aquatic plants and algae have all attracted considerable attention for the capacity to eliminate heavy metal. Indeed, some species and ecotypes of algae can live in the presence of toxic metal concentrations in the environment that are lethal for other species. Previous researchers demonstrate (Aziz and Ng, 1993; Oswald, 1995) that microalgae play an important role during the tertiary treatment of domestic wastewater in maturation ponds and/or in the treatment of smallmiddle-scale municipal wastewater in facultative or aerobic ponds. Microalgae enhance the removal of nutrients, organic contaminants, heavy metals, and pathogens from domestic wastewater and can furnish an interesting raw material e.g. algae metabolites and biogas for the production of highvalue chemicals.

Adverse effects of heavy metals on the environment and their accretion through the food chain have lead to research in the development of efficient, low cost techniques for wastewater treatment (Pan et al., 2009; Sahan et al., 2010; Singh et al., 2007), with methods using algae biomass receiving a great deal of attention (Mehta and Gaur, 2005; Singh et al., 2007; Tuzen and 2010).To their ubiquity in aquatic Sari. ecosystems, the use of algae to monitor heavy metal toxicity is increasing due to their involvement in the most aquatic processes.

For example, *Spirulina* and *chlorella* are two separate micro-algae organisms which have existed on earth since the dawn of time. Both were revered as powerful super foods in many traditional societies, and today are more relevant than ever for achieving overall health and wellbeing. Many studies have reported that *Spirogyra* and *Cladophora* spp. have a very high capacity for binding with metals due to the presence of polysaccharides, proteins, or lipids on the surface of cell walls (Alimohamadi *et al.*, 2005; Deng *et al.*, 2007; Gupta and Rastogi, 2008; Tuzen and Sari, 2010).

McLean et al. (1972) first reported the presence of Cd-binding material in a freshwater blue green algae, Anacyslis nidulans, but no attempt was made further characterization. Indeed, metalbinding proteins have been reported for different algae, i.e. Chlorella ellipsoida (Nagano et al., 1982), C. pyrinoidosa (Hart and Bertram, 1980), Dunaliella bioculata (Heuillet et al., 1988), Synechococcus sp. (Olafson et al., 1979), Euglena gracilis (Gingrich et al., 1986), Scenedesmus acutiformis (Stokes et al., 1977) and S. quadricauda (Reddy and Prasad, 1989). All such proteins are within a mol. wt range of 8000-10,000. Photosynthetic aeration is therefore especially interesting to reduce operation costs and limit the risks for pollutant volatilization under mechanical aeration and recent studies have shown that microalgae can indeed support the degradation of various aerobic hazardous

contaminants (Munoz *et al.*, 2004; Safonova *et al.*, 2004).

It has been reported that microalgae are aquatic organisms presenting molecular mechanisms that allow them to discriminate non-essential heavy metals from those essential ones for their growth. Stokes *et al.* (1977) first discovered MtIII complex synthesis in the microalga *Scenedesmus acutiformis.* The different detoxification processes executed by algae are reviewed with special emphasis on those involving the peptides metallothioneins, mainly the post transcriptionally synthesized class III metallothioneins or phytochelatins (Gekeler *et al.*, 1988).

Heavy metals represent an important group of polluted and hazardous contaminants often found in industrial wastewater (Kratochvil and Volesky, 1998; Volesky, 2001). Microalgae, related eukaryotic photosynthetic organisms, and some fungi have preferentially developed the production of peptides capable to bind heavy metals. These molecules, as organometallic complexes, are further partitioned inside vacuoles to facilitate appropriate control of the cytoplasmic concentration of heavy metal ions, thus preventing or neutralizing their potential toxic effect (Cobbett and Goldsbrough, 2002). In contrast to this mechanism used by eukaryotes, prokaryotic cells employ ATP consuming efflux of heavy metals or enzymatic change of speciation to achieve detoxification (Nies, 1999).

For example, Cd is believed to have no nutritive value for algae. Canizares-Villanueva (2000) have reported that the microalgae can be efficiently use to remove heavy metals and a fixperinetal uptake of 15mg $g^{-1}_{Biomass}$ at 99% removal efficiency has been reported. This data showed that the process is competitive compared to other treatment methods. Stokes *et al.* (1977) have discovered MtIII complex synthesis in the microalga *Scenedesmus acutiformis*.

3. VITAMINS

Dietary micronutrients such as vitamins E and C as well as carotenoids have also been regarded as antioxidant defenses e.g in fish (Wilhelm-Filho, 1996; Hamre, 2010). Vitamin C (ascorbic acid) and vitamin E (d-alpha-tocopherol) are recognized as essential nutrients for all species of animals. In other words, these vitamins have been shown to have protective effect against metal induced toxicity (Rao and Sharma, 2001; Jiraungkoorskul *et al.* 2007). Vitamin C is an electron donor, and this property accounts for all its known functions. Antioxidant effects of vitamin C have been demonstrated in many experiments in vitro. Vitamin C is an electron donor and therefore a reducing agent. All known physiological and biochemical actions of vitamin C are due to its action as an electron donor. For this purpose, Vitamin C is called an antioxidant because, by donating its electrons, it prevents other compounds from being oxidized (Padayatty et al., 2003). Alpha-tocopherol (vitamin E) is a naturally occurring antioxidant in biological systems and is present in the cell membrane of various tissues, including the intestine and stomach. Vitamin E prevents free radical-induced injury by blocking the free radical chain reaction (Solar, 1959).

Layachi and Kechrid (2012), have reported that the supplementation of Vit-C and Vit- E or there combination in rat exposed to Cd significantly increase of the glutathione level in comparison to groups exposed cadmium. Additionally, the normalization of glutathione levels (GSH), glutathione peroxidase (GSH-Px) and catalase (CAT) activities following vitamin C or vitamin E treatment could be explained by the fact that these vitamins caused a decline in lipid peroxidation (LPO) accompanied by an increase in the activities/level of, GSH, GSH-Px and CAT in liver rat. In addition, it has been reported that the body weights decrease significantly while the liver weight of rats exposed to Cd significantly increase as compared to the control group. However, in the group with vitamin C, vitamin E and vitamin C + vitamin E supplies, the body weight gain became significantly greater and the liver weight decreased than in rats exposed to cadmium (Layachi and Kechrid, 2012). The same authors reported that, the activities of serum hepato specific enzymes serum alanine aminotransaminase (GOT), glutamate-pyruvate transaminase (GPT) and alkaline phosphatase (ALP) were generally significantly decreased in rat groups treated with vitamin C and vitamin E either alone or in combination. Moreover, the combination of vitamin C and E showed more efficacy than vitamin C or vitamin E alone when comparing to group Cd-vit C and Cd-vit E with Cd-vit C-vit E animals. Erdogan et al., 2005 reported that the reduction in weight gain could have been due to the decrease in food intake, or could be related to the overall increased of lipids and proteins degeneration as a result of cadmium Amelioration blood toxicity. of glucose concentration in cadmium animals treated was observed after addition of vitamin E or vitamin C

alone or in combination (Layachi and Kechrid, 2012). Cadmium is one of the heavy metals which induce membrane damage. Cadmium toxicity was attenuated by the pretreatment with vitamine E even at the highest concentrations tested (Mattie and Freedman, 2001). The co-administration of vitamin C and/or vitamin E to the cadmium treated animals improved body and liver weights. The protective effect of vitamine-C (ascorbic acid) on heavy metal toxicity via its free radical scavenging mechanism and detoxification effect is well reported (Suzuki, 1990). However, according to Sodhi et al., (2008), vitamin E due to its solubility in lipids, plays an important role in protecting lipid-rich structure like hepatic tissue from free radicals damage and an effective inhibitor of autocatalytic process of lipid peroxidation (LPO).

4. MINERAL SUPPLEMENTATION

One of the major mechanisms behind metal toxicity has been attributed to oxidative stress. The biological activity of heavy metals can be markedly affected by the presence of metal chelators which may reverse their toxicity. Much interest has recently been focused on the uptake of cadmium by marine and estuarine organisms. Many of these studies have been prompted by a concern over public health hazards caused by accumulation of this trace metal in food.

Several studies indicated that calcium (Ca2+) can reduce heavy metal toxicity by forming complex compounds with them which are then, either eliminated or unable to cross biological membranes. The mechanism of acute Cd toxicity appears to be interference with whole-body calcium regulation resulting in hypocalcaemia, but this interference only occurs at industrial-type concentrations. Furthermore, during such exposures, Cd accumulates in the whole body of the organism to levels (>50 μ mol g⁻¹) as much as 5-fold greater than normal whole-body Ca concentration (10–15 μ mol g⁻¹). This raises interesting questions about the interaction of Ca and Cd uptake in this animal.

It's well known that Ca plays diverse role in the living organisms. Indeed, in the vertebrate's organisms it represents a major component of the skeleton but in addition, it also has vital functions in the body fluids and soft tissues. For enzymatic processes, Ca acts as a cofactor in various and couples stimulus excitation reactions (e.g in muscle contraction or the secretion of glands). Both Ca and Cd are divalents and they use the same transitional channel interacting with each other antagonistically. For example, it have been reported that high concentrations of Ca either in water or diet clearly envisages ameliorating protective effects on water borne Cd toxicity in fish *Oreochromis mossambicus* (Zohouri *et al.*, 2001).

In teleostean fish *Oreochromis mossambicus*, Cd induces significant alterations in the levels of lipid peroxidation (LPO) and certain enzymatic status of antioxidant enzymes (superoxide dismutase and catalase) in liver and kidney tissues. However, these activities were progressively reversed after using trace element supplements like Ca and/or Zn (Jamakala and Rani, 2012).

Ca and/or Zn supplementation significantly counteracted the enhancement of LPO caused by Cd exposure. This finding is in a perfect agreement with the findings of Ng *et al.*, (2009) who reported that elevated Ca protects against Cd induced toxicity in rainbow trout. It has been reported that a lower level of LPO means a lower degree of membrane damage, and Ca and Zn might have alleviated the Cd- induced membrane damage and aids protect the cell (Jamakala and Rani, 2012).

Therefore, the enhanced LPO in the liver, kidney and other tissues might result from the reduction of their SOD activity. The decrease in SOD activity could be due to its inhibition by the excess production of ROS as evidenced by LPO following Cd exposure. The supplementation of Ca and/or Zn if fish (Oreochromis Mossambicus) exposed to Cd causes a significant increase in SOD activity in both liver and kidney tissue (Jamakala and Rani, 2012). Similar findings were reported rainbow trout (Baldisserotto et al., 2004) and rats (Patra et al., 2001) exposed to Cd and subjected to Ca and Zn supplementation. Several study reported that Cd exposure decrease CAT activity in Oreochromis Mossambicus, Heteropneustes fossilis and Sparus aurata (Jamakala and Rani, 2012; Radhakrishnan et al., 1999; Vaglio and Landriscina, 2009). However, after supplementation of Ca and/or Zn, CAT activity levels were significantly increased especially in liver and kidney tissue in fish.

Several elements have been shown to have a protective effect against Cd-induced injury. Accordingly, it was reported that selenium (Se) is considered one of the most efficient and an important nutritional trace element (Shilo *et al.*, 2010), which contributes significantly to host

IJPBA, Jan - Feb, 2013, Vol. 4, Issue, 1

immune responses and antioxidant protection (Brigelius-Flohe and Flohe, 2003). Among the other antioxidants, selenium (Se) is an essential trace mineral in animal nutrition obtained partly from the surrounding water (Lall and Bishop, 1977), but mostly from the diet (Halver, 2002). The importance of Se to oxidative stress involves its presence at the active site of the antioxidant enzyme GPX (Felton et al., 1996), which expense of reduced $reducesH_2O_2$ at the glutathione (Arteel and Sies, 2001). Se represents one of the important nonenzymatic antioxidant defense systems, which may modulate its toxicity by an antioxidative mechanism. Se acts as a of unusual component the amino acids selenocysteine (Se-Cys) and selenomethionine (Se-Met).

Many proteins contain selenium, among them glutathione peroxidase (GSH-Px), plasma cytoplasm GSH-Px, mitochondrial GSH-Px, cell membrane phospholipid hydroperoxide glutathione peroxidase (PHGSH-Px) and iodothyronine deiodinase (Sies, 1991). Glutathione peroxidase (PHGSH-Px), a selenoenzyme, reduces in situ the oxidized phospholipid polyunsaturated fatty acids, restoring cellular membranes to normal (Bock et al., 1991). Combs et al., (2009) reported that Se acts as an antioxidant in the body as an antioxidant, and is also involved in thyroid hormone metabolism, redox reactions, reproduction, and immune function. Se plays a role in protecting cells against free radicals and oxidative stress (Bansal and Kaur, 2005). Additionally, numerous studies have shown that Se can protect against Cd toxicity in mammals in vitro and in vivo (El-Sharaky et al., 2007, Lazarus et al., 2009; Messaoudi et al., 2009, 2010). In addition, Se acts as a co-factor for the reduction of antioxidant enzymes, including glutathione peroxidases and certain forms of thioredoxin reductase (Chen et al., 2012). Moreover, the protective effect of Se against the toxicity of Cd might, at least partially, be attributed to stimulate the level of heat shock protein (HSPs). Therefore, Se can be considered a potential therapeutic nutrient to protect against toxicity induced by Cd.

REFERENCES

1. Abdel-Tawwab M, Mousa MAA, Abbass, FE. Growth performance and physiological response of African catfish, *Clarias gariepinus* (B.) fed organic selenium prior to the exposure to environmental copper toxicity. Aquaculture 2007; 272(1-4): 335–345.

- Alimohamadi M, Abolhamd G, Keshtkar A. Pb(II) and Cu(II) biosorption on *Rhizopus arrhizus* modeling mono- and multi-component systems. Miner Eng 2005; 18: 1325–1330.
- 3. Arteel GE, Sies H. The biochemistry of selenium and the glutathione system. Environ Toxicol Phar 2001; 10: 153–158.
- 4. Athar M, Vohora SB. Heavy metals and environment, New Delhi, New Age International Publisher, 2001, p. 3–40.
- Aziz MA, Ng WJ. Industrial wastewater treatment using an activated algae-reactor. Water Sci Technol 1993; 28: 71–76.
- B6ck A, Forchhammer K, Heider J, Baron C. Selenoprotein synthesis, an expansion of the genetic code. Trends Biochem Sci 1991; 16: 463-7.
- Baldisserotto B, Kamunde C, Matsuo A, Wood CM. A protective effect of dietary calcium against acute waterbone cadmium uptake in rainbow trout. Aquat Toxicol 2004; 67: 57-73.
- 8. Bansal MP, Kaur P. Selenium, a versatile trace element: Current research implications. Indian J Exp Biol 2005; 43: 1119–1129.
- Brando-Neto J, Mduriera G, Mendonca BB, Bloise W, Castro AV. Endocrine interaction between and zinc and prolactin. An interpretive review. Biol Trace Elem Res 1995; 49: 139–149.
- Brigelius-Flohe R, Flohe L. Is there a role of glutathione peroxidases in signaling and differentiation. Biofactors 2003; 17: 93– 102.
- 11. Canizares-Villanueva RO. Heavy metals biosorption by using microbial biomasa. Rev Latinoam Microbiol 2000; 131–143 (in Spanish).
- 12. Chen X, Zhu YH, Cheng XY, Zhang ZW, Xu SW. The protection of selenium against cadmiu minduced cytotoxicity via the heat shokpro tein pathway in chickensplenic lymphocyt es. Molecules. 2012; 17(12):14565-14572.
- 13. Cobbett C, Goldsbrough P. Phytochelatin and metallothioneins: Roles in heavy metal detoxification and homeostasis. Annu Rev Plant Biol 2002; 53: 159–182.
- 14. Combs GFJr, Midthune DN, Patterson KY, Canfield WK, Hill AD, Levander,

OA, Taylor PR, Moler JE, Patterson BH. Effects of selenomethionine supplementation on selenium status and thyroid hormone concentrations in healthy adults. Am J Clin Nutr 2009; 89: 1808– 1814.

- 15. Das S, Khangarot BS. Bioaccumulation and toxic effects of cadmium on feeding and growth of an Indian pond snail *Lymnaea luteola* L. under laboratory conditions. J Hazard Mat 2010; 15: 763– 70.
- 16. Davies PH, Gorman WC, Carlson CA, Brinkman SR. Effect of hardness on bioavalibility and toxicity of cadmium to rainbow trout. Chem Speciat Bioavailab 1993; 5: 67–77
- 17. Deng LP, Su YY, Su H, Wang XT, Zhu XB. Sorption and desorption of lead(II) from wastewater by green algae *Cladophora fascicularis*. J Hazard Mater 2007; 143: 220–225.
- El-Sharaky AS, Newairy AA, Badreldeen MM, Eweda SM, Sheweita SA. Protective role of selenium against renal toxicity induced by cadmium in rats. Toxicology 2007; 235: 185–193.
- 19. Erdogan Z, Erdogan S, Celik S, Unlu V. Effects of ascorbic acid on cadmiuminduced oxidative stress and performance of broilers. Biol Trace Elem Res 2005; 104:19-31.
- 20. Ersoy B, Celik M. Essential elements and contaminants in tissues of commercial pelagic fish from the Eastern Mediterranean Sea. J Sci Food Agric 2009; 89(9):1615–1621.
- 21. Felton SP, Landolt ML and Grace R. Effects of selenium dietary enhancement hatchery-reared on coho salmon, Oncorhynchus kisutch (Walbaum), when compared with wild coho: hepatic enzymes and adaptation seawater evaluated. Aquacult Res 1996; 27: 135-142.
- 22. Gekeler W, Grill E, Winnacker EL, Zenk MH. Algae sequester heavy metals via synthesis of phytochelatin complexes. Arch Microbiol 1988; 150: 197–202.
- 23. Ghosh M, Singh SP. Review on phytoremediation of heavy metals and utilization of its byproducts. Appl Ecol Env Res 2005; 3 (1): P. 1–18.

- 24. Gingrich DJ, Weber DN, Shaw CF, Garvevj S, Petering DH. Characterization of a highly negative and labile binding protein induced in *Euglena gracilis* by cadmium. Envir Hlth Persp 1986; 65: 77 85.
- 26. Gupta VK, Rastogi A. Biosorption of lead from aqueous solutions by green algae *Spirogyra* species: kinetics and equilibrium studies. J Hazard Mater 2008; 152: 407–414.
- 27. Halver JE. The vitamins. In Fish Nutrition, 2nd ed., pp. 62–132 [JE Halver and R Hardy, editors]. New York: Academic Press: 2002
- 28. Hamre K. Metabolism, interactions, requirements and functions of vitamin E in fish. Aquacult Nutr 2010; 17: 98–115.
- 29. Hansen JA, Welsh PG, Lipton J, Suedkamp MJ. The effects of long-term cadmium exposure on the growth and survival of juvenile bull trout (*salvelinus confleuentus*). Aquat Toxicol 2002; 58: 165–174.
- 30. Hart BA, Bertram PE. Cadmium binding protein in a cadmium tolerant strain of *Chlorella pyrenoidosa*. Envir Exp But 1980; 20: 175-180.
- Heullet E, Guerbette F, Guenou C, Kadar JC. Induction of a cadmium binding protein in a unicellular alga. Int J Biochem 1988; 20: 203 210.
- 32. Jadhav JP, Kalyani DC, Telke AA, Phugare SS, Govindwar SP. Evaluation of the efficacy of a bacterial consortium for the removal of color, reduction of heavy metals, and toxicity from textile dye effluent. Bioresource Technol 2010; 101: 165–173.
- 33. Jamakala O, Rani UA. Protective role of trace elements against cadmium induced alterations in the selected oxidative stress enzymes in liver and kidney of fresh water teleost, oreochromis mossambicus (tilapia). Int J Pharm Pharm Sci 2012;4(5).
- 34. Jarup L, Akesson A. Current status of cadmium as an environmental health problem. Toxicol Appl Pharmacol 2009; 238(3):201–208.
- 35. Jiraungkoorskul W, Sahaphong S, Kosai P, Kim MH. The effect of ascorbic acid on cadmium exposure in the gills of Puntius altus. Int J ZoolRes 2007; 3:77-85.

- 36. Kaviraj A, Ghosal, TK. (Effects of poultry litter on the chronic toxicity of cadmium to common carp (*Cyprinus carpio*). Bioresource Technol 1997; 60: 239–243.
- Kratochvil D, Volesky B. Advances in the biosorption of heavy metals. Trends Biotechnol 1998; 16: 291–300.
- 38. Lall SP., Bishop FJ. Studies on mineral and protein utilization by Atlantic salmon (*Salmo salar*) grown in sea water. Fisheries and Marine Service, Environment Canada, Ottawa, ON, Technical Report No. 688, pp. 16:1977.
- 39. Layachi N, Kechrid Z. Combined protective effect of vitamins C and E on cadmium induced oxidative liver injury in rats. Afr J Biotechnol 2012; 11: 16013-16020.
- 40. Lazarus M, Orct T, Jurasoviae J, Blanusa M. The effect of dietary selenium supplementation on cadmium absorption and retention in suckling rats. Biometals 2009; 22: 973–983.
- 41. Lin YH, Shiau SY. The effects of dietary selenium on the oxidative stress of grouper, *Epinephelus malabaricus*, fed high copper. Aquaculture 2007; 267: 38–43.
- 42. Maclean FI, Lucis OJ, Shaikh ZA, Jansz ER. The uptake and subcellular distribution of Cd and Zn in microorganisms. Proc Fedn Am Soc Exp Biol 1972; 31:699.
- 43. Mattie MD, Freedman JH. Protective effects of aspirin and vitamin E (alphatocopherol) against copper- and cadmiuminduced toxicity. Biochem Biophys Res Commun 2001; 285(4):921-5. 44. Mehta SK, Gaur JP, Use of algae for removing heavy metal ions from wastewater: progress and prospects. Crit Rev Biotechnol 2005; 25: 113–152.
- 44. Messaoudi I, El Heni J, Hammouda F, Saïd K, Kerkeni A. Protective effects of selenium, zinc, or their combination on cadmium-induced oxidative stress in rat kidney. Biol Trace Elem Res 2009; 130(2):152-61
- 45. Messaoudi I, Hammouda F, El Heni J, Baati T, Saïd K, and Kerkeni A. Reversal of cadmium-induced oxidative stress in rat erythrocytes by selenium, zinc or their combination. Exp Toxicol Pathol 2010; 62:281–288.

- 46. Meyers JS. A mechanistic explanation for the ln(LC50) vs ln (hardness) adjustment equation for metals. Environ Sci Technol 1999; 33: 908–912
- 47. Munoz R, Kollner C, Guieysse B, Mattiasson B. Photosynthetically oxygenated salicylate biodegradation in a continuous stirred tank photobioreactor. Biotechnol Bioeng 2004; 87: 797–803.
- 48. Nagano T, Watanase Y, Hida K., Suketa Y, Okada S. Property of cadmium-binding protein in *Chlorella ellipsoida*. Eiseikagaku 1982; 28: 114-117.
- 49. Ng TYT, Klink JS, Wood CM. Does dietary Ca protect against toxicity of a low diet borne Cd exposure to the rainbow trout? Aquat Toxicol 2009; 91: 75-86.
- 50. Nies DH. Microbial heavy-metal resistance. Appl Microbiol Biotechnol 1999; 51: 730–750.
- 51. Olafson RW, Abel K, Sim RG. Prokaryotic metallothionein: preliminary characterization of a blue-green alga heavy metal binding protein. Biochem biophys Res Commun 1979; 89: 36 43.
- 52. Oswald WJ. Ponds in the twenty-first century. Water Sci Technol 1995; 31: 1–8.
- 53. Padayatty SJ, Katz A, Wang Y, Eck P, Kwon O, Lee JH, Chen S, Corpe C, Dutta A, Dutta SK, Levine M. Vitamin C as an antioxidant: evaluation of its role in disease prevention. J Am Coll Nutr 2003; 22(1):18-35.
- 54. Pan R, Cao L, Zhang, R. Combined effects of Cu, Cd, Pb, and Zn on the growth and uptake of consortium of Cu-resistant Penicillium sp. A1 and Cd-resistant Fusarium sp. A19. J Hazard Mater 2009; 171: 761–766.
- 55. Patra RC, Swarup D, Dwivedi SK. Antioxidant effects of alpha tocopherol, ascorbic acid and L-methionine on lead induced oxidative stress to the liver, kidney and brain in rats. Toxicology 2001; 162: 81 - 88.
- 56. Perales-Vela HV, Pena-Castro JM, Canizares-Villanueva RO. Heavy metal detoxification in eukaryotic microalgae. Chemosphere 2006; 64: 1–10.
- 57. Price NM, Morel FMM. Cadmium and cobalt substitution for zinc in a marine diatom. Nature 1990; 344: 658–660.
- 58. Radhakrishnan MV. Effect of cadmium on catalase activity in four tissues of fresh

water fish *Heteropneustes fossilis* (Bloch). The Internet Journal of Veterinary Medicine 2009; ISSN: 1937-8165.

- 59. Raja P, Veerasingam S, Suresh G, Marichamy, G, Venkatachalapathy R. Heavy metals concentration in our commercially valuable marine edible fish species from Parangipettai Coast, South East Coast of India. Int J Anim Vet Adv 2009; 1(1):10–14.
- 60. Rao MV, Sharma PS. Protective effect of vitamin E against mercuric chloride reproductive toxicity in male mice. Reprod Toxicol 2001; 15:705-712.
- 61. Reddy GN, Prasad MNV. Cadmium inducible proteins in *Scenedesmus quadricauda*. Curr Sci 1989; 58: 1381-1382.
- 62. Safonova E, Kvitko KV, Iankevitch MI, Surgko LF, Afri IA, Reisser W. Biotreatment of industrial wastewater by selected algal–bacterial consortia. Eng Life Sci 2004; 4: 347–353.
- 63. Sahan T, Ceylan H, Sahiner N, Aktas N. Optimization of removal conditions of copper ions from aqueous solutions by *Trametes versicolor*. Bioresour Technol 2010; 101: 4520–4526.
- 64. Shilo S, Pardo M, Aharoni-Simon M, Glibter S, Tirosh O. Selenium supplementation increases liver MnSOD expression: Molecular mechanism for hepato-protection. J Inorg Biochem 2008; 102: 110–118.
- 65. Sies H. Oxidative Stress. pp. 327-9, 541-55: 1991.
- 66. Singh A, Kumar D, Gaur JP. Copper (II) and lead(II) sorption from aqueous solution by non-living *Spirogyra neglecta*. Bioresour Technol 2007; 98: 3622–3629.
- 67. Sodhi S, Shamara A, Brar APS, Brar RS. Effect of tocopherol and selenium on antioxidant status, lipid peroxidation and hepatotoxicty induced by malathion in chicks. Biochem Physiol 2008; 90:82-86.
- 68. Solar CJ. Vitamin E and ulcers. Digest Nutr 1959; 18: 745
- 69. Stokes PM, Maler T, Riordan JR. A low molecular weight copper-binding protein in a copper tolerant strain of Scenedesmus

aculiformis. Pages 146-154 in D. D. HEMPHIL, ed. Trace substances in environmental health, Vol. X1. University of Missouri Press, Columbia, MO: 1977.

- 70. Stokes PM, Maler T, Riordan JR. A low molecular weight copper-binding protein in a copper tolerant strain *Scenedesmus acutiformis*. In: Hemphil, D.D. (Ed.), Trace Substances in Environmental Health. University of Missouri Press, Columbia, pp. 146–154: 1977.
- 71. Stomiňska I, Jezierska B. The effect of Heavy metals on post embryonic development of common carp, *Cyprinus carpio* L. Arch Pol Fish 2000; 8: 119–128.
- 72. Suhaimi F, Wong SP, Lee VLL, Low LK. Heavy metals in fish and shellfish found in local wet markets. Singapore J Primary Ind 2005; 32:1–18
- 73. Suzuki Y. Synergism of ascorbic acid and glutathione in reduction of hexavalent chromium *in vitro*. Ind Health 1990; 28:9-19.
- 74. Tuzen M, Sari A. Biosorption of selenium from aqueous solution by green algae (*Cladophora hutchinsiae*) biomass: equilibrium, thermodynamic and kinetic studies. Chem Eng J 2010; 158: 200–206.
- 75. Vaglio A, Landriscina C. Changes in liver enzyme activity in the teleost *Sparus aurata* in response to cadmium intoxication. Ecotoxicol Environ Saf 1999; 43: 111-116.
- 76. Volesky B. Defixation of metal bearing effluents: biosorption for the next century. Hydrometallurgy. 2001; 59: 203–216.
- 77. Wilhelm-Filho D. Fish antioxidant defenses a comparative approach. Braz J Med Biol Res 1996; 29; 1735–1742.
- 78. Yılmaz, A.B., Yılmaz, L. (2007). Influences of sex and seasons on levels of heavy metals in tissues of green tiger shrimp (*Penaeus semisulcatus* de Hann, 1844). Food Chem. 101:1664–1669
- Zohouri MA, Pyle GG, Wood CM. Dietary Ca inhibits waterborne Cd uptake in Cdexposed rainbow trout, *Oncorhynchus mykiss*. Comp Biochem Physiol 2001; 130C (2/3): 347-356.